

The Free Internet Journal for Organic Chemistry

Pnaphthaaper

Archive for Organic Chemistry

Arkivoc **2021**, part iii, 0-0 to be inserted by editorial office

Visible light-promoted Selectfluor-mediated quinone functionalization with unactivated Csp³-H components

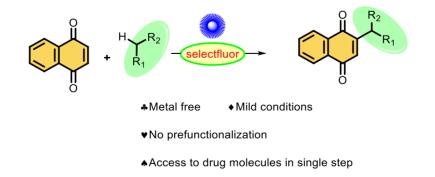
Arun D. Kulthe,^{a,b†} Maheshwara Reddy Nadiveedhi,^{c†} Prathama S. Mainkar ^{*a,b} and Srirama Murthy Akondi ^{*a,b}

^aDepartment of Organic Synthesis and Process Chemistry, CSIR-Indian Institute of Chemical Technology (CSIR-IICT), Hyderabad-500007, India. ^bAcademy of scientific and innovative research (AcSIR), Ghaziabad 201002, India. ^cDepartment of Chemistry, Sri Venkateswara University, Tirupati-517502, India. [†]These authors contributed equally to this work Email: sriramakondi@iict.res.in; sriramiict@gmail.com

Dedicated to Professor Lanny Liebeskind on the occasion of his 70th anniversary

Received mm-dd-yyyy	Accepted mm-dd-yyyy	Published on line mm-dd-yyyy
Dates to be inserted by editorial office		
Abstract		

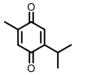
A visible light-promoted metal-free cross-dehydrogenative-coupling (CDC) method for the alkylation of 1,4naphthoquinones is reported using Selectfluor as a hydrogen atom transfer (HAT) agent. This protocol is suitable for a variety of 1,4-naphthoquinones and Csp³-H compounds and it facilitates the formation of pharmaceutically important quinone scaffolds under mild conditions. Using this methodology, the antimalarial drug, parvaquone, was synthesized in a single step.

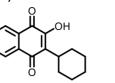


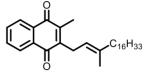
Keywords: Quinone functionalization, cross-dehydrogenative-coupling, Selectfluor and visible light

Introduction

The 1,4-quinone scaffold is one of the most investigated templates in the field of organic synthesis, as it features in many functional molecules, bioactive natural products and pharmaceuticals.¹⁻⁵ Among the derivatives of 1,4-quinone, alkylated 1,4-naphthoquinones and 1,4-benzoquinones (Figure 1) are extremely important as they exhibit a wide range of biological properties such as anti-microbial, anti-tumor, anti-inflammatory and neurological activities (Figure 1).⁶⁻¹⁰

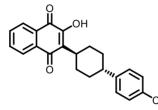


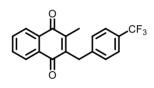




Thymoquinone (anticancer)

Parvaquone (antiparasitic) Vitamin K1 (coagulation)





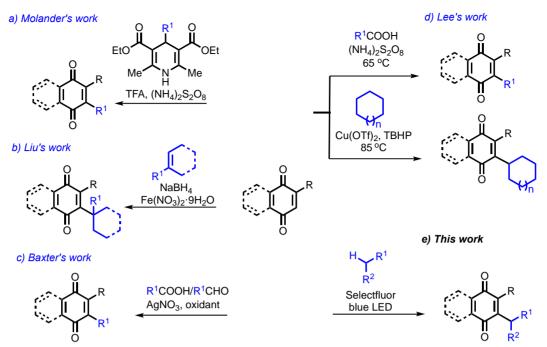
Atovaquone (antiparasite)

Plasmodione (antimalarial lead drug)

Figure 1. Some biologically active alkylated quinones

Owing to the importance of alkylated 1,4-quinone frameworks, several radical alkylation protocols for the functionalization of 1,4-quinones have been proposed in recent years (Scheme 1). For instance, Molander and co-workers reported alkylation of 1,4-quinones *via* oxidative homolysis of 1,4-dihydropyridines (Scheme 1a).¹¹ Liu's group developed an Fe(III) mediated radical alkylation of quinones with olefins (Scheme 1b).¹² Baxter and co-workers reported a silver-catalysed radical alkylation of quinones (Scheme 1c).¹³ Also, a persulfate mediated alkylation of 1,4-quinones using carboxylic acids¹⁴ and a copper catalysed direct cross-coupling of 1,4-quinones with cycloalkanes¹⁵ were demonstrated by Lee's group (Scheme 1d). Nevertheless, each of these methods has limitations such as pre-functionalization of the alkyl partner, requirement of transition metal reagents and explosive peroxides as oxidants. Hence, more efficient, milder and sustainable methods are still desirable.

Visible light-promoted Selectfluor-mediated direct functionalization of C-H bonds via the hydrogen atom transfer (HAT) mechanism has attracted attention in recent years. Pioneering work by Lei's group¹⁶⁻¹⁷ demonstrated the Csp³-H heteroarylation of cycloalkanes, ethers and alcohols mediated by Selectfluor under visible-light irradiation. At the same time, Jin's group¹⁸ used similar reaction conditions for the alkylation of heteroarenes via the generation of alkyl radicals from unactivated Csp³-H components. However, a visible light-promoted, Selectfluor-mediated C-H functionalization for the alkylation of quinones has not yet been studied.



Scheme 1. Radical alkylative functionalization of quinones

Herein, we report a new method of introducing alkyl groups into the 1,4-naphthoquinone skeleton without pre-functionalization of the alkyl partner (Scheme 1d). This method utilizes visible light and the green oxidant Selectfluor without the requirement for radical initiator or transition metal photocatalyst.

Results and Discussion

With an overall aim of developing a direct Csp^2-Csp^3 cross coupling strategy for the alkylation of quinones under mild conditions, we started our investigation by irradiating a mixture of 2-acetoxy-1,4-naphthoquinone (1a), cyclohexane (2a) and Selectfluor in acetonitrile under a N₂ atmosphere at room temperature using a Penn PhD photoreactor M2 (blue LED, 450 nm) (Table 1, entry 1).

Table 1. Screening of optimal reaction conditions^a



entry	variation from above conditions	yield (%) ^b
1	none	42
2	white LED instead of blue LED	trace
3	80 °C in the dark	trace
4	2 eq CH ₃ COOH as additive	43
5	2 eq CF ₃ COOH as additive	39
6	acetone as a solvent	28

7	cyclohexane as a solvent	nd
8	2 eq Et₃N as additive	nd
10	1a in CH ₃ CN was added over 20 min through syringe pump	54

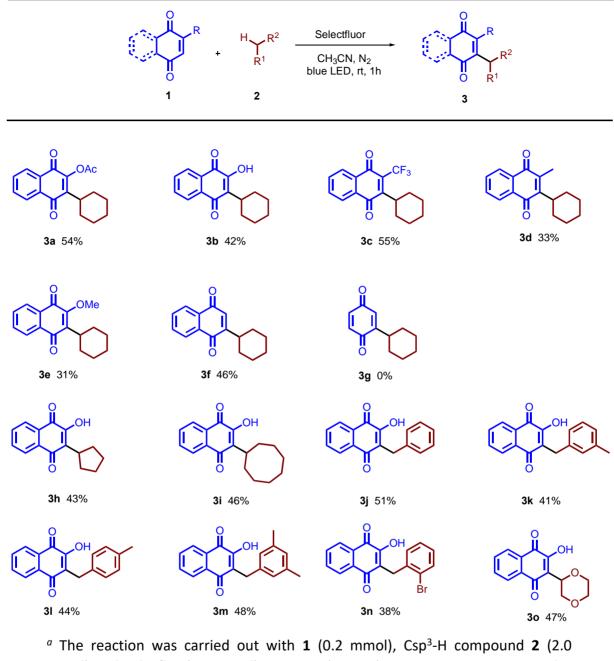
^{*a*} The reaction was carried out with **1a** (0.2 mmol), cyclohexane (2.0 mmol) and Selecfluor (0.4 mmol) in MeCN (2.0 mL) at room temperature under N₂ using Penn PhD photoreactor M2 (blue LED 450 nm) for 1 h. ^{*b*} Isolated yields.

To our delight, the desired cross coupled product **3a** was formed in 42% yield within one hour. No desired product was observed when the light source was changed to 14 W white LED (Table 1, entry 2). Also, heating the reaction mixture to 80 °C in the absence of light, also did not yield any of the desired product (Table 1, entry 3). Use of additives such as AcOH and TFA didn't improve the yield of **3a** (Table 1, entries 4 and 5) and Et₃N completely diminished the yield of **3a** (Table 1, entry 8). Use of the solvent acetone led to lower yield of **3a** (Table 1, entry 6) and reaction didn't take place when cyclohexane was used as a solvent (Table 1, entry 7). Finally, when quinone **1a** was added to a mixture of cyclohexane and Selectfluor over a period of 20 minutes through syringe pump, the isolated yield of **3a** was improved to 54% (Table 1, entry 10).

The scope and generality of this cross-coupling strategy was then investigated using various 1,4-quinones and aliphatic C-H components under the optimized conditions (Table 2). The reaction of 2-hydroxy-1,4-naphthoquinone with cyclohexane under the optimized conditions afforded the antimalarial drug molecule parvaquone (**3b**) in 42% yield. 1,4-Naphthoquinone bearing a CF₃ group at the 2-position reacted smoothly and furnished the desired product (**3c**) in 55% yield. Whereas, 2-methyl- and 2-methoxy-1,4-napthoquinones afforded the desired products (**3d** and **3e**) in lower yields. Unsubstituted 1,4-naphthoquinone gave the desired product **3f** in 46% yield. *p*-Benzoquinone did not afford the desired product and its complete decomposition was observed under the reaction conditions.

Next, the scope of different aliphatic C-H components was examined using 2-hydroxy-1,4-naphthoquinone as reaction partner. The reaction with cycloalkanes such as cyclopentane and cyclooctane was successful and furnished the desired alkylated quinones (**3h** and **3i**) in moderate yields. Later, a series of methylbenzenes were investigated. Toluene, *m*-xylene, *p*-xylene and mesitylene were all found to be suitable substrates for this methodology and furnished the desired compounds (**3j**-**3m**). Notably, *ortho*-bromotoluene reacted well without loss of Br giving **3n**, thus affording the potential for further cross-couplings. In addition, cyclic ether 1,4-dioxane was also a competent substrate, giving the desired product (**3o**) in 52% yield.

Table 2. Substrate scope of 1,4-quinones and aliphatic C-H components.



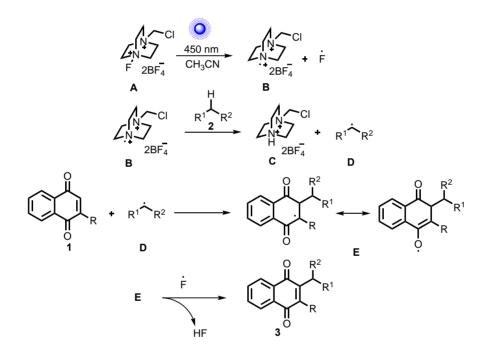
mmol) and Selecflor (0.4 mmol) in MeCN (2.0 mL) at room temperature under N_2 using Penn PhD photoreactor M2 (blue LED 450 nm) for 1 h. ^b Isolated yields.

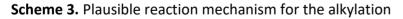
To gain mechanistic insight, we next performed radical inhibition experiments (Scheme 2). When the reaction of 2-hydroxy-1,4-naphthoquinone with toluene was conducted in the presence of three equivalents of TEMPO and butylated hydroxytoluene (BHT) (2,6-bis(1,1-dimethylethyl)-4-methylphenol) under the standard conditions, formation of the product **3j** was completely suppressed. These experiments clearly indicate a radical reaction pathway.



Scheme 2. Radical inhibition experiments

A plausible mechanism is proposed based on the control experiments above and preceding literature.¹⁵⁻¹⁷ Firstly, the N-radical cation **B** formed by the homolytic fission of Selectfluor **A** abstracts a hydrogen atom from the alkane **2** to give the alkyl radical intermediate **D** and dication **C**. The generated nucleophilic alkyl radical **D** adds to the electron-deficient quinone **1** to give the radical intermediate **E**. Finally, abstraction of a hydrogen atom by fluorine radical from intermediate **E** generates the product **3** (Scheme 3).





Conclusions

In conclusion, we have developed a visible-light induced, Selectfluor mediated Csp²-Csp³ cross coupling strategy without the need of prefunctionalization of Csp³ partner for the alkylation of 1,4-naphthoquinones. This mild protocol is suitable for various 1,4-naphthoquinones and aliphatic Csp³-H components to afford pharmaceutically important quinone frameworks in moderate yields.

Experimental Section

General. Unless otherwise noted, reagents obtained from commercial suppliers were used without further purification. Reactions were monitored using thin-layer chromatography (SiO₂). Thin layer chromatography was performed on Merck silica gel plates and visualized by UV light. Column chromatography was carried out using silica gel (100-200 mesh) packed in glass columns. NMR spectra were recorded at 300, 400, 500 MHz (H) and at 75, 100, 125 MHz (C), respectively. Chemical shifts (δ) are reported in ppm, using the residual solvent peak in CDCl₃ (H: δ = 7.26 and C: δ = 77.0 ppm) as internal standard, and coupling constants (J) are given in Hz. HRMS were recorded using ESI-TOF techniques. Melting points of solids were recorded using Electrothermal (IA9100) melting point apparatus. Irradiation was performed with Penn PhD photoreactor M2 (PR M2) (blue LED, 450 nm) purchased from Sigma- Aldrich.

General procedure for the synthesis of alkylated 1,4-naphthoquinones. Selectfluor (0.4 mmol) and aliphatic Csp^3 -H component 2 (2.0 mmol) were weighed in a vial with rubber septum and MeCN (1.0 mL) was added to this mixture. The vial was back filled with N₂ and it was introduced into Penn PhD photoreactor M2 (blue LED 450 nm). To this mixture, a solution of 1,4-naphthoquinone 1 (0.2 mmol) in MeCN (1.0 mL) was added over a period of 20 min through syringe pump. After completion of reaction (1 h), the reaction mixture was concentrated using rotary evaporation and the crude reaction mixture was purified by column chromatography to get the desired compound **3**.

3-Cyclohexyl-1,4-dioxo-1,4-dihydronaphthalen-2-yl acetate (3a). Yellow syrup (32.2 mg, 54%); ¹H NMR (400 MHz, CDCl₃) δ 8.12 – 8.03 (m, 2H), 7.77 – 7.67 (m, 2H), 3.01 (tt, *J* 11.9, 3.4 Hz, 1H), 2.42 (s, 3H), 1.88 – 1.64 (m, 7H), 1.43 – 1.27 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 184.6, 178.5, 168.2, 151.2, 142.4, 134.1, 133.7, 132.4, 130.7, 126.8, 126.4, 36.4, 30.0, 26.7, 26.0, 20.6; IR (neat, cm⁻¹): 3020, 2930, 1651, 1513, 1215, 746; HRMS (ESI) calcd for C₁₈H₁₈O₄ [M]⁺: 298.1205; found: 298.1191.

2-Cyclohexyl-3-hydroxynaphthalene-1,4-dione (3b). Yellow solid (21.5 mg, 42%); mp 132-134 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.10 (d, *J* 7.6 Hz, 1H), 8.05 (d, *J* 7.5 Hz, 1H), 7.74 (t, *J* 7.4 Hz, 1H), 7.65 (t, *J* 7.4 Hz, 1H), 7.46 (s, 1H), 3.09 – 3.04 (m, 1H), 2.03 – 1.92 (m, 2H), 1.82 – 1.79 (m, 2H), 1.73 – 1.71 (m, 1H), 1.64 – 1.60 (m, 2H), 1.41 – 1.27 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 184.6, 182.0, 152.9, 134.9, 133.2, 132.7, 129.2, 127.9, 127.0, 125.9, 35.2, 29.7, 29.2, 26.8, 26.0; HRMS (ESI) calcd for C₁₆H₁₇O₃ [M+H]⁺: 257.1178; found: 257.1168. NMR data and m.p. (lit.133-135 °C) matches literature values.¹⁵

2-Cyclohexyl-3-(trifluoromethyl)naphthalene-1,4-dione (3c). Yellow syrup (33.9 mg, 55%); ¹H NMR (400 MHz, CDCl₃) δ 8.10 – 7.99 (m, 2H), 7.82 – 7.69 (m, 2H), 3.04 (tt, *J* 11.8, 3.1 Hz, 1H), 2.20 – 2.05 (m, 2H), 1.90 – 1.81 (m, 2H), 1.80 – 1.73 (m, 1H), 1.72 – 1.64 (m, 2H), 1.42 – 1.28 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 184.5, 180.9, 157.5, 134.3, 134.1, 132.1, 131.6, 126.6, 126.3, 122.4 (q, *J* 279.4 Hz), 42.2, 30.5, 26.7, 25.7; ¹⁹F NMR (377 MHz, CDCl₃) δ -56.4; IR (neat, cm⁻¹): 2928, 2857, 1673, 1595, 1281, 1137, 720; HRMS (ESI) calcd for C₁₇H₁₅F₃O₂ [M]⁺: 308.1024; found: 308.0978.

2-Cyclohexyl-3-methylnaphthalene-1,4-dione (3d). Yellow solid (16.8 mg, 33%); mp 78-79 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.07 – 8.00 (m, 2H), 7.71 – 7.62 (m, 2H), 2.88 (tt, *J* 12.1, 3.3 Hz, 1H), 2.23 (s, 3H), 2.11 – 2.01 (m, 2H), 1.89 – 1.82 (m, 2H), 1.78 – 1.73 (m, 1H), 1.65 – 1.59 (m, 2H), 1.39 – 1.31 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 185.7, 185.2, 150.6, 143.1, 134.7, 134.5, 133.4, 133.1, 132.8, 131.8, 127.5, 126.6, 126.2, 126.0, 40.8, 29.9, 27.0, 26.0, 12.6; HRMS (ESI) calcd for C₁₇H₁₈O₂ [M]⁺: 254.1307; found: 254.1313. NMR data and m.p. (lit.78 °C) matches literature values.¹⁴

2-Cyclohexyl-3-methoxynaphthalene-1,4-dione (3e). Yellow paste (16.7 mg, 31%); ¹H NMR (400 MHz, CDCl₃) δ 8.01 – 7.92 (m, 2H), 7.66 – 7.56 (m, 2H), 3.99 (s, 3H), 3.03 (tt, *J* 12.2, 3.3 Hz, 1H), 1.90 – 1.78 (m, 2H), 1.78 – 1.70 (m, 2H), 1.69 – 1.63 (m, 1H), 1.55 – 1.47 (m, 2H), 1.36 – 1.16 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 184.6, 180.9, 157.3, 138.7, 132.7, 132.1, 131.3, 130.4, 125.4, 124.9, 60.3, 34.9, 29.0, 25.9, 25.0; HRMS (ESI) calcd for C₁₇H₁₈O₃ [M+H]⁺: 270.1255; found: 270.1229. NMR data matches literature values.¹⁴

2-CyclohexyInaphthalene-1,4-dione (3f). Yellow paste (22.1 mg, 46%); ¹H NMR (500 MHz, CDCl₃) δ 8.12 – 8.07 (m, 1H), 8.07 – 8.01 (m, 1H), 7.75 – 7.68 (m, 2H), 6.73 (s, 1H), 2.95 – 2.85 (m, 1H), 1.88 – 1.81 (m, 4H), 1.80 – 1.75 (m, 1H), 1.51 – 1.40 (m, 2H), 1.30 – 1.17 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 185.7, 184.8, 156.3, 133.6, 133.6, 133.1, 132.5, 131.9, 126.7, 125.9, 36.7, 32.3, 26.4, 26.1; HRMS (ESI) calcd for C₁₆H₁₆O₂ [M+H]⁺: 240.1150; found: 240.1163. NMR data matches literature values.¹⁴

2-Cyclopentyl-3-hydroxynaphthalene-1,4-dione (3h). Yellow syrup (20.8 mg, 43%); ¹H NMR (500 MHz, CDCl₃) δ 8.12 (dd, *J* 7.7, 0.9 Hz, 1H), 8.06 (dd, *J* 7.6, 1.0 Hz, 1H), 7.75 (td, *J* 7.6, 1.3 Hz, 1H), 7.67 (td, *J* 7.5, 1.2 Hz, 1H), 7.44 (s, 1H), 3.51 – 3.37 (m, 1H), 1.97 – 1.81 (m, 6H), 1.68 – 1.64 (m, 2H).; ¹³C NMR (101 MHz, CDCl₃) δ 184.7, 181.6, 152.8, 134.9, 133.3, 132.8, 129.3, 127.6, 126.9, 126.0, 34.9, 30.8, 26.8; IR (neat, cm⁻¹): 2926, 2858, 1653, 1456, 1389, 1275, 770; HRMS (ESI) calcd for C₁₅H₁₅O₃ [M+H]⁺: 243.1021; found: 243.1026.

2-Cyclooctyl-3-hydroxynaphthalene-1,4-dione (3i). Brown color gum (26.1 mg, 46%); ¹H NMR (400 MHz, CDCl₃) δ 8.10 (dd, *J* 7.7, 1.1 Hz, 1H), 8.06 (dd, *J* 7.6, 1.1 Hz, 1H), 7.75 (td, *J* 7.6, 1.4 Hz, 1H), 7.67 (td, *J* 7.5, 1.3 Hz, 1H), 7.35 (s, 1H), 3.30 (tt, *J* 10.4, 3.1 Hz, 1H), 2.12 – 1.97 (m, 2H), 1.87 – 1.75 (m, 2H), 1.74 – 1.49 (m, 10H); ¹³C NMR (101 MHz, CDCl₃) δ 184.5, 182.1, 152.0, 134.9, 133.1, 132.7, 131.0, 129.3, 126.9, 125.9, 34.4, 31.4, 26.7, 26.5, 26.4; NMR data matches literature values.¹⁵

2-Benzyl-3-hydroxynaphthalene-1,4-dione 3j. Yellow solid (26.9 mg, 51%); mp 173-175 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.12 (dd, *J* 7.7, 1.0 Hz, 1H), 8.07 (dd, *J* 7.6, 1.1 Hz, 1H), 7.74 (td, *J* 7.6, 1.3 Hz, 1H), 7.67 (td, *J* 7.5, 1.3 Hz, 1H), 7.44 (s, 1H), 7.39 (d, *J* 7.3 Hz, 2H), 7.26 (t, *J* 7.3 Hz, 2H), 7.17 (t, *J* 7.3 Hz, 1H), 3.95 (s, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 184.4, 181.7, 153.1, 138.8, 138.1, 135.1, 133.0, 132.8, 129.9, 129.4, 128.3, 127.1, 127.0, 126.2, 126.1, 123.2, 29.0, 21.4; NMR data and m.p. (lit.174-175 °C)matches literature values.¹⁹

2-Hydroxy-3-(3-methylbenzyl)naphthalene-1,4-dione 3k. Brown color gum (22.8 mg, 41%); ¹H NMR (500 MHz, CDCl₃) δ 8.12 (dd, *J* 7.7, 1.0 Hz, 1H), 8.07 (dd, *J* 7.6, 1.0 Hz, 1H), 7.74 (td, *J* 7.6, 1.4 Hz, 1H), 7.67 (td, *J* 7.5, 1.3 Hz, 1H), 7.42 (s, 1H), 7.22 – 7.17 (m, 2H), 7.15 (t, *J* 7.7 Hz, 1H), 6.99 (d, *J* 7.3 Hz, 1H), 3.91 (s, 2H), 2.30 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 184.4, 181.7, 153.1, 138.8, 138.1, 135.0, 1323.0, 132.8, 129.9, 129.4, 128.3, 127.1, 127.0, 126.2, 126.1, 123.2, 29.0, 21.4; IR (neat, cm⁻¹): 3022, 1659, 1518, 1215, 747; HRMS (ESI) calcd for C₁₈H₁₄O₃ [M]⁺: 278.0943; found: 278.0947.

2-Hydroxy-3-(4-methylbenzyl)naphthalene-1,4-dione 3I. Yellow solid (24.5 mg, 44%); mp 162-164 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.11 (dd, *J* 7.7, 0.9 Hz, 1H), 8.06 (dd, *J* 7.6, 1.0 Hz, 1H), 7.74 (td, *J* 7.6, 1.4 Hz, 1H), 7.66 (td, *J* 7.5, 1.3 Hz, 1H), 7.41 (s, 1H), 7.28 (d, *J* 8.0 Hz, 2H), 7.07 (d, *J* 7.8 Hz, 2H), 3.91 (s, 2H), 2.28 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 184.4, 181.7, 152.9, 135.9, 135.0, 133.0, 129.2, 129.1, 126.9, 126.1, 123.3, 28.67, 21.1; HRMS (ESI) calcd for C₁₈H₁₄O₃ [M]⁺: 278.0943; found: 278.0947. NMR data and m.p. (lit.162-164 °C) matches literature values.¹⁹

2-(3,5-Dimethylbenzyl)-3-hydroxynaphthalene-1,4-dione 3m. Brown color gum (28.1 mg, 48%); ¹H NMR (500 MHz, CDCl₃) δ 8.05 (dd, *J* 7.7, 0.9 Hz, 1H), 7.99 (dd, *J* 7.6, 1.0 Hz, 1H), 7.69 – 7.65 (m, 1H), 7.62 – 7.58 (m, 1H), 7.34 (s, 1H), 6.92 (s, 2H), 6.74 (s, 1H), 3.80 (s, 2H), 2.19 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 184.4, 181.7, 153.0, 138.7, 137.9, 134.9, 132.9, 129.4, 128.0, 126.9, 126.9, 126.1, 123.2, 28.9, 21.3; IR (neat, cm⁻¹): 3020, 1658, 1520, 1215, 744; HRMS (ESI) calcd for C₁₉H₁₆O₃ [M]⁺: 292.1099; found: 292.1105.

2-(2-Bromobenzyl)-3-hydroxynaphthalene-1,4-dione 3n. Yellow solid (26.0 mg, 38%); mp 163-165 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.17 – 8.08 (m, 2H), 7.80 – 7.75 (m, 1H), 7.74 – 7.68 (m, 1H), 7.45 (s, 1H), 7.56 (dd, *J* 8.0, 0.9 Hz, 1H), 7.19 – 7.11 (m, 2H), 7.08 – 7.02 (m, 1H), 4.09 (s, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 184.2, 181.4, 154.0, 137.9, 135.1, 133.1, 132.8, 129.7, 129.4, 127.9, 127.3, 127.1, 126.3, 124.7, 121.4, 29.6; HRMS (ESI) calcd for C₁₇H₁₁BrO₃ [M]⁺: 341.9892; found: 341.9912. NMR data and m.p. (lit.162-164 °C) matches literature values.¹⁹

2-(1,4-Dioxan-2-yl)-3-hydroxynaphthalene-1,4-dione 3o. Brown color gum (24.5 mg, 47%); ¹H NMR (300 MHz, CDCl₃) δ 9.28 (s, 1H), 8.08 (t, *J* 6.7 Hz, 2H), 7.79 – 7.62 (m, 2H), 5.19 (dd, *J* 10.1, 2.7 Hz, 1H), 4.05 – 3.67 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 183.3, 180.5, 156.4, 134.7, 133.3, 132.0, 130.2, 126.5, 126.4, 117.5, 74.0, 68.7, 67.6, 66.3; IR (neat, cm⁻¹): 2928, 1728, 1680, 1650, 1278, 1118; HRMS (ESI) calcd for C₁₄H₁₃O₅ [M+H]⁺: 261.0763; found: 261.0764.

Acknowledgements

ADK thanks the Council of Scientific & Industrial Research (CSIR) and MRN thanks the Department of Science and Technology (DST), New Delhi, for a research fellowship. SMA is thankful to CSIR for funding the project (Ref. No. 34/1/TD-CLP/NCP-FBR 2020-RPPBDD-TMD-Se-MI) (IICT/pubs./2020/342).

Supplementary Material

¹H, ¹³C and ¹⁹F NMR associated with this article are available as supplementary data.

References

- 1. Asche, C. *Mini-Rev. Med. Chem.* **2005**, *5*, 449–467. https://doi.org/10.2174/1389557053765556.
- 2. Puder, C.; Wagner, K.; Vettermann, R.; Hauptmann, R.; Potterat, O. J. Nat. Prod. **2005**, *68*, 323–326. <u>https://doi.org/10.1021/np040150d</u>.
- 3. Inks, E. S.; Josey, B. J.; Jesinkey, S. R.; Chou, C. J. *ACS Chem. Biol.* **2012**, *7*, 331–339. <u>https://doi.org/10.1021/cb200134p</u>.
- 4. Sunassee, S. N.; Davies-Coleman, M. T. *Nat. Prod. Rep.* **2012**, *29*, 513–535. <u>https://doi.org/10.1039/c2np00086e</u>.
- 5. Son, E. J.; Kim, J. H.; Kim, K.; Park, C. B. *J. Mater. Chem. A*, **2016**, *4*, 11179–11202. https://doi.org/10.1039/c6ta03123d.
- 6. Breyer, S.; Effenberger, K.; Schobert, R. *ChemMedChem* **2009**, *4*, 761–768. <u>https://doi.org/10.1002/cmdc.200800430</u>.
- Khader, M.; Eckl, P. M. Iran. J. Basic Med. Sci. 2014, 17, 950–957. https://doi.org/10.22038/ijbms.2015.3851.
- 8. Josey, B. J.; Inks, E. S.; Wen, X.; Chou, C. J. *J. Med. Chem.* **2013**, *56*, 1007–1022. <u>https://doi.org/10.1021/jm301485d</u>.
- 9. Verma, R. Anticancer Agents Med. Chem. **2008**, *6*, 489–499. https://doi.org/10.2174/187152006778226512.
- 10. McHardy, N.; Hudson, A. T.; Morgan, D. W. T. *Res. Vet. Sci.* **1983**, *35*, 347–352. <u>https://doi.org/10.1016/s0034-5288(18)32031-9</u>.
- 11. Gutiérrez-Bonet, Á.; Remeur, C.; Matsui, J. K.; Molander, G. A. *J. Am. Chem. Soc.* **2017**, *139*, 12251–12258.

https://doi.org/10.1021/jacs.7b05899.

- 12. Liu, S.; Shen, T.; Luo, Z.; Liu, Z. Q. *Chem. Commun.* **2019**, *55*, 4027–4030. <u>https://doi.org/10.1039/c9cc01704f</u>.
- 13. Hamsath, A.; Galloway, J. D.; Baxter, R. D. *Synthesis* **2018**, *50*, 2915–2923. https://doi.org/10.1055/s-0037-1610005.

- 14. Sutherland, D. R.; Veguillas, M.; Oates, C. L.; Lee, A. L. *Org. Lett.* **2018**, *20*, 6863–6867. <u>https://doi.org/10.1021/acs.orglett.8b02988</u>.
- 15. Baral, E. R.; Kim, S. H.; Lee, Y. R. *Asian J. Org. Chem.* **2016**, *5*, 1134–1141. <u>https://doi.org/10.1002/ajoc.201600219</u>.
- 16. Liang, X. A.; Niu, L.; Wang, S.; Liu, J.; Lei, A. *Org. Lett.* **2019**, *21*, 2441–2444. <u>https://doi.org/10.1021/acs.orglett.9b00744</u>.
- 17. Niu, L.; Liu, J.; Liang, X. A.; Wang, S.; Lei, A. *Nat. Commun.* **2019**, *10*, 1–7. <u>https://doi.org/10.1038/s41467-019-08413-9</u>.
- 18. Zhao, H.; Jin, J. *Org. Lett.* **2019**, *21*, 6179–6184. https://doi.org/10.1021/acs.orglett.9b01635.
- 19. Ramachary, D. B.; Anif Pasha, M.; Thirupathi, G. *Angew. Chemie Int. Ed.* **2017**, *56*, 12930–12934. <u>https://doi.org/10.1002/anie.201706557</u>.

This paper is an open access article distributed under the terms of the Creative Commons Attribution (CC BY) license (<u>http://creativecommons.org/licenses/by/4.0/</u>)