Newly discovered naturally occurring organohalogens

Gordon W. Gribble

Department of Chemistry, Dartmouth College, Hanover, NH 03755, USA
E-mail: ggribble@dartmouth.edu

Dedicated to the memories of these pioneers in the field of marine natural products:
Paul J. Scheuer, Richard E. Moore, and D. John Faulkner

Received 05-02-2018  Accepted 08-26-2018  Published on line 11-03-2018

Abstract

Last year more than 200 naturally occurring organohalogens were described for the first time. These natural products were isolated and characterized from marine algae, sponges, corals, tunicates, bryozoans, marine and terrestrial fungi and bacteria, cyanobacteria, terrestrial plants, slime molds, red ants, and interstellar space. The following examples are illustrative of the extraordinary synthetic virtuosity of nature.

Keywords: Organohalogen, organochlorine, organobromine, organoiodine, marine natural products, terrestrial natural products, bacteria, fungi, sponges, algae, corals, marine organisms

DOI: https://doi.org/10.24820/ark.5550190.p010.610
Table of Contents

1. Introduction
2. Discussion
   2.1 Marine plants
   2.2 Sponges
   2.3 Corals
   2.4 Tunicates
   2.5 Bryozoans
   2.6 Marine fungi
   2.7 Marine bacteria
   2.8 Cyanobacteria
   2.9 Other marine organisms
   2.10 Terrestrial plants
   2.11 Terrestrial fungi
   2.12 Terrestrial bacteria
   2.13 Slime mold
   2.14 Miscellaneous
3. Conclusions
4. Acknowledgements
5. References

1. Introduction

From fewer than 50 known naturally occurring organohalogens in 1968, the number today – fifty years later – is more than 5,000. Three comprehensive reviews are available, as are more recent separate compilations of halogenated alkaloids, marine natural products, and halogenated heterocyclic compounds that occur naturally. A recent review of naturally occurring organoiodides has also appeared, and the annual review of marine natural products routinely covers new halogenated marine compounds for that year. It should be noted that previously known organohalogens and non-halogenated natural products discovered in the cited papers are not included in this review.

2. Discussion

2.1 Marine plants
The most prolific source of marine organohalogen natural products is the seaweed “limu kohu” (Asparagopsis taxiformis), a delicacy of native Hawaiians and which has yielded more 100 organohalogenes. The major component of this red alga is bromoform (CHBr₃), along with several lachrymatory α-haloketones. This author believes that the “smell of the ocean” is in large part due to the omnipresent organohalogens exuded by algae, CHBr₃, CH₂Cl₂, CHCl₃, CH₃Br, CH₃I, CH₂Br, CH₂Br₂, CH₂I₂, CH₃, CHCl₂Br, CHClBr₂, CHBr₂I, CHBrl₂, CHClBrI, CH₃CH₂Br, CH₃CH₂I, CH₃CH₂CH₂Br, CH₃CH₂CH₂I, (CH₃)₂CHI, and others.
The ubiquitous red algal genus *Laurencia* is a fertile source of organohalogens and last year saw the discovery of several new such metabolites. A collection of *Laurencia obtusa* from the Red Sea yielded the new maneonene, jeddahenyne A (1), and the novel isomaneonene, 12-debromo-12-methoxy isomaneonene A (2) along with several known compounds. An examination of *Laurencia viridis* from the Canary Islands has furnished four new acetogenins 3 - 6, which are variants of the known pinnatifidenyne C_{15} acetogenins (Figure 1). Oxirane 5 shows modest activity against four human solid tumor cell lines (GI_{50} = 13 - 48 µM), and 4 is active against two of the cell lines (GI_{50} = 33 - 45 µM).

**Figure 1.** Structures of new *Laurencia* sp. Organohalogens.

Eleven halogenated chamigrenes, compositacins (7 - 17), were isolated from the red alga *Laurencia composita* Yamada collected along the coast of Nanji Island in the East China Sea (Figure 2). Only compositacin G (13) shows good activity against the fungus *Microsporum gypseum* (MIC_{80} = 4 µg/mL), while compositacins D and G exhibit marginal cytotoxicity towards the A-549 human lung adenocarcinoma cell line (49 - 85 µM). The absolute configurations of compositacin B (8) was determined by the time-dependent density functional theory electronic circular dichroism (TDDFT-ECD) method, and those of compositacins A (7) and C - K (9 - 17) were established on biosynthetic grounds by comparison to compositacin B and to related sesquiterpenoids.
Four separate collections of Laurencia sp. (L. okamurae and L. nipponica) from Japanese coastal waters yielded the new omaezol (18), intricatriol (19), and hachijojimallenes A (20) and B (21) (Figure 3). Omaezol (18) and hachijojimallene A (20) demonstrate potent antifouling activity against larvae of the barnacle Amphibalanus amphitrite (EC$_{50}$ = 0.31 - 0.59 µM), comparable to CuSO$_4$ (EC$_{50}$ = 0.71 µM).

Figure 2. Structures of new Laurencia composita halogenated chamigrenes.

Figure 3. Structures of new Laurencia sp. halogenated metabolites from Japanese coastal waters.
The Chinese red alga *Symphyocladia latiuscula*, gathered from Qingdao coastal waters, contains ten new polybrominated phenols, the symphyocladins H–Q (22–31) (Figure 4).14 A biosynthetic pathway involving the coupling of a quinone methide from 2,3,6-tribromo-4,5-dihydroxybenzyl alcohol, a known natural phenol, and citric acid is proposed, but remains unsupported by experimental evidence.

![Figure 4. Structures of new *Symphyocladia latiuscula* polybrominated phenols.](image)

A Japanese sample of the red alga *Odonthalia corymbifera* afforded the two new bromophenols odonthalol (32) and odonthadione (33), the latter of which contains the cyclopentenedione unit previously unknown in natural products (Figure 5).15 Both phenols possess tyrosinase inhibitory and antioxidant activity.

Although naturally occurring organoiiodides are rare,2,3,8 a collection of the red alga *Callophyccus* sp. from Fiji furnished five novel iodinated meroditerpenes, iodocallophycoic acid A (34) and iodocallophycols A–D (35–38), and the related bromophycoic acid F (39) and bromophycoic acid A methyl ester (40) (Figure 6).16 Iodocallophycoic acid A (34) displays moderate activity against methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant *Enterococcus faecium* (VREF) (MIC = 1.4 and 2.2 µg/mL, respectively). This metabolite also potentiates the anti-MRSA activity of oxacillin leading to an 8-fold increase in potency of the latter antibiotic.
A large collection of polyhalogenated acetogenins was identified in the red alga *Ptilonia magellanica* collected off the coast of Chile. These included ptilonines A - F (41 - 46), magellenediol (47), and pyranosylmagellanicus D (48) and E (49). The absolute configuration of the previously known
pyranosylmagellanicus A (not shown) was determined by derivatization with (R)- and (S)-α-methoxy-α-phenylacetic acids.

Figure 7. Structures of new *Ptilonia magellanica* halogenated acetogenins.

Five tropical seaweeds, *Kappaphycus alvarezii*, *Padina australis*, *Sargassum binderi*, *Sargassum siliquosum*, and *Turbinaria conoides*, emit these well known halocarbons: CHBr₃, CH₂Br₂, CH₃I, CH₂I₂, CH₂BrI, CH₂BrCl, CHBrCl₂, and CHBr₂Cl. The effect of seawater pH on these emissions was recently examined, and it was found that decreasing pH led to an increase in selective halocarbon emissions.¹⁸

2.2 Sponges

Like most marine plants, but unlike fish and marine animals, sponges are anchored to the reef and must employ chemical defense for survival against predation. As we will see, the structural diversity of sponge metabolites is astonishing. It must be noted that the actual origin of some of these metabolites may be bacteria or microalgae associated with the sponge. This point will be visited later. These marine sponge antifouling metabolites were reviewed in 2017.¹⁹–²¹ Also reviewed last year were the metabolites from the sponge genus *Agelas*,²² and the cyclic azole-homologated peptides from sponges.²³

A common structural motif adopted by marine sponges is the pyrrole heterocycle. For example, 32 bromopyrrole alkaloids were isolated from the sponge *Stylissa massa* collected in Hainan Island, China. Of these metabolites five are new, stylisines A - E (50 - 54), which include the enantiomeric pair 53 and 54 (Figure 8).²⁴
Figure 8. Structures of new *Stylissa massa* brominated pyrroles.

The sponge *Agelas* sp. is a fruitful producer of bromopyrrole alkaloids and a recent collection of *Agelas* sp. from the South China Sea afforded four new dimeric bromopyrrole metabolites, hexazosceptrin (55), agelestes A and B (56, 57), and (9S,10R,9'S,10'R)-nakamuric acid (58) (Figure 9). The absolute configuration of all four metabolites was established. None of these compounds show activity against lymphoma U937 and lung cancer PC9 cells.

Figure 9. Structures of new *Agelas* sp. brominated pyrroles.
A deep-water (140 m) Palau Topsentia sp. sponge contains two novel brominated indoles, tulongicin A (59) and dihydrospogotive C (60), in addition to two known analogues (Figure 10). Their absolute configuration was determined, and both metabolites display strong antimicrobial activity against Staphylococcus aureus (MIC = 1.2 - 3.7 µg/mL).

![Figure 10. Structures of new Topsentia sp. brominated indoles.](image)

An examination of the Indonesian sponge Oceanapia sp. yielded the indole metabolite 6-bromo-8-keto-conicamin A (61), which displays strong activity against the human pancreatic cancer cell line PANC-1 (IC$_{50}$ = 1.5 µM). Four novel thiazole containing biakamides A - D (62 - 65) were isolated from another Indonesian sponge, Petrosaspongia sp (Figure 11). These unique polyketides are also active against PANC-1 (IC$_{50}$ = 0.5 - 4.0 µM). Total syntheses of all possible stereoisomers from the optically pure monoprotected 2,4-dimethyl-1,5-diol established the absolute configurations of the two secondary methyl groups.

![Figure 11. Structures of new Indonesian sponge metabolites.](image)
Sponges have a proclivity for incorporating the bromotyrosine unit in their metabolites\textsuperscript{2,3} and several new examples were reported in 2017. The Madagascan sponge *Amphimedon* sp. contains amphimedonoic acid (66) and psammaplyssene E (67), along with the known 3,5-dibromo-4-methoxybenzoic acid.\textsuperscript{29} Neither compound is active against human epidermoid carcinoma KB cells (IC\textsubscript{50} > 10 µg/mL). A sponge from Western Australia, *Pseudoceratina* cf. *verrucosa*, yielded pseudoceratinamide A (68) and B (69) and the enantiomer 70 of a previously known bromotyrosine (Figure 12).\textsuperscript{30}

![Figure 12. Structures of new sponge bromotyrosines.](image)

The Indonesian sponge *Iotrochota* cf. *iota* yielded seven new halogenated tyrosines, enisorines A - E (71 - 75), (+)-1-\textit{O}-methylhemibastadinol 2 (76), and (+)-1-\textit{O}-methylhemibastadinol 4 (77) (Figure 13).\textsuperscript{31} All seven metabolites inhibit T3SS-dependent YopE secretion, which is a virulence factor employed by many Gram-negative pathogens that injects bacterial effector proteins into host cells to negate host cell defenses.

![Figure 13. Structures of new *Iotrochota* cf. *iota* halogenated tyrosines.](image)
2.3 Corals

Whereas a few hard (stony) corals liberate organohalogen metabolites, soft corals (octocorals, sea fans, gorgonians) are extraordinarily generous producers of halogen-containing metabolites.\textsuperscript{2,3} These stunning marine animals, gently swaying in the reef currents, are both a delight for the scuba diver and a treasure trove for the marine natural products chemist seeking new metabolites.

The briarane (bicyclo[8.4.0]) diterpenoid carbon skeleton is pervasive in soft corals, as more than 600 examples are known from marine octocorals and gorgonians, many of which contain chlorine.\textsuperscript{2-4} A collection of the octocoral \textit{Briareum excavatum} from Taiwanese waters furnished three new briarenols, one of which, briarenol E (78), contains chlorine.\textsuperscript{32} The gorgonian coral \textit{Junceella fragilis} from Hainan Island, China, contains four pairs of chlorinated briarane diterpenes, five of which are new (79 - 83) (Figure 14).\textsuperscript{33} Interestingly, these pairs of isomers undergo acetyl migration (i.e., 80 ⇄ 81, and 82 ⇄ 83), which was observed for the first time. The acetyl migration of 79 yields the previously known fragilide J (2-deacetylpraeholide), which is also present in this coral. All of these metabolites inhibit the production of nitric oxide in RAW 264.7 cells.

\textbf{Figure 14.} Structures of new briarane chlorinated diterpenoids.

In addition to several known diterpenoids, another collection of \textit{Junceella fragilis} from Hainan Island by this same research group yielded seventeen new diterpenoids, ten of which contain chlorine (84 - 93) (Figure 15).\textsuperscript{34} Fragilolides D (86), G (89), and P (93) are inhibitory towards hepatitis B e antigen (HBeAg), but not active against the expression of hepatitis B surface antigen (HBsAg). None of these metabolites are cytotoxic in a panel of tumor cell lines.
2.4 Tunicates
Tunicates (ascidians, sea squirts) belong to subphylum Urochordata (or Tunicata) of the phylum Chordata, Class Ascidiacea. Like sponges, tunicates are filter feeders and rely on chemical defense for survival. They may be solitary or colonial marine animals. Some 3,000 species of tunicates have been described, and a recent review is available. The Korean tunicate *Pseudodistoma antinboja* yielded four new cadiolides J–M (94–97) (Figure 16), the only halogenated tunicate metabolites described in 2017.
2.5 Bryozoans

Unlike the majestic soft corals and brightly colored sponges, bryozoans (Phylum Bryoza = Ectoprocta) are nondescript “moss animals” that are ignored by scuba divers and snorkelers (in my experience). Nevertheless, these inauspicious marine animals produce remarkably complex halogen-containing metabolites.2,3

The Arctic bryozoan *Securiflustra securifrons* has furnished securamines H - J (98 - 100) (Figure 17).38 These halogenated indole-imidazole alkaloids were found in bryozoans living off the coast of Hjelmsøya, Norway. Securamines H and I are cytotoxic against these human cancer cell lines A2058 (melanoma), HT-29 (colon adenocarcinoma), and MCF-7 (breast adenocarcinoma) with IC50 µM values of 1.4 - 2.7, 1.9- 2.5, and 2.1 -2.4, respectively. Securamine J (100) is inactive in all three cell lines (IC50 > 50 µM), and securamine H (98) is slightly more active than securamine I (99).

Two new brominated indoles, terminoflustrindoles B (101) and C (102), were extracted from and identified in the bryozoan *Terminoflustra membranaceatruncata* that was collected in the White Sea (Figure 18).39
Figure 18. Structures of new bromoindoles from *Terminoflustra membranaceutruncata*.

**2.6 Marine fungi**

A relatively new source of metabolites is marine fungi and several examples were reported in 2017.

For example, the fungus *Hansfordia sinuosae*, found with the sponge *Niphates* sp. From the South China Sea, afforded the novel chlorinated resorcinol hansfordiols H - J (103 - 105), in addition to seven non-chlorinated analogues.\(^{40}\) Hansfordiols H and I show good antioxidant activity comparable to Trolox, but no antibacterial activity or cytotoxicity against several cell lines. The fungus *Penicillium* sp. SCS-KFD09, found growing on the marine worm *Sipunculus nudus*, is the source of the six new meroterpenoids, two of which contain chlorine, chrodrimanins K (106) and L (107).\(^{41}\) The former meroterpenoid displays influenza A virus (anti-H1N1) activity (IC\(_{50}\) = 74 µM), compared to the positive control ribavirin (IC\(_{50}\) = 103 µM). Of five new meroterpenoids isolated from a *Penicillium* sp. SCS-KFD09 fungus living with the marine worm *Sipunculus nudus* from Haikou Bay, China, one contains chlorine: the unusual trichlorinated chrodrimanin O (108) (Figure 19).\(^{42}\) It displays inhibition of protein tyrosine phosphatase 1B (PTP1B) (IC\(_{50}\) = 71.6 µM), but no cytotoxicity towards three tumor cell lines (A549, HepG2, and HeLa) at 10 µM.

Figure 19. Structures of new chlorinated metabolites from marine fungi.

Another set of meroterpenoids, the chartarolides A - C (109 - 111), was identified in the fungus *Stachybotrys chartarum* WGC-25C-6, which is found with the sponge *Niphates recondite* (Figure 20).\(^{43}\) All three compounds exhibit significant antitumor activity; for example, chartarolide A shows inhibition of these
human cell lines: HCT-116 (colon, IC$_{50} = 1.9$ µM), HepG2 (liver, IC$_{50} = 1.8$ µM), BGC-823 (gastric, IC$_{50} = 1.3$ µM), NCI-H1650 (lung, IC$_{50} = 5.5$ µM), A2780 (ovarian, IC$_{50} = 1.5$ µM), MCF7 (ovarian, IC$_{50} = 1.4$ µM). Chartarolides A and B also inhibit several tumor-related protein kinases (IC$_{50} = 2.6 – 20.3$ µM).

Figure 20. Structures of new chlorinated metabolites from Stachybotrys chartarum WGC-25C-6.

The marine fungus Penicillium sp. F37 is found with the Brazilian coastal sponge Axinella corrugata and produces arvoredol (112), a novel chlorinated polyketide that inhibits biofilm formation on the sponge.$^{44}$ This metabolite also inhibits biofilm formation of the human pathogen Staphylococcus epidermidis without serving as an antibiotic. The mangrove soil-derived fungus Penicillium janthinellum HK1-6 has yielded two chlorine-containing azaphilones, penicilones C (113) and D (114).$^{45}$ Both compounds show potent anti-MRSA (Staphylococcus aureus ATCC 43300 and ATCC 33591) activity (MIC 3.13–6.25 µg/mL). Penicilone D also displays strong activity against vancomycin-resistant Enterococcus faecalis. This suggests that the penicilones (including the two non-chlorinated penicilones A and B) may have the capability to circumvent antibiotic cross-resistance. Another marine-derived Penicillium sp. 5CS10 sof 101 fungus from a South China Sea sediment furnished three new chlorinated emodacidamides C (115), F (116), and G (117), in addition to five non-chlorinated analogues (Figure 21).
2.7 Marine bacteria

Bacteria live everywhere, including in the marine environment. A few halogenated bacterial metabolites from the oceans were described in 2017. Cultures of the marine bacterium *Pseudovibrio denitrificans* Ab134, which were isolated from the sponge *Arenosclera brasiliensis*, yielded several known bromotyrosine-derived alkaloids that were previously only isolated from marine sponges.\(^{47}\) This work shows for the first time that bromotyrosine-derived alkaloids can be biosynthesized by a marine bacterium. The isolated or spectroscopically identified alkaloids are fistularin-3, 11-hydroxyaerothionin, verongidoic acid, aerothionin, homopurpuroceratic acid B, purealidin L, and alysinamisine II, all previously isolated only from *Verongida* sponges.\(^{2,3}\)

A Palau sediment sample yielded a new streptomycete strain that produces marinocyanins A - F (118 - 123), which are novel brominated phenazinone meroterpenoids (Figure 22).\(^{48}\) Marinocyanin A is the most potent antifungal agent of the six marinocyanins against amphotericin-resistant *Candida albicans* (MIC = 0.95 µM). Both marinocyanins A and B display potent cytotoxicity against HCT-116 human colon carcinoma (IC\(_{50}\) = 0.049 and 0.029 µM, respectively).
A sediment sample from the Great Salt Lake, Utah, contains *Streptomyces* sp. GSL-6B that afforded three heptapeptides, the bonnevillamides A - C (124 - 126) (Figure 23). These novel natural products feature
unprecedented non-proteinogenic amino acids, and all three contain the bonnevilllic acid unit (3-(3,5-dichloro-4-methoxyphenyl)-2-hydroxyacrylic acid). Bonnevillamide A (124) has the extremely rare 4-methylazetidine-2-carboxylic acid methyl ester moiety. These metabolites have no antimicrobial activity, but bonnevillamide B (125) modulates heart growth and cardiac function in zebrafish embryo.

2.8 Cyanobacteria

As of 2009 more than 1,000 natural products were identified in cyanobacteria, also known as blue-green algae. Many of these organisms contain organochlorines and some are highly toxic to humans. For example, Nostoc cyanobacteria are well recognized as a threat to humans when they infest drinking water. The frequent infestations of Lake Erie by Microcystis aeruginosa cyanobacteria result in serious health concerns.

The global marine cyanobacterium Trichodesmium thiebautii (order Oscillatoriales) collected from a bloom near Padre Island, Texas, in the Gulf of Mexico afforded trichophycin A (127). This novel metabolite exhibits moderate cytotoxicity against Neuro-2A and HCT-116 cells (EC50 = 6.5 and 11.7 µM, respectively). Another collection of a Trichodesmium bloom from the Gulf of Mexico yielded the dichlorinated polyketide trichothiazole A (128). This metabolite expresses moderate cytotoxicity towards Neuro-2A cells (EC50 = 13.3 µM). The Panamanian cyanobacterium cf. Symploca sp. produces the novel gem-dichlorovinylidene containing caracolamide A (129) (Figure 24). Although not cytotoxic to NCI-H460 human non-small-cell lung cancer cells (IC50 > 10 µM), caracolamide A has calcium influx and calcium channel oscillation modulatory activity as low as 10 pM. Its structure was confirmed by synthesis from 2-phenethylamine.

A Malaysian cyanobacterium, Moorea bouillonii, contains the novel chlorinated fatty acid amides columbamides D (130) and E (131), the structures of which were confirmed by total synthesis of all four stereoisomers (of 130). Subsequent chiral-phase HPLC determined the absolute configuration. These two (synthetic) compounds are noncytotoxic at 22 µM to MCF7 (breast) and H460 (lung) human cancer cells. An examination of Moorea producens from Okinawa revealed three new malyngamides, 132 – 134 (Figure 25). These compounds stimulated glucose uptake in cultured L6 myotubes.
Figure 25. Structures of new chlorinated metabolites from *Moorea* sp. cyanobacteria.

An *Okeania* sp. of cyanobacteria from the Red Sea afforded two new chlorinated lyngbyabellins O (135) and P (136) in addition to several previously known analogues (Figure 26). Both compounds display strong antifouling activity against the barnacle *Amphibalanus amphitrite* (EC$_{50}$ = 0.38 - 0.73 µM), and lyngbyabellin P (136) has potent inhibitory action against MCF-7 breast cancer cells (GI$_{50}$ = 9 µM).

Figure 26. Structures of new chlorinated metabolites from *Okeania* sp. cyanobacteria.
A bloom of *Lyngbya* sp. cyanobacteria growing in the Kemp Channel in the Florida keys yielded kempopeptin C (137). This novel cyclic depsipeptide shows antiproteolytic activity against trypsin, plasmin, and matriptase (IC$_{50}$ = 0.19, 0.36, and 0.28 µM, respectively). A rare brominated cyclodepsipeptide, odobromoamide (138), was characterized from the cyanobacterium *Okeania* sp. found off the coast of Odo, Okinawa (Figure 27). This metabolite is active against HeLa S3 cells (IC$_{50}$ = 0.31 µM).

![Figure 27](image1.png)

**Figure 27.** Structures of new halogenated metabolites from *Lyngbya* sp. and *Okeania* sp. cyanobacteria.

### 2.9 Other marine organisms

Marine brittle stars (Ophiuroidea) are a very large group of reclusive, solitary echinoderms that inhabit the world’s oceans. They are typically found hiding under rocks.

A brittle star, *Ophionereis reticulata*, from the coast of Brazil contains two known chamigrene sesquiterpenes in addition to the novel acetyl isoobtusadiene (139). The known metabolites found in this animal are elatol and isoobtusadiene, which are common in *Laurencia* red algae, suggesting a dietary origin for these compounds. The halogenated diterpene dolabellol A (140) was characterized from the opisthobranch *Dolabella auricularia* found living off the Japanese coast (Figure 28). The absolute configuration of 140 was established by a combination of spectroscopy, chemical degradation, and X-ray crystallography. As in the previous study, it is suggested that this animal acquires dolabellol A from its algae diet. Accordingly, dolabellol A is structurally similar to the metabolites obtusadiol, rogioldiol A, laurenditerpenol, and 14-bromoobtus-1-ene-3,11-diol, all of which are found in the algae on which *Dolabella auricularia* feeds.

![Figure 28](image2.png)

**Figure 28.** Structures of new halogenated metabolites from marine animals.

### 2.10 Terrestrial plants

Lacking the huge concentration of halide (i.e., chloride, bromide) in the oceans, terrestrial organisms manufacture far fewer halogen-containing metabolites than their marine counterparts. Nevertheless, last year several new examples of organochlorine compounds in terrestrial plants, fungi, and bacteria were reported. An excellent review of chlorinated plant steroids and their biological activities has appeared.
series of steroidal withanolides, including eleven new examples and two known chlorine-containing ones, were isolated from the plant *Physalis peruviana* L. (Solanaceae), and were evaluated for their cytotoxicity against prostate and renal cancer cells. The two known chlorinated examples (not shown) physalolactone and 4-deoxyphysalolactone are inactive.

The small shrub *Uvaria alba* Merr. from Luzon Island, Philippines, and reported to have anti-infective and cytotoxic activities, contains two novel chlorine-containing polyoxygenated seco-cyclohexenes, albanols A (141) and B (142). Both albanols show modest activity against *Mycobacterium tuberculosis* H37Rv (MIC = 26 - 38 µM), and albanol A exhibits cytostatic activity towards HeLa cells. A collection of the traditional medicinal plant *Cleistochlamys kirkii* from Tanzania yielded 13 new metabolites including two chlorinated cyclohexenes, cleistenechlorohydrins A (143) and B (144) (Figure 29). This plant is a member of the genus of the family Annonaceae and is native to several eastern and southern African countries. For example, in Mozambique this plant is used to treat wound infections, rheumatism, and tuberculosis.

Chinese agarwood (*Aquilaria sinensis* Lour.) Gilg. (Thymelaeaceae) is the source of three new chlorinated 2-(2-phenylethyl)chromones, 145 - 147, in addition to two non-chlorinated analogues and 11 known compounds. The absolute configurations were determined and these compounds exhibit strong inhibition of nitric oxide production in RAW 264.7 cells (IC$_{50}$ = 3.8 – 7.3 µM). A collection of *Seidlitzia rosmarinus* from the Sinai desert shoreline of the Gulf of Aqaba, Egypt, yielded the novel, isomeric α-chloroferuloylamides 148 and 149 (Figure 30).
The traditional Chinese medicine plant *Curculigo orchioides* (“Xianmao”) collected from Yunnan Province, China, afforded five new chlorinated phenolic glycosides, curculigines J - N (150 - 154). Three related glycosides were isolated from *Przewalskia tangutica* (Solanaceae), przewatangosides A - C (155 - 157), a plant found in the Tibet region of China (Figure 31). Only 155 shows (weak) activity against SMMC-7721 (liver carcinoma) (IC$_{50}$ = 38.1 µM).

![Structures of new chlorinated phenolic glycosides from terrestrial plants](image)

**Figure 31.** Structures of new chlorinated phenolic glycosides from terrestrial plants.

The Chinese herb *Valeriana jatamansi* (Caprifoliaceae), which is an important traditional Chinese medicine for the treatment of nervous disorders, epilepsy, insanity, snake poisoning, and skin diseases, furnished three new chlorinated iridoids, chlorovalt rates P - R (158 - 160), in addition to five previously known chlorinated iridoids. A collection of *Phlomis likiangensis* (Lamiaceae) from Yunnan, China, yielded six new iridoids including the two new chlorine-containing phlolosides E (161) and F (162) (Figure 32). Like *Valeriana jatamansi* (*vide supra*), the *Phlomis* genus is used as a herbal tea to cope with gastrointestinal diseases and other disorders. Chlorine-containing iridoids are probably the largest collection of organochlorines present in the terrestrial environment.²,³
Another traditional Chinese medicine plant for pain relief dating back to the 1st century is *Rhododendron molle* G.Don. A gathering of the fruits of this plant from Guangxi Province, China, led to the discovery of three new chlorinated diterpenoids rhodomollein XXXI - XXXIII (163 - 165) in a group of 12 new compounds isolated in this study (Figure 33). All three metabolites show significant antinociceptive activity, especially 164 and 165 at a very low dose (2 mg/kg).

In contrast to chlorinated terpenoids (*vide supra*), chlorinated plant alkaloids are exceedingly rare. Two examples were identified in 2017. The plant *Ficus fistulosa* var. *tengerensis* (Moraceae) from Malaysia contains the novel tengechlorenine (166) as a pair of phenanthroindolizidine enantiomers. This alkaloid shows strong cytotoxicity against three breast cancer cell lines, MDA-MB-468, MDA-MB-231, and MCF-7 (IC$_{50}$ = 0.038 - 0.91 µM). The widely distributed plant *Rauvolfia vomitoria* (Apocynaceae) contains the unusual alkaloid rauvomine A (167). This plant is found in the tropical regions of Africa and Asia, and has been used to treat fever, gastrointestinal and liver diseases, pain, and some cancers.
2.11 Terrestrial fungi

Terrestrial fungi are prodigious fabricators of natural products, many of which contain halogen. New examples are discovered yearly.\(^2,3\)

The genus *Colletotrichum* includes a large group of fungal plant pathogens that cause disease to many crops, and this fungus is extremely detrimental to agriculture.\(^8^2\) An examination of the fungus *Colletotrichum higginsianum* afforded the novel colletochlorins G (168) and H (169) along with five known related metabolites.\(^8^3\) The absolute configuration of the colletochlorins could not be determined. Five new and five known metabolites were isolated from the ubiquitous fungus *Aspergillus unguis* that includes the novel aspergillusethers C (170) and D (171) (Figure 35).\(^8^4\) This fungal sample was collected from Surat Thani province in Thailand. The dichlorinated 171 is 4 to 8 times more active than the monochlorinated 170 in antifungal activity towards *Candida albicans*, *Cryptococcus neoformans*, and *Penicillium marneffei* (MIC = 16, 8, 16 µg/mL, respectively).

Figure 34. Structures of new chlorinated alkaloids from terrestrial plants.

Figure 35. Structures of new chlorinated metabolites from terrestrial fungi.

Fermentation broths of *Penicillium concentricum*, an endophytic fungus of the Liverwort *Trichocolea tomentella* (Trichocoleaceae), produced an array (more than 20) of metabolites including the novel 172 - 175.\(^8^5\) Metabolites 172 and 173/174 are cytotoxic to MCF-7 breast cancer cells (IC\(_{50}\) = 8.4 and 9.7 µM,
This liverwort was collected in Newport, Virginia. The mangrove *Bruguiera sexangula* var. *rhynchopetala* from the South China Sea is host to the fungus *Penicillium citrinum* HL-5126, and the latter produces the chlorinated xanthone 176 and anthraquinone 177 (Figure 36). Metabolite 177 is antibacterial against *Vibrio parahaemolyticus* (MIC = 10 µM).

![Chemical structures](image)

**Figure 36.** Structures of new halogenated metabolites from terrestrial *Penicillium* sp. fungi.

The fungal pathogen *Cochliobolus australiensis* was isolated from infected leaves of the invasive weed “buffel grass” *Pennisetum ciliare* (syn. *Cenchrus ciliaris*), which is a major problem in southern Arizona. From this pathogen were isolated the new metabolites chloromonilinic acids C (178) and D (179), in addition to some previously known metabolites. The stem bark of the Chinese tree *Melia azedarach* Linn. from Jiangsu Province, China, is associated with the fungus *Pestalotiopsis* sp. Extraction of this fungus led to the novel pestalchloride G (180) as a racemate. It shows strong antimicrobial activity against several pathogenic bacteria (*Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and *Bacillus subtilis*; MIC<sub>50</sub> = 4.1, 15.0, 13.5, 16.5 µg/mL, respectively). Both enantiomers of 180 show similar bioactivities. Cultures of the fungus *Helminthosporium velutinum* yone96, which was isolated from dead twigs of a woody plant from Yakushima Island, Japan, led to cyclohelminthol X (181). This complex metabolite shows activity against...
COLO 201 (colon) and (especially) HL-60 (leukemia) cells (IC₅₀ = 16 and 0.35 µM, respectively), and displays proteasome inhibition. Some fungal metabolites are structurally simple, such as 8-chloroxylarinol A (182) isolated from *Malbranchea flavorosea* (Figure 37). Metabolite 182 is a strong inhibitor of α-glucosidase. The genus *Malbranchea* (Myxotrichaceae) is a worldwide soil-based fungus.

![Image of metabolites 178, 179, 180, 181, and 182](image)

**Figure 37.** Structures of new chlorinated metabolites from terrestrial fungi.

A classic antifungal agent is griseofulvin and a new derivative of it was isolated from *Penicillium griseofulvum* CPCC 400528, 4'-demethoxy-4'-N-isopentylisogriseofulvin (183), along with several other metabolites including griseofulvin. This new metabolite is active against HIV (IC₅₀ = 33.2 µM). The mangrove plant *Sonneratia caseolaris* in Hainan Province, China, is associated with the endophytic fungus *Penicillium janthinellum* HDN13-309 and the latter yielded a series of alkaloids, including the chlorine-containing penicisulfuranols A (184) and D (185) along with four non-chlorinated analogues (Figure 38). Metabolite 184 shows potent cytotoxicity towards HeLa and HL-60 cells (IC₅₀ = 0.5 and 0.1 µM, respectively). These novel compounds possess a rare 1,2-oxazadecalin core and the unusual spiro-furan ring.

![Image of metabolites 183, 184, and 185](image)

**Figure 38.** Structures of new chlorinated metabolites from terrestrial *Penicillium* fungi.
The legume-infesting fungus *Diaporthe toxica* causes fatal liver disease in lupin-fed sheep. The major responsible toxin is phomopsin A. The present study discovered a new metabolite of this fungus, phomopsin F (186), the *N*-methylated derivative of phomopsin A. The new dichlorinated dehydrocurvularin 187 was characterized from *Alternaria* sp. AST0039, which is a fungal endophyte of *Astragalus lentiginosus* (Fabaceae, “spotted locoweed”), collected in central Arizona. The chlorinated lepistatins A - C (188 - 190) were isolated from the culture broth of the basidiomycete *Lepista sordida* (Figure 39). No significant antibacterial and antiproliferative activities of these lepistatins are observed at 25 µg/mL.

![Figure 39. Structures of new chlorinated metabolites from terrestrial fungi.](image)

*B. laricina* is a fungus associated with the moss *Rhodobryum umgiganteum* living in Yunnan Province, China. This fungus produces the new chlorinated cyclohexenones botrysphones A (191) and C (192), along with a suite of other metabolites, including the known chlorosphaeropsidone. The novel cosmochlorins D (193) and E (194) were characterized from endophytes associated with the shrub *Ficus ampelas* (Moraceae) (Figure 40). The producing organism is *Phomopsis* sp. N-125. Both 193 and 194 are cytotoxic towards HL-60 cells (IC$_{50}$ = 6.1 and 1.8 µM, respectively).

![Figure 40. Structures of new chlorinated metabolites from terrestrial fungi.](image)
2.12 Terrestrial bacteria

No other organism produces metabolites more complex than those produced by terrestrial bacteria. Some of these natural products are life-saving antibiotics, like the glycopeptide vancomycin. An examination of *Actinoallomurus* sp. ID145698 revealed the presence of new angucyclinones, the allocyclinones A–D (195–198). These metabolites are active towards Gram positive bacteria with MIC values of 0.25–1 µg/mL, with the exception of *Enterococcus faecium*. The activity increases with the increasing number of chlorines (195 > 198 > 197 > 196). A soil-derived *Actinomadura* strain produces the novel polyether polyketide nonthmicin (199) (absolute configuration shown), which shows potent antimicrobial activity (IC_{50} = 0.0013–0.005 µg/mL) against *Kocuria rhizophia*, *Bacillus cereus*, *Staphylococcus aureus*, and *Enterococcus faecalis* (Figure 41). Interestingly, the dechlorinated ecteinamycin metabolite is much less active against all four organisms (MIC = 0.01 µg/mL).

![Structures of new metabolites from terrestrial bacteria](image)

**Figure 41.** Structures of new chlorinated metabolites from terrestrial bacteria.

The rare *Microbacterium* sp. was isolated from the gut of the carrion beetle (*Nicrophorus concolor*) and found to produce the novel chlorinated cyclic peptides microphorusamides A (200) and B (201). The former metabolite 200 is eight times more active than 201 against several pathogenic bacterial strains (*Staphylococcus aureus*, *Enterococcus faecalis*, *Enterococcus faecium*, and *Salmonella enterica*) (MIC = 8–16 µg/mL), but both 200 and 201 are inactive towards several pathogenic fungi and display no cytotoxicity against various human cancer cell lines. The strain *Streptomyces* sp. KCB13F003 has led to two ulleungmycins A (202) and B (203), which are very similar to the microphorusamides 200 and 201 (Figure 42).
Figure 42. Structures of new chlorinated cyclic peptides from terrestrial bacteria.

Another *Streptomyces* sp. (DSM14386) afforded seven new halogenated peptides, the svetamycins A - G (204 - 210) (Figure 43). Svetamycin G (210) is the most active of these metabolites against the growth of *Mycobacterium smegmatis* (MIC$_{80}$ = 2 µg/mL). Svetamycins A (204) and C (206) are cytotoxic towards HepG2 (hepatoma), IC$_{50}$ = 11.0 and 3.6 µg/mL, respectively. A study of the several microsclerodermins from terrestrial myxobacteria uncovered the new microsclerodermin L (211).
2.13 Slime mold

Once classified as a fungus, slime mold (slime mould) is the name given to several unrelated organisms that either live freely as single cells or as unified structures. Some 900 species are known worldwide.

The bacteria-eating slime mold *Dictyostelium monochasioides* produces eight chlorinated alkylresorcinols, monochasiols A–H (212–219), the structures of which were confirmed by synthesis (Figure 44).\(^{104}\) Monochasiol A (212) inhibits the concanavalin A-induced interleukin-2 production in Jurkat cells, which is a human T lymphocyte cell line.

**Figure 43.** Structures of new chlorinated cyclic peptides from terrestrial bacteria.
Figure 44. Structures of new chlorinated alkylresorcinols from *Dictyostelium monochasoides*.

2.14 Miscellaneous

Two other new, and quite surprising, natural sources of organohalogens were described in 2017.

As do termites, red wood ants produce CH$_3$Cl, CHCl$_3$, CCl$_4$, and CHBr$_3$. For example, the average concentrations in the nests of *Formica rufa* and *Formica polyctena* are up to three-fold higher than the atmospheric background and as much as 70-fold higher than volcanic emissions, which are generally considered as one of the main geogenic sources of CHCl$_3$. Bromoform was up to 20-fold higher than atmospheric background.

An astounding finding was the interstellar detection of CH$_3$Cl and CH$_3$F in the gas surrounding the protostar IRAS 16293-2422, and the presence of CH$_3$Cl in the coma of the comet 67P/Churyumov-Gerasimenko. The authors of this discovery calculate that approximately 600 tons/year of CH$_3$Cl could have been delivered to the young earth based on the cometary CH$_3$Cl abundance and during the heavy bombardment over 80 million years. This amounts to 50 gigatons of CH$_3$Cl.

Halocarbon emissions from marine phytoplankton as influenced by climate change was reviewed in 2017. Some 40 low-molecular weight organohalogens were evaluated in these studies. Also evaluated were the formations of haloacetic acids and trihalomethanes that are produced by peracetic acid and chlorine drinking water disinfection processes. No new organohalogens were described in either of these three investigations.
3. Conclusions

Contrary to the widespread belief – pervasive prior to 1980 – that “nature would never make halogen-containing natural products,” the past several decades have shown that literally thousands of organohalogen natural products have now been identified,\(^2,3\) approximating 6,000! As we have seen in this brief review, the frequency of the discovery of new naturally occurring organohalogen compounds — 100–200 per year — has sustained for the year 2017. Marine and terrestrial organisms alike continue to amaze with their inexorable output of novel halogen-containing compounds. Given the fact that of the 500,000 estimated marine organisms\(^1\) — which are the source of most organohalogens — only a small percentage have been investigated for their chemical content, it is certain that myriad new natural organohalogens are awaiting discovery. Of the estimated 1.5 million species of fungi, secondary metabolites have been characterized from only 5,000 species.\(^2\) The future is bright for the collector of naturally occurring organohalogens!

This increased activity in natural products research, of all types, may be attributed to modern collection methods (e.g., SCUBA and remote submersibles for the collection of previously inaccessible marine organisms), selective bioassay for identifying biologically active compounds, powerful multidimensional NMR and mass spectral techniques for characterizing sub-milligram amounts of compounds, and new separation and purification techniques (e.g., counter-current chromatography and HPLC). All of this ensures that even the most structurally intricate natural products can be identified. Finally, knowledge and appreciation of folk medicine and ethobotany will continue to guide chemists to new biologically active natural products, including organohalogen compounds.

But what is the \textit{raison d'être} for the existence of natural organohalogens? This is the big question! Some organohalogens function as pheromones and hormones, as antifeedants and antifoulants, as natural pesticides, as recyclers of halogen, and as structural proteins. Many organohalogen compounds display enormous biological activity that may lead to clinical drugs of benefit to mankind.

4. Acknowledgements

The author is indebted to Wendy Berryman for typing this manuscript and drawing the chemical structures.

5. References

   \url{https://doi.org/10.1098/rspb.1968.0052}
   \url{https://doi.org/10.1071/EN15002}
   \url{https://doi.org/10.3390/md13074044}
   \url{https://doi.org/10.3987/REV-11-SR(P)5}
   \url{https://doi.org/10.1039/C4RA09833A}
   https://doi.org/10.1039/C7NP00052A
   https://doi.org/10.1016/j.phytochem.2017.08.001
   https://doi.org/10.3390/md16010005
   https://doi.org/10.1016/j.phytochem.2017.08.001
   https://doi.org/10.3390/md15090267
   https://doi.org/10.3390/md15120374
   https://doi.org/10.1016/j.tetlet.2017.09.044
   https://doi.org/10.1021/acs.joc.7b00096
   https://doi.org/10.1016/j.phytochem.2017.06.010
   https://doi.org/10.7717/peerj.2918
   https://doi.org/10.3390/md15090263
   https://doi.org/10.3390/md15090266
   https://doi.org/10.3390/md15090266
   https://doi.org/10.3390/md15110351
   https://doi.org/10.1039/C7OB02628E
   https://doi.org/10.3390/md16010009
   https://doi.org/10.1016/j.tet.2017.03.078


https://doi.org/10.1016/j.fitote.2017.08.002
https://doi.org/10.1016/j.tetlet.2017.03.079
https://doi.org/10.1016/j.phytol.2017.04.010
https://doi.org/10.1021/acs.jnatprod.6b01179
https://doi.org/10.1021/acs.jnatprod.7b00269
https://doi.org/10.1021/acs.jnatprod.6b00838
https://doi.org/10.1016/j.tet.2017.03.003
https://doi.org/10.3390/md15070195
https://doi.org/10.1021/np800751j
https://doi.org/10.1016/j.copbio.2010.01.005
https://doi.org/10.1351/pac198052010165
https://doi.org/10.2307/1297756
https://doi.org/10.3390/md15010010
https://doi.org/10.1016/j.tetlet.2017.09.027
https://doi.org/10.1021/acs.jnatprod.7b00367
https://doi.org/10.1021/acs.orglett.7b01869
   https://doi.org/10.1021/acs.jnatprod.7b00449
   https://doi.org/10.3390/md15090290
   https://doi.org/10.1246/bcsj.20160417
   https://doi.org/10.1246/cl.170756
   https://doi.org/10.20546/ijcrbp.2017.411.009
   https://doi.org/10.1021/acs.jnatprod.7b00106
   https://doi.org/10.1002/cbdv.200590087
https://doi.org/10.1016/j.phytochem.2017.05.010
https://doi.org/10.1016/j.tetlet.2017.06.078
https://doi.org/10.1016/j.tet.2017.12.017
https://doi.org/10.1021/acs.jnatprod.7b00500
https://doi.org/10.1021/acs.orglett.7b01723
https://doi.org/10.1111/j.1364-3703.2011.00783.x
https://doi.org/10.1016/j.tet.2017.10.018
https://doi.org/10.1016/j.tet.2017.08.039
https://doi.org/10.1021/acs.jnatprod.6b01069
https://doi.org/10.1038/ja.2017.52
https://doi.org/10.1021/acs.jnatprod.7b00583
https://doi.org/10.1016/j.phytol.2017.10.006
https://doi.org/10.1021/acs.joc.7b00393
https://doi.org/10.1021/acs.jnatprod.6b00977
https://doi.org/10.1021/acs.jnatprod.6b00829
https://doi.org/10.1021/acs.jnatprod.6b00483
https://doi.org/10.1021/acs.jnatprod.6b00662


    https://doi.org/10.1016/j.chemosphere.2017.09.059

    https://doi.org/10.1016/B978-0-12-624004-7.50011-1

    https://doi.org/10.1002/ps.2780390210