

Convenient syntheses of new quinoline nucleosides bearing amino acid esters

Ibrahim A. I. Ali

Department of Chemistry, Faculty of Science, Suez Canal University, Ismailia, Egypt
E.mail: ibrahim3369@yahoo.com

Dedicated to Prof. Ezz El-Din M. S. Salem for his 70th birthday

Abstract

Quinoline reverse nucleosides **4**, **5** were prepared by reaction of quinolines **1**, **2** with methyl 2,3-*O*-isopropylidene-5-*O*-*p*-toluenesulfonyl- β -D-ribofuranoside (**3**) in the presence of sodium hydride. Quinoline nucleosides bearing an amino acid ester residue **12-16** were prepared by azide coupling method from ester **4**. The synthesized compounds were characterized by elemental analysis, MALDI MS and NMR data.

Keywords: Nucleosides, quinoline, amino acids and dipeptide, azide coupling, ribose-series

Introduction

The synthesis of quinoline and its derivatives have attracted considerable attention of organic and medicinal chemists for many years.¹⁻⁴ The structural core of quinoline is frequently associated with medicinal applications, such as anti-cancer,⁵ antimicrobial,⁶ HIV-1 integrase inhibitors,⁷ HIV protease inhibitors⁸ antileishmanial activity,⁹ NK-3 receptor antagonists¹⁰ and pLT antagonists.¹¹⁻¹³

Quinoline carboxamides are bioisosteres of the NK-3 antagonist SB 218795 and were reported to possess local anesthetic, potent cholinesterase inhibitor activities¹⁴ and potent and orally active PDE4 inhibitor.¹⁵

The intensive efforts to find effective therapeutic agents with antiviral and antitumor activities have directed many researchers to synthesize a series of modified nucleosides or their analogues. Among the enormous number of published articles on modified heterocyclic nucleosides there are only a few publications that report the amino acid coupling of heterocyclic nucleosides.¹⁶⁻¹⁸

This type of modified amino acid nucleosides brings the potential of simultaneous binding and recognition of two complementary structure residues; amino acid and heterocyclic ring, which might interact with the poly nucleotides of the receptor recognition sites.

The aim of the present study was to synthesize a series of quinoline nucleosides in the ribo-series substituted at position 4 by a spacer linked with a series of amino acids and dipeptides carboxamides as potential chemotherapeutic agents.

Result and Discussion

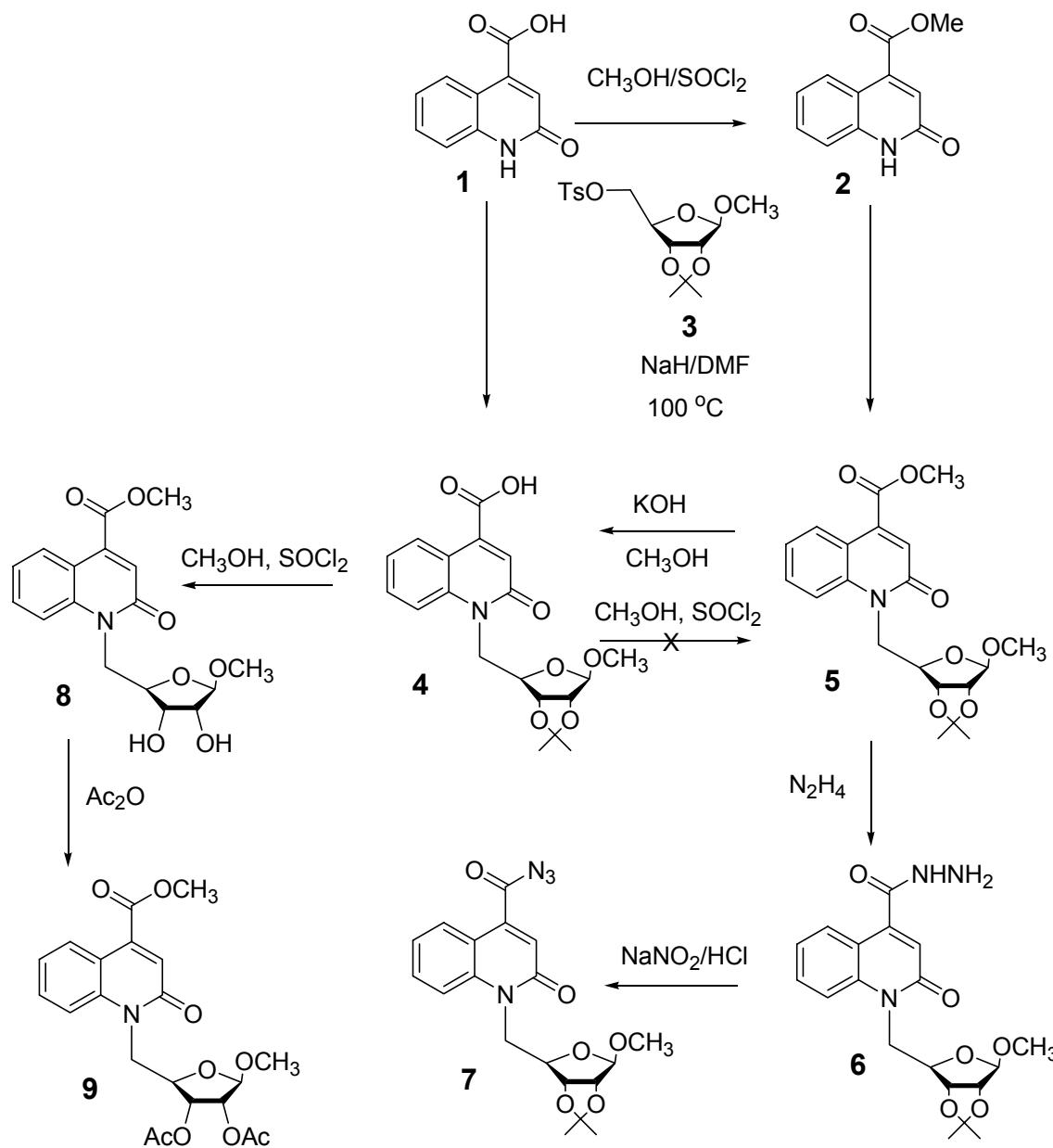
In view of these facts and in continuation of our efforts in synthesizing various bioactive molecules,¹⁹⁻²³ we have found it desirable to synthesize a series of quinoline nucleoside bearing amino acid methyl esters and dipeptides at position 4.

Treatment of quinoline **1** and **2**²⁴ with protected ribose tosylate **3** in the presence of NaH in dry DMF afforded methyl 1-(methyl 5-deoxy-2,3-*O*-isopropylidene-β-D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydroquinoline-4-carboxylate (**4**) and acid **5**, respectively in 85-90 % yields according to a slightly modified procedure by the displacement of the tosyloxy group.^{25,26}

The structure of reverse nucleoside **5** was chemically confirmed by an equivocal synthesis from the ester derivative **4** by hydrolysis in aqueous alcoholic KOH. On the other hand, the attempted esterification of **5** with thionyl chloride in absolute methanol afforded fully deprotected “reversed” nucleoside ester **8** in 60 % yield. The subsequent acetylation of **8** gave diacetyl derivative **9** (Scheme 1).

Carboxamides play a key role in medicinal chemistry. They are neutral, stable, have both hydrogen bond donating and accepting properties necessary for biological molecule recognition.²⁷

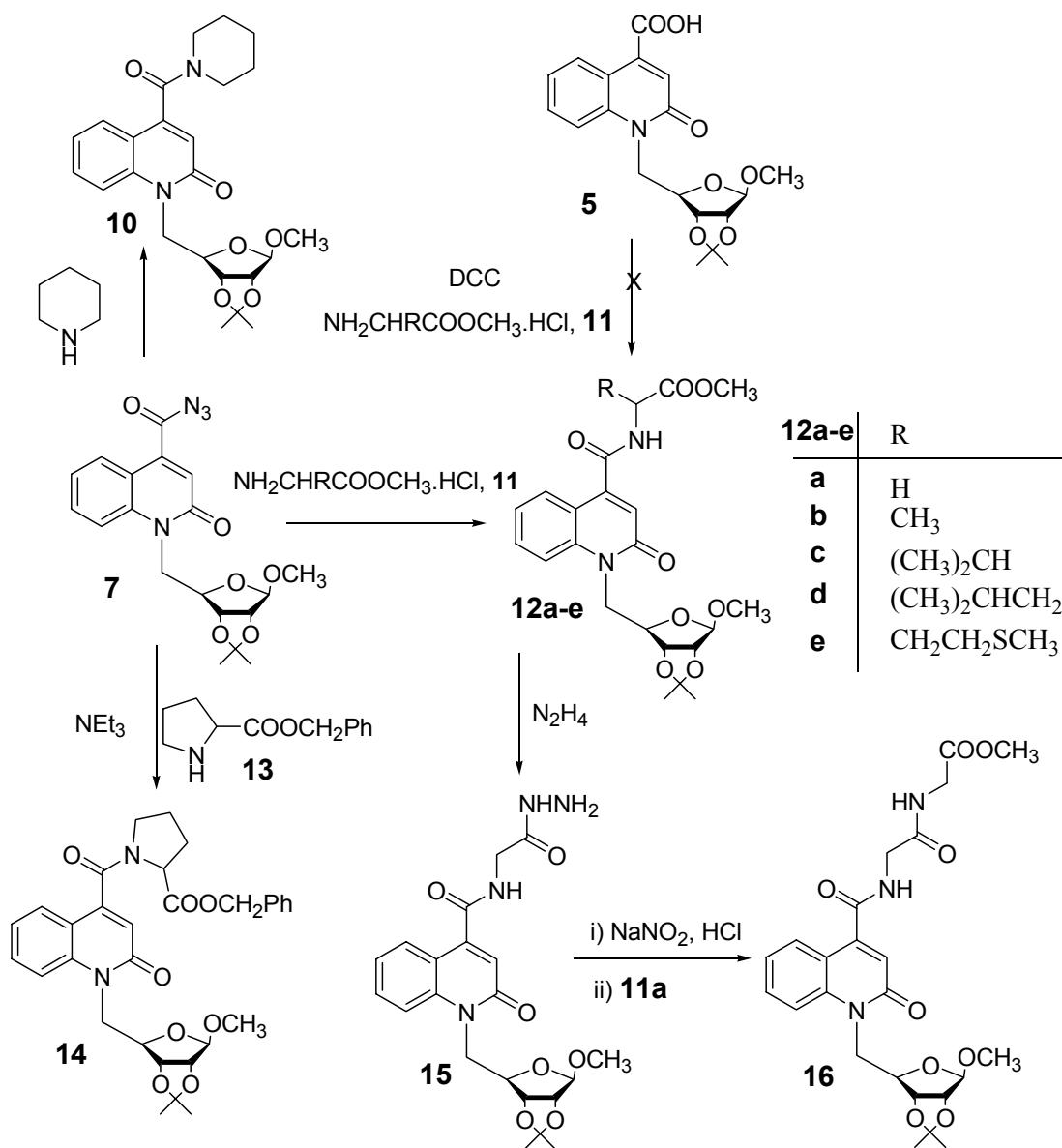
Both ester **4** and acid **5** are excellent key intermediates for the simple chemical modification of the quinoline skeleton to produce a carboxamide moiety at position 4. The ester **4** was boiled with hydrazine hydrate in ethyl alcohol to afford the hydrazide **6**, which was subsequently converted into azide **7** by treatment with NaNO₂ and HCl mixture.

**Scheme 1**

The synthesis of the target carboxamide **10** and amino acid derivatives **12a-e**, **14** was efficiently performed from key intermediate ester **4** via the azide coupling method,^{22, 28} which was reported to minimize the degree of racemization in amino acid coupling. The azide **7** reacted with piperidine in ethyl acetate to give the corresponding 1-(methyl 5-deoxy-2,3-*O*-isopropylidene- β -D-ribofuranosyl-5-yl)-2-oxo-4-(piperidine-1-carbonyl) 1,2-dihydroquinoline (**10**) in 77 % yield.

Similarly, the azide **7** reacted with amino acid methyl ester hydrochloride **11a-e** (glycine, L-alanine, L-valine, L-leucine, L-methionine) and **13** in the presence of triethyl amine in ethyl

acetate to afford methyl-2-{{[1-(methyl 5-deoxy-2,3-O-isopropylidene- β -D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} alkanoate **12a-e** and **14** in good yield.



Scheme 2

The DCC coupling is one of the major tools employed in literature to introduce peptide bonds by the reaction of acid with amino acid methyl ester. The synthesis of the target amino acid derivatives **12a-e** from the corresponding acid **5** via DCC coupling could be valuable for chemical structure confirmation. However, the reaction of acid **5** with glycine methyl ester hydrochloride **11a** in the presence of DCC, HOBr and triethyl amine failed to give the desired product **12a**.

Further development of azide coupling was obtained by the synthesis of the dipeptide derivative **16**. Hydrazinolysis of the amino acid ester **12a** (Gly) afforded the hydrazide **15**. Nitrosation of hydrazide **15** gave the azide. The *in situ* generated azide solution in ethyl acetate reacted with glycine methyl ester hydrochloride **11a**, in the presence of triethyl amine to afford the dipeptide **16** (Scheme 2).

The structure assignment of the amino acid esters **12a-e** and **16** is based on spectral analysis, (Figure 1). The ¹H NMR spectrum of the glycine derivative **12a** exhibits three singlet signals at δ 4.99, 3.75 and 3.31 ppm corresponding to H-1 and 2OCH₃ respectively. The ¹³C NMR spectrum of **12a** shows 107.8, 54.9, 41.5 ppm attributed to (C-1), OCH₃ and NCH₂ respectively. The ¹H NMR for all amino acid derivatives **12b** (Ala), **12c** (Val), **12d** (Leu), **12e** (Met), gives triplet or doublet centered at δ 6.60 ppm attributed to NH.

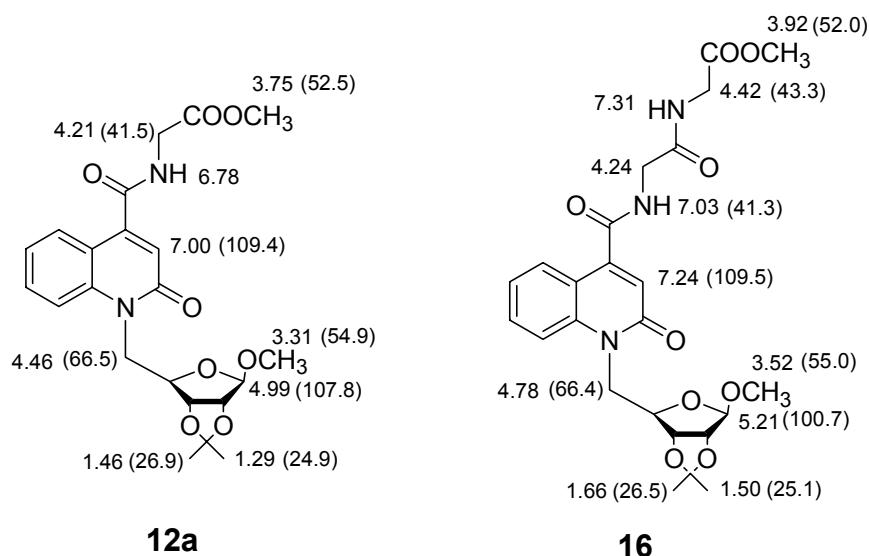


Figure 1. Selected ¹H NMR and ¹³C NMR spectral data of amino acid derivative **12a** and dipeptide **16**.

Conclusions

In conclusion, an efficient and very simple method for the syntheses of various quinoline reverse nucleosides bearing an amino acid ester residue **12-16** were prepared by azide coupling method from ester **4** in high yields as potential chemotherapeutic agents.

Experimental Section

General Procedures. Solvents were purified and dried in the usual way. The boiling range of the petroleum ether used was 40-60°C. Thin layer chromatography (TLC): silica gel 60 F₂₅₄

plastic plates (E. Merck, layer thickness 0.2 mm) detected by UV absorption. Melting points were determined on a Buchi 510 melting-point apparatus and the values are uncorrected. NMR spectra measured with Bruker AC 250 (250 MHz). TMS (0.00 ppm) as internal standard. MALDI-MS were measured with a KRATOS Analytical Compact, using 2,5 dihydroxybenzoic acid (DHB) as matrix. The $(M+Na)^+$ and $(M+K)^+$ ions were peak matched using ions derived from the 2,5-dihydroxybenzoic acid matrix. Elemental analyses were performed on a Flash EA-1112 instrument at the Microanalytical laboratory, Faculty of Science, Suez Canal University, Ismailia, Egypt. The starting compounds **1**, **2**²⁶ and **3**^{24,25} were prepared according to the method described.

Preparation of compound 4

A mixture of quinoline **1** (2.0 g, 10.0 mmol) and NaH (0.24 g, 10.0 mmol) in dry DMF (50 ml) was stirred at 100°C for 1 h. The sugar derivative **3** (3.42 g, 10.0 mmol) was added and the mixture was stirred at 100°C for 5 h. The solution was evaporated to dryness. The residue was purified by column chromatography petroleum ether/ethyl acetate (8:1) as eluent to give **4**.

Methyl 1-(methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydroquinoline-4-carboxylate (4). White powder (3.50 g, 90 %); mp 156-157 °C. ¹H NMR (250 MHz, CDCl₃): δ 8.51 (1H, d, *J* = 7.7 Hz, ArH); 7.75 (1H, d, *J* = 7.7 Hz, ArH); 7.62-7.58 (3H, m, ArH, CH); 4.97 (1H, s, H-1); 4.76 (1H, d, *J* = 5.9 Hz, H-2); 4.61 (1H, d, *J* = 5.9 Hz, H-3); 4.52-4.47 (1H, m, H-4); 4.43-4.37 (2H, m, H-5, H-5'); 3.88 (3H, s, OCH₃); 3.28 (3H, s, OCH₃); 1.44 (3H, s, CH₃); 1.27 (3H, s, CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 166.0; 160.0 (2CO); 147.3; 137.9; 129.8; 127.7; 125.5; 125.3; 122.0; 115.2 (Ar-C); 112.6 (CMe₂); 109.4 (C-1); 85.2 (C-2); 84.4 (C-3); 82.0 (C-4); 66.3 (C-5); 54.8; 52.5 (2OCH₃); 26.4; 24.9 (2CH₃). (MALDI, positive mode, matrix: DHB): m/z = 388.8 (M)⁺; 411.3 (M+Na)⁺. Anal. Calcd. For C₂₀H₂₃NO₇ (389.40): C, 61.69 %, H, 5.95 %, N, 3.60 %; found C, 61.42 %, H, 5.88 %, N, 3.42 %.

Preparation of compound 5

Method A. A mixture of quinoline **2** (1.90 g, 10.0 mmol) and NaH (0.48 g, 20.0 mmol) in dry DMF (50 ml) was stirred at 100°C for 1 h. The sugar derivative **3** (3.42 g, 10.0 mmol) was added and the mixture was stirred at 100°C for 5 h. The solution was evaporated to dryness and the residue was acidified by dil HCl and extracted with CHCl₃ (100 mL). The combined organic extracts were dried over Na₂SO₄, filtered and evaporated to dryness and the residue was chromatographed using petroleum ether/ethyl acetate (6:1) as eluent.

1-(Methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydroquinoline-4-carboxylic acid (5). White crystals (3.20 g, 85 %); mp 212-213 °C. ¹H NMR (250 MHz, CDCl₃): δ 12.83 (1H, bs, OH); 8.36 (1H, d, *J* = 8.2 Hz, ArH); 7.60-7.45 (2H, m, ArH); 7.28-7.19 (2H, m, ArH, CH); 5.01 (1H, s, H-1); 4.74 (1H, d, *J* = 5.8 Hz, H-2); 4.63 (1H, d, *J* = 5.8 Hz, H-3); 4.54-4.49 (1H, m, H-4); 4.44-4.41 (2H, m, H-5, H-5'); 3.34 (3H, s, OCH₃); 1.48 (3H, s, CH₃); 1.32 (3H, s, CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 164.6, 163.5 (2CO); 140.2;

138.9; 138.7; 131.2; 126.4; 124.4; 123.4; 112.3 (Ar-C); 112.8 (CMe₂); 109.6 (C-1); 85.2 (C-2); 84.0 (C-3); 81.7 (C-4); 66.1 (C-5); 55.1 (OCH₃); 26.4; 24.9 (2CH₃). (MALDI, positive mode, matrix: DHB): m/z = 397.6 (M+Na)⁺. Anal. Calcd. For C₁₉H₂₁NO₇ (375.37): C, 60.79 %, H, 5.64 %, N, 3.73 %; found C, 61.02 %, H, 5.35 %, N, 4.11 %.

Method B. A solution of **4** (1.95 g, 5.0 mmol) in methanol (40 mL) and 5% KOH (6 mL) was stirred for 6 h. The solution was evaporated to dryness and the residue was dissolved in water and acidified with dil HCl. The white precipitate was filtered and chromatographed using petroleum ether/ethyl acetate (6:1) to give **5** (1.20 g, 64 %).

Preparation of hydrazide **6**

To a solution of ester **4** (3.90 g, 10.0 mmol) in methyl alcohol (50 mL) was added hydrazine hydrate (2.4 mL, 50.0 mmol). The reaction mixture was refluxed for 4 hours, cooled and the resultant precipitate was filtered off, washed with ethanol and ether then crystallized from methanol to yield the hydrazide **6**.

1-(Methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydro-quinoline-4-carboxylic acid hydrazide (6). White crystals (3.40 g, 87 %); mp 197-198 °C. ¹H NMR (250 MHz, CDCl₃): δ 8.05 (1H, d, J = 7.1 Hz, ArH); 7.72 (1H, d, J = 7.1 Hz, ArH); 7.62-7.32 (2H, m, ArH); 6.95 (1H, s, CH); 4.97 (1H, s, H-1); 4.58 (1H, d, J = 6.0 Hz, H-2); 4.56-4.45 (4H, m, H-3, H-4, H-5, H-5'); 3.88 (3H, s, OCH₃); 3.20 (2H, bs, NH₂); 1.43 (3H, s, CH₃); 1.25 (3H, s, CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 167.6, 160.5 (2CO); 143.2; 134.2; 130.2; 127.7; 125.1; 121.7; 112.5; 112.1 (Ar-C); 111.4 (CMe₂); 109.4 (C-1); 85.2 (C-2); 84.5 (C-3); 82.0 (C-4); 66.3 (C-5); 54.9 (OCH₃); 26.4; 24.9 (2CH₃). Anal. Calcd. For C₁₉H₂₃N₃O₆ (389.40): C, 58.60 %, H, 5.95 %, N, 10.79 %; found C, 58.46 %, H, 5.81 %, N, 10.92 %.

Preparation of azide **7**

To a cold solution (-5°C) of hydrazide derivative **6** (3.89 g, 10.0 mmol) in HOAc (10 ml), 1 N HCl (30 ml), and water (150 ml) was added a solution of NaNO₂ (1.0 g, 15.0 mmol) in cold water (10 ml). After stirring at -5°C for 15 min, the yellow syrup was formed. The azide was taken in cold ethyl acetate (100 ml), washed with cold 3% NaHCO₃, H₂O and finally dried (Na₂SO₄). The solvent was evaporated in vacuo to give the pure azide **7**.

1-(Methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydro-quinoline-4-carbonyl azide (7). Yellow powder (2.60 g, 65 %); mp 68-69 °C. ¹H NMR (250 MHz, CDCl₃): δ 7.89-7.60 (2H, m, ArH); 7.51-7.45 (2H, m, ArH); 6.71 (1H, s, CH); 5.03 (1H, s, H-1); 4.81 (1H, d, J = 5.8 Hz, H-2); 4.65 (1H, d, J = 5.8 Hz, H-3); 4.62-4.50 (3H, m, H-4, H-5, H-5'); 3.34 (3H, s, OCH₃); 1.48 (3H, s, CH₃); 1.32 (3H, s, CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 178.5, 160.6 (2CO); 144.5; 136.2; 130.6; 127.7; 124.7; 122.3; 115.7 (Ar-C); 111.4 (CMe₂); 109.4 (CH); 107.8 (C-1); 85.3 (C-2); 84.5 (C-3); 82.1 (C-4); 66.3 (C-5); 54.9 (OCH₃); 26.5, 25.0 (2CH₃). (MALDI, positive mode, matrix: DHB): m/z = 422.1 (M+Na)⁺. Anal. Calcd. For C₁₉H₂₀N₄O₆ (400.39): C, 57.00 %, H, 5.03 %, N, 13.99 %; found C, 56.74 %, H, 5.31 %, N, 14.17 %.

Preparation of compound 8

To a solution of **5** (3.75 g, 10.0 mmol) in absolute methanol (70 ml) at 0°C; thionyl chloride (0.8 ml, 10.0 mmol) was added dropwise under vigorous stirring for 3 hours at 5°C. The reaction mixture was kept overnight at room temperature, then the solvent was evaporated under reduced pressure, afterwards the residue was treated with absolute methanol (20 ml) with subsequent evaporation. This process was repeated three times to remove excess HCl. The residue was chromatographed using petroleum ether/ethyl acetate (3:1) as eluent.

Methyl 1-(methyl 5-deoxy- β -D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydroquinoline-4-carboxylate (8). White crystals (2.10 g, 60 %); mp 187-188 °C. ^1H NMR (250 MHz, DMSO): δ 8.85 (1H, d, J = 8.1 Hz, ArH); 8.0 (1H, d, J = 8.1 Hz, ArH); 7.81 (1H, s, CH); 7.75-7.52 (2H, m, ArH); 5.41-5.34 (1H, m, H-2); 5.15-5.05 (1H, m, H-3); 4.95 (1H, s, H-1); 4.66-4.58 (2H, m, H-4, H-5); 4.41-4.38 (1H, m, H-5'); 3.62 (3H, s OCH₃); 3.38 (3H, s, OCH₃); 2.15 (2H, bs, 2OH). ^{13}C NMR (62.5 MHz, DMSO): δ 174.1; 164.6 (2CO); 152.2; 142.8; 138.3; 131.2; 128.7; 126.5; 122.2; 115.9 (Ar-C); 107.0 (C-1); 80.3 (C-2); 76.1 (C-3); 71.3 (C-4); 66.0 (C-5); 54.0 (OCH₃); 53.6 (OCH₃). (MALDI, positive mode, matrix: DHB): m/z = 371.3 (M+Na)⁺. Anal. Calcd. For C₁₇H₁₉NO₇ (349.34): C, 58.45 %, H, 5.48 %, N, 4.01 %; found C, 58.02 %, H, 5.39 %, N, 3.79 %.

Acetylation of compound 8

Quinoline **8** (0.70 g, 2.0 mmol) was treated with acetic anhydride (20 mL) and pyridine (20 mL). The reaction mixture was stirred at room temperature for 24 h.; concentrated under reduced pressure, and purified by flash chromatography (petroleum ether/ethyl acetate, 8:1).

Methyl 1-(methyl 5-deoxy-2,3-di-O-acetyl- β -D-ribofuranosyl-5-yl)-2-oxo-1,2-dihydroquinoline-4-carboxylate (9). White powder (0.60 g, 69 %); mp 161-162 °C. ^1H NMR (250 MHz, CDCl₃): δ 7.56-7.43 (2H, m, ArH); 7.29 (1H, s, CH); 7.28-7.21 (2H, m, ArH); 5.43-5.32 (1H, m, H-2); 5.25 (1H, d, J = 5.0 Hz, H-3); 4.91 (1H, s, H-1); 4.64-4.58 (1H, m, H-4); 4.47-4.42 (2H, m, H-5, H-5'); 3.63 (3H, s, OCH₃); 3.36 (3H, s, OCH₃); 2.10 (3H, s, CH₃); 2.04 (3H, s, CH₃). ^{13}C NMR (62.5 MHz, CDCl₃): δ 174.1; 170.6; 170.1; 168.2 (4CO); 153.1; 141.5; 139.1; 132.4; 129.2; 127.2; 124.2 (Ar-C); 116.4 (CMe₂); 106.5 (C-1); 81.2 (C-2); 76.6 (C-3); 70.8 (C-4); 66.4 (C-5); 54.2 (OCH₃); 54.4 (OCH₃); 20.5; 20.2 (2AcO). (MALDI, positive mode, matrix: DHB): m/z = 454.7 (M+Na)⁺. Anal. Calcd. For C₂₁H₂₃NO₉ (433.41): C, 58.20 %, H, 5.35 %, N, 3.23 %; found C, 58.42 %, H, 5.41 %, N, 3.52 %.

General procedure for preparation of Compounds 10, 12a-e, 14

To azide solution **7** (4.0 g, 10.0 mmol) in ethyl acetate (50 ml), the appropriate nucleophile [piperidine; amino acid ester hydrochloride **11a-e** or proline ester hydrochloride **13** (10.0 mmol)] in ethyl acetate (50 ml) containing 2.0 ml of triethyl amine was added. The reaction mixture was kept at -5°C for 24 h, then at 25°C for another 24 h, followed by washing with 0.5 N HCl, water, 3 % solution of NaHCO₃ and finally dried (Na₂SO₄). The solution was evaporated to dryness, and the residue was recrystallized from petroleum ether/ ethyl acetate to give the desired product.

1-(Methyl 5-deoxy-2,3-*O*-isopropylidene- β -D-ribofuranosyl-5-yl)-2-oxo-4-(piperidine-1-carbonyl) -1,2-dihydroquinoline (10). White powder (3.40 g, 77 %); mp 171-172 °C. ^1H NMR (250 MHz, CDCl_3): δ 7.80 (1H, d, $J = 8.3$ Hz, ArH); 7.66-7.57 (2H, m, ArH); 7.38-7.32 (1H, m, ArH); 6.82 (1H, s, CH); 5.00 (1H, s, H-1); 4.80 (1H, d, $J = 6.0$ Hz, H-2); 4.63 (1H, d, $J = 6.0$ Hz, H-3); 4.59-4.51 (2H, m, H-4, H-5); 4.47-4.42 (1H, m, H-5'); 3.62 (3H, s, OCH_3); 3.41 (4H, m, 2NCH_2); 1.67-1.61 (6H, m, 3CH_2); 1.47 (3H, s, CH_3); 1.30 (3H, s, CH_3). ^{13}C NMR (62.5 MHz, CDCl_3): δ 166.2, 160.7 (2CO); 146.5; 145.8; 130.0; 129.8; 127.6; 124.7; 121.4 (Ar-C); 112.3 (CMe_2); 109.3 (CH); 100.9 (C-1); 85.1 (C-2); 84.4 (C-3); 81.9 (C-4); 66.2 (C-5); 54.7 (OCH_3); 47.9 (CH_2); 42.4 (CH_2); 33.8 (2 CH_2); 42.4 (CH_2); 26.4; 24.2 (2 CH_3). (MALDI, positive mode, matrix: DHB): $m/z = 464.5$ ($\text{M}+\text{Na}^+$). Anal. Calcd. For $\text{C}_{24}\text{H}_{30}\text{N}_2\text{O}_6$ (442.50): C, 65.14 %, H, 6.83 %, N, 6.33 %; found C, 65.40 %, H, 7.15 %, N, 6.07 %.

Methyl-2-{{[1-(methyl 5-deoxy-2,3-*O*-isopropylidene- β -D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} acetate (12a). White powder (3.60 g, 81 %); mp 148-149 °C. ^1H NMR (250 MHz, CDCl_3): δ 8.05 (1H, d, $J = 7.3$ Hz, ArH); 7.77 (1H, d, $J = 7.3$ Hz, ArH); 7.63-7.57 (1H, m, ArH); 7.41-7.32 (1H, m, ArH); 7.00 (1H, s, CH); 6.78 (1H, t, $J = 5.4$ Hz, NH); 4.99 (1H, s, H-1); 4.79 (1H, d, $J = 6.0$ Hz, H-2); 4.61 (1H, d, $J = 6.0$ Hz, H-3); 4.57-4.48 (1H, m, H-4); 4.46-4.40 (2H, m, H-5, H-5'); 4.21 (2H, d, $J = 5.4$ Hz, NHCH_2); 3.75 (3H, s, OCH_3); 3.31 (3H, s, OCH_3); 1.46 (3H, s, CH_3); 1.29 (3H, s, CH_3). ^{13}C NMR (62.5 MHz, CDCl_3): δ 169.9, 167.1, 160.5 (3CO); 146.9; 144.2; 130.2; 128.9; 127.7; 125.2; 121.5 (Ar-C); 111.2 (CMe_2); 109.4 (CH); 107.8 (C-1); 85.2 (C-2); 84.5 (C-3); 82.0 (C-4); 66.3 (C-5); 54.9 (OCH_3); 52.5 (OCH_3); 41.5 (NHCH_2); 26.9; 24.9 (2 CH_3). (MALDI, positive mode, matrix: DHB): $m/z = 468.0$ ($\text{M}+\text{Na}^+$). Anal. Calcd. For $\text{C}_{22}\text{H}_{26}\text{N}_2\text{O}_8$ (446.45): C, 59.19 %, H, 5.87 %, N, 6.27 %; found C, 58.87 %, H, 5.92 %, N, 6.41 %.

Methyl-2-{{[1-(methyl 5-deoxy-2,3-*O*-isopropylidene- β -D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} propanoate (12b). White powder (3.50 g, 76 %); mp 162-163 °C. ^1H NMR (250 MHz, CDCl_3): δ 8.11 (1H, s, CH); 7.73-7.68 (2H, m, ArH); 7.48-7.42 (1H, m, ArH); 7.18-7.11 (1H, m, ArH); 6.60 (1H, d, $J = 7.6$ Hz, NH); 4.98 (1H, s, H-1); 4.79 (1H, d, $J = 6.0$ Hz, H-2); 4.62 (1H, d, $J = 6.0$ Hz, H-3); 4.59-4.47 (2H, m, H-4, H-5); 4.42-4.34 (2H, m, NHCH_2 , H-5'); 3.68 (3H, s, OCH_3); 3.30 (3H, s, OCH_3); 1.46 (3H, s, CH_3); 1.40 (3H, d, $J = 7.2$ Hz, CH_3); 1.29 (3H, s, CH_3). ^{13}C NMR (62.5 MHz, CDCl_3): δ 174.8; 162.3; 154.7 (3CO); 146.9; 143.7; 131.4; 129.4; 128.0; 123.5; 119.9 (Ar-C); 112.3 (CMe_2); 109.4 (CH); 100.7 (C-1); 85.3 (C-2); 84.5 (C-3); 82.1 (C-4); 65.7 (C-5); 54.8 (OCH_3); 52.6 (OCH_3); 48.9 (NHCH_2), 26.4; 24.9 (2 CH_3); 18.6 (CH_3). (MALDI, positive mode, matrix: DHB): $m/z = 482.6$ ($\text{M}+\text{Na}^+$). Anal. Calcd. For $\text{C}_{23}\text{H}_{28}\text{N}_2\text{O}_8$ (460.48): C, 59.99 %, H, 6.13 %, N, 6.08 %; found C, 60.12 %, H, 5.95 %, N, 6.41 %.

Methyl-2-{{[1-(methyl 5-deoxy-2,3-*O*-isopropylidene- β -D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} 3-methylbutanoate (12c). White powder (3.10 g, 63 %); mp 129-130 °C. ^1H NMR (250 MHz, CDCl_3): δ 7.78-7.65 (2H, m, ArH); 7.52 (1H, s, CH); 7.47-7.41 (2H, m, ArH); 6.68 (1H, d, $J = 8.5$ Hz, NH); 4.96 (1H, s, H-1); 4.78 (1H, d, $J = 6.1$ Hz, H-2); 4.59 (1H, d, $J = 6.1$ Hz, H-3); 4.56-4.48 (2H, m, H-4, H-5); 4.43-4.34 (2H, m, H-5').

NHCH); 3.63 (3H, s, OCH₃); 3.28 (3H, s, OCH₃); 2.14-2.03 (1H, m, CH); 1.45 (3H, s, CH₃); 1.28 (3H, s, CH₃); 0.93 (3H, d, *J* = 6.8 Hz, CH₃); 0.83 (3H, d, *J* = 6.8 Hz, CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 173.8; 162.5; 155.4 (3CO); 146.9; 144.1; 133.4; 129.5; 128.0; 123.5; 120.2 (Ar-C); 111.1 (CMe₂); 109.5 (CH); 100.5 (C-1); 85.4 (C-2); 84.6 (C-3); 82.2 (C-4); 65.8 (C-5); 58.4 (NHCH); 54.9 (OCH₃); 52.3 (OCH₃); 31.3 (CH); 26.5, 25.0 (2CH₃); 19.1; 17.9 (2CH₃). (MALDI, positive mode, matrix: DHB): m/z = 511.0 (M+Na)⁺; 526.0 (M+K)⁺. Anal. Calcd. For C₂₅H₃₂N₂O₈ (488.53): C, 61.46 %, H, 6.60 %, N, 5.73 %; found C, 61.12 %, H, 6.43 %, N, 5.86 %.

Methyl-2-{{[1-(methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} 4-methylpentanoate (12d). White powder (3.70 g, 73 %); mp 107-108 °C. ¹H NMR (250 MHz, CDCl₃): δ 8.00-7.76 (1H, m, ArH); 7.73-7.69 (2H, m, ArH); 7.51-7.45 (1H, m, ArH); 7.42 (1H, s, CH); 6.46 (1H, d, *J* = 8.1 Hz, NH); 4.98 (1H, s, H-1); 4.77 (1H, d, *J* = 5.9 Hz, H-2); 4.59 (1H, d, *J* = 5.9 Hz, H-3); 4.55-4.44 (2H, m, H-4, H-5); 4.41-4.33 (2H, m, H-5', NHCH); 3.73 (3H, s, OCH₃); 3.30 (3H, s, OCH₃); 1.79-1.56 (2H, m, CH₂); 1.47 (3H, s, CH₃); 1.29 (3H, s, CH₃); 1.26-1.22 (1H, m, CH); 3.93 (6H, 2d, *J* = 3.8 Hz, 2CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 175.6; 162.2; 154.7 (3CO); 146.7; 143.4; 130.0; 129.2; 123.3; 119.6; 117.9 (Ar-C); 112.2 (CMe₂); 109.3 (CH); 100.3 (C-1); 85.1 (C-2); 84.4 (C-3); 82.0 (C-4); 65.5 (C-5); 54.7 (OCH₃); 52.5 (OCH₃); 51.6 (NHCH); 41.4 (CH); 26.3 (CH₃); 25.0 (CH₂); 24.8 (CH₃); 22.6 (CH₃); 21.7 (CH₃). (MALDI, positive mode, matrix: DHB): m/z = 525.7 (M+Na)⁺; 540.6 (M+K)⁺. Anal. Calcd. For C₂₆H₃₄N₂O₈ (502.56): C, 62.14 %, H, 6.82 %, N, 5.57 %; found C, 62.43 %, H, 7.10 %, N, 5.41 %.

Methyl-2-{{[1-(methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} 4-methylsulfanylbutanoate (12e). White powder (2.80 g, 54 %); mp 81-82 °C. ¹H NMR (250 MHz, CDCl₃): δ 8.12 (1H, s, CH); 7.77-7.71 (2H, m, ArH); 7.55-7.47 (1H, m, ArH); 7.25-7.17 (1H, m, ArH); 6.68 (1H, d, *J* = 8.0 Hz, NH); 4.99 (1H, s, H-1); 4.80 (1H, d, *J* = 6.0 Hz, H-2); 4.62 (1H, d, *J* = 6.0 Hz, H-3); 4.58-4.50 (2H, m, H-4, H-5); 4.46-4.33 (2H, m, H-5', NHCH); 3.69 (3H, s, OCH₃); 3.32 (3H, s, OCH₃); 2.51 (1H, t, *J* = 7.3 Hz, CH₂); 2.18-2.05 (2H, m, CH₂S); 2.01 (3H, s, SCH₃); 1.47 (3H, s, CH₃); 1.30 (3H, s, CH₃). ¹³C NMR (62.5 MHz, CDCl₃): δ 173.7, 162.3, 154.8 (3CO); 146.9; 143.6; 133.5; 129.5; 128.0; 123.5; 119.9 (Ar-C); 112.3 (CMe₂); 109.4 (CH); 100.9 (C-1); 85.2 (C-2); 84.4 (C-3); 82.1 (C-4); 65.7 (C-5); 54.6 (NHCH); 52.6 (OCH₃); 52.4 (OCH₃); 31.7 (CH₂); 29.9 (CH₂); 26.4 (CH₃); 24.9 (CH₃); 15.3 (SCH₃). (MALDI, positive mode, matrix: DHB): m/z = 542.5 (M+Na)⁺. Anal. Calcd. For C₂₅H₃₂N₂O₈S (520.60): C, 57.68 %, H, 6.20 %, N, 5.38 %; found C, 57.52 %, H, 5.95 %, N, 5.21 %.

Benzyl 1-{{[1-(methyl 5-deoxy-2,3-O-isopropylidene-β-D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carbonyl}pyrrolidin-2-carboxylate (14). White powder (3.80 g, 67 %); mp 68-69 °C. ¹H NMR (250 MHz, CDCl₃): δ 7.80 (1H, d, *J* = 8.4 Hz, ArH); 7.71 (1H, s, CH); 7.61-7.54 (4H, m, ArH); 7.34-7.28 (4H, m, ArH); 5.28 (1H, d, *J_{gem}* = 12.3 Hz, CHph); 5.21 (1H, d, *J_{gem}* = 12.3 Hz, CHph); 5.01 (1H, s, H-1); 4.83 (1H, d, *J* = 6.0 Hz, H-2); 4.63 (1H, d, *J* = 6.0 Hz, H-3); 4.62-4.51 (2H, m, H-4, H-5); 4.44-4.36 (2H, m, H-5', NHCH); 3.86-3.60 (2H, m,

CH_2); 3.34 (3H, s, OCH_3); 2.23-2.01 (4H, m, 2CH_2); 1.48 (3H, s, CH_3); 1.31 (3H, s, CH_3). ^{13}C NMR (62.5 MHz, CDCl_3): δ 177.7; 172.4; 162.6 (3CO); 153.2; 146.9; 143.3; 135.2; 129.3; 128.6; 128.5; 128.2; 123.6; 118.9; 117.9 (Ar-C); 112.3 (CMe_2); 109.4 (CH); 100.3 (C-1); 85.3 (C-2); 84.5 (C-3); 82.1 (C-4); 67.4 (C-5); 65.7 (CH_2Ph); 59.6 (NCH); 54.8 (OCH_3); 46.6 (CH_2); 29.5 (CH_2); 26.4 (CH_3); 24.9 (CH_3); 24.3 (CH_2). (MALDI, positive mode, matrix: DHB): m/z = 584.4 ($\text{M}+\text{Na}^+$). Anal. Calcd. For $\text{C}_{31}\text{H}_{34}\text{N}_2\text{O}_8$ (562.23): C, 66.18 %, H, 6.09 %, N, 4.98 %; found C, 65.92 %, H, 5.85 %, N, 5.26 %.

General procedure for preparation of hydrazide 15

The hydrazinolysis of **12a** was processed as described above and the product **15** was purified by crystallisation from ethanol.

2-{{[1-(methyl 5-deoxy-2,3-O-isopropylidene- β -D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido} acetohydrazide (15)}. White powder (3.20 g, 72 %); mp 101-103 °C. ^1H NMR (250 MHz, CDCl_3): δ 8.23 (1H, d, J = 7.5 Hz, ArH); 7.79 (2H, d, J = 7.5 Hz, ArH); 7.71-7.59 (2H, m, ArH); 7.01 (1H, s, CH); 6.85 (1H, bs, NH); 5.05 (1H, s, H-1); 4.61-4.54 (2H, m, H-2, H-3); 4.50-4.43 (3H, m, H-4, H-5, H-5'); 4.13 (2H, d, J = 5.7 Hz, NHCH_2); 3.85 (3H, s, OCH_3); 3.50 (2H, bs, NH₂); 1.47 (3H, s, CH_3); 1.21 (3H, s, CH_3). (MALDI, positive mode, matrix: DHB): m/z = 468.3 ($\text{M}+\text{Na}^+$). Anal. Calcd. For $\text{C}_{21}\text{H}_{26}\text{N}_4\text{O}_7$ (446.45): C, 56.50 %, H, 5.87 %, N, 12.55 %; found C, 56.72 %, H, 5.98 %, N, 12.

General procedure for preparation of dipeptide 16

The azide coupling was processed as described above and the product **16** was purified by crystallisation from petroleum ether/ethyl acetate.

Methyl-2-{{[1-(methyl 5-deoxy-2,3-O-isopropylidene- β -D-ribofuranosyl-5-yl)]-2-oxo-1,2-dihydroquinoline-4-carboxamido}-2'acetamido} acetate (16)}. White powder (2.80 g, 56 %); mp 144-145 °C. ^1H NMR (250 MHz, CDCl_3): δ 8.28 (1H, d, J = 8.3 Hz, ArH); 8.00 (1H, d, J = 8.3 Hz, ArH); 7.84 (1H, dd, J = 1.4, 8.3 Hz, ArH); 7.60 (1H, dd, J = 1.4, 8.3 Hz, ArH); 7.31 (1H, bs, NH); 7.24 (1H, s, CH); 7.03 (1H, bs, NH); 5.21 (1H, s, H-1); 5.00 (1H, d, J = 6.0 Hz, H-2); 4.82 (1H, d, J = 6.0 Hz, H-3); 4.78-4.64 (3H, m, H-4, H-5, H-5'); 4.43 (2H, d, J = 5.3 Hz, NHCH_2); 4.24 (2H, d, J = 5.3 Hz, NHCH_2); 3.92 (3H, s, OCH_3); 3.52 (3H, s, OCH_3); 1.66 (3H, s, CH_3); 1.50 (3H, s, CH_3). ^{13}C NMR (62.5 MHz, CDCl_3): δ 170.0; 168.6; 167.5; 160.6 (4CO); 147.1; 144.2; 135.3; 130.3; 127.8; 125.3; 121.6 (Ar-C); 111.5 (CMe_2); 109.5 (CH); 100.7 (C-1); 85.4 (C-2); 84.6 (C-3); 82.2 (C-4); 66.4 (C-5); 55.0 (OCH_3); 52.5 (OCH_3); 43.3, 41.3 (2 NHCH_2), 26.5; 25.1 (2 CH_3). (MALDI, positive mode, matrix: DHB): m/z = 525.5 ($\text{M}+\text{Na}^+$). Anal. Calcd. For $\text{C}_{24}\text{H}_{29}\text{N}_3\text{O}_9$ (503.50): C, 57.25 %, H, 5.81 %, N, 8.35 %; found C, 56.93 %, H, 6.04 %, N, 8.12 %.

References

1. Balasubramanian, M.; Keay, J. G. In Katritzky, A. R.; Rees, C. W.; Scriven, E. F. V., Eds., *Comprehensive Heterocyclic Chemistry II*; Vol. 5, Pergamon: Oxford, 1996.
2. De, D.; Byers, L. D.; Krogstad, D. J. *J. Heterocycl. Chem.* **1997**, *34*, 315.
3. Gilchrist, T. *J. Chem. Soc., Perkin Trans. I* **2001**, 2491.
4. Kouznetsov, V.; Mendez, L.; Gomes, C. *Curr. Org. Chem.* **2005**, *9*, 141.
5. Elderfield, R. C.; Le Von , E. F. *J. Org. Chem.* **1960**, *25*, 1576.
6. Musiol, R.; Jampilek, J.; Buchta, V.; Silva, L.; Niedbala, H.; Podeszwa, B.; Palka, A.; Majerz-Maniecka, K.; Oleksyn, B.; Polanski, J. *Bioorg. Med. Chem.* **2006**, *14*, 3592.
7. Bénard, C.; Zouhiri, F.; Normand-Bayle, M.; Danet, M.; Desmaële, D.; Leh, H.; Mouscadet, J-F.; Mbemba, G.; Thomas, C-M.; Bonnenfant, S.; Le Bret, M.; d'Angelo, J. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 2473.
8. Garrouste, P.; Pawlowski, M. Tonnaire T.; Sicsic, S.; Dumy, P.; De Rosny, E.; Reboud-Ravaux, M.; Fulcrand, P.; Martinez, J. *Eur. J. Med. Chem.* **1998**, *33*, 423.
9. Desrivot, J.; Herrenknecht, C.; Ponchel, G.; Garbi, N.; Prina, E.; Fournet, A.; Bories, C.; Figadère, B.; Hocquemiller, R.; Loiseau, P. M. *Biomed. Pharmacother.* **2007**, *61*, 441.
10. Borioni, A.; Mustazza, C.; Sestili, I.; Sbraccia, M.; Turchetto, L.; Del Giudice, M. F. *Arch. Pharm.* **2007**, *340*, 17.
11. Sprecher, A.; Gerspacher, M.; Beck, A.; Kimmel, S.; Wiestner, H.; Anderson, G. P.; Niederhauser, U.; Subramanian, N.; Bray, M. A. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 965.
12. Galemmo, R. A.; Gavai, A.; Huang, F-C. *Curr. Opin. Ther. Patents* **1992**, *8*, 811.
13. Musser, J. H.; Kreft, A. F. *Drugs Fut.* **1990**, *15*, 73.
14. Miescher, K. *Helv. Chim. Acta* **1932**, *15*, 163.
15. Billah, M.; Buckley, G. M.; Cooper, N.; Dyke, H. J.; Egan, R.; Ganguly, A.; Gowers, L.; Haughan, A. F.; Kendall, H. J.; Lowe, C.; Minnicozzi, M.; Montana, J. G.; Oxford, J.; Peake, J. C.; Picken, C. L.; Piwinski, J. J.; Naylor, R.; Sabin, V.; Shih, N.-Y.; Warneck, J. B. H. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1617.
16. Zhang, D; Bender, D. M.; Victor, F.; Peterson, J. A.; Boyer, R. D.; Stephenson, G. A.; Azman, A.; McCarthy, J. R. *Tetrahedron Lett.* **2008**, *49*, 2052.
17. Threlfall, R.; Davies, A.; Howarth, N.; Richard Cosstick, R. *Nucleosides, Nucleotides & Nucleic acids* **2007**, *26*, 611.
18. Knapp, S. *Chem. Rev.* **1995**, *95*, 1859. (b) Casiraghi, G.; Zanardi, F.; Rassu, G.; Spanu, P. *Chem. Rev.* **1995**, *95*, 1677.
19. Ali, I. A. I.; Al-Masoudi, I. A.; Saeed, B.; Al-Masoudi, N. A.; La Colla, P. *Heteroatom Chem.* **2005**, *16*, 148.
20. Fathalla, W.; Ali, I. A. I. *Heteroatom Chem.* **2007**, *18*, 637.
21. Fathalla, W.; El Rayes, S. M.; Ali, I. A. I. *ARKIVOC* **2007**, (xvi), 173.
22. El Rayes, S. M.; Ali, I. A. I.; Fathalla, W. *ARKIVOC* **2008**, (xi), 86.
23. Ali, I. A. I.; Fathalla, W.; El Rayes, S. M. *ARKIVOC* **2008**, (iii), 179.

24. Okada, K.; Sakuma, H.; Kondo, M.; Inoue, S. *Chem. Lett.* **1979**, 213.
25. Abdel-Rahman, A. A.-H.; Abdel-Megied, A. E.-S.; Goda, A. E.-S.; Zeid, I. F.; El Ashry, E. S. H. *Nucleosides, Nucleotides & Nucleic acids* **2003**, 22, 2027.
26. Kissman, H. .M.; Baker, B. R. *J. Am. Chem. Soc.* **1957**, 79, 5534.
27. Ghose, A. K.; Viswanadhan, V. N.; Wendoloski, J. *J. Comb. Chem.* **1999**, 55.
28. Sahin, G.; Palaska, E.; Ekizoglu, M.; Ozalp, M. *Farmaco* **2002**, 57, 539.