Synthesis of the 5'-phosphonate of 4(S)-(6-amino-9H-purin-9-yl) tetrahydro-2(S)-furanmethanol [S,S-IsoddA]

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Abstract

4(*S*)-(6-Amino-9H-purin-9-yl)tetrahydrofuran-2(*S*)-ylmethyl phosphonic acid, a 5'-C-phosphonate analog of the potent anti-HIV compound *S*,*S*-IsoddA, was synthesized in order to bypass the critical initial intracellular phosphorylation. Key phases in this multistep synthesis were the Arbuzov reaction of the 5-iodofuranose with triethylphosphite and the Mitsunobu/coupling reaction of the phosphonate with adenine. The structure of the final product was confirmed by HRMS and multinuclear NMR data.

Keywords: Synthesis, isonucleoside, phosphonate, anti-HIV

Introduction

The unique properties of phosphonate analogs of the natural phosphoric acid esters make them suitable for use in a continuously increasing variety of applications. Replacement of an Ophosphate group in a biologically-active molecule by a C-phosphonic acid group might be expected to have interesting biological effects. This modification can confer greater stability on these isosteres as the C-P bond that replaces the C-O-P bond cannot be hydrolyzed by the enzymes involved in O-phosphate ester cleavage. In 1986, an acyclic nucleoside phosphonate analog, (S)-9-(3-hydroxy-2-phosphonomethoxy propyl)adenine, was found to be active against a number of viruses.² Since that time, the use of the apparently membrane permeable phosphonate moiety has been intensively studied in the design of more useful antiviral agents. For example, the anti-HIV activities of dideoxynucleosides are critically dependent on their initial intracellular phosphorylation.^{3,4} One way to overcome the difficulty of the first phosphorylation step is to work with prodrugs that deliver intracellularly the monophosphate forms.^{5,6} Another approach to bypass the first phosphorylation step more completely is through phosphonate analogs, that, after intracellular conversion to their diphosphate forms, can serve as inhibitors/ chain terminators in the HIV RT reaction.⁷⁻¹⁰ A unique feature common to all nucleoside phosphonates is their prolonged antiviral action.

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Results and Discussion

3,5-Dideoxy-5-iodo-1,2-O-isopropylidene- α -D-xylo-furanose (3)¹⁵ was prepared in six steps starting from 1,2-O-isopropylidene-α-D-xylofuranose (2) (Scheme 1). The Arbuzov reaction of 3 with triethylphosphite gave 3,5-dideoxy-1,2-O-isopropylidene-5-diethylphosphonyl-α-D-xylofuranose (4) in 85% yield. Acid-catalyzed methanolysis of 4 afforded the α - and β -methylglycosides 5. Reductive demethoxylation of 5 utilized a methodology¹¹ which first involved protection of the 2-hydroxyl group by silvlation followed in situ, by treatment of this product with triethylsilane and TMS-triflate in 1,2-dichloroethane, which produced the tetrahydrofuran 6 in 46% yield. Compound 6 can be converted into 4(S)-(6-amino-9H-purin-9-yl)-2(S)diethylphosphonylmethyltetrahydrofuran (7) either under Mitsunobu conditions or by mesylation (89%) followed by condensation with adenine (49%). COSY and NOESY NMR data of this compound established unambiguously that only the β-isomer was obtained during the condensation step. Dealkylation of the phosphonate ester 7 with bromotrimethylsilane gave the corresponding phosphonic acid, 4(S)-(6-amino-9H-purin-9-yl)tetra-hydrofuran-2(S)-ylmethyl phosphonic acid (8) which was purified by preparative HPLC (85% yield). It was characterized by ¹H, ¹³C and ³¹P NMR data, and high-resolution negative ion mass spectral data. However, evaluation of the compound for in vitro anti-HIV activity in infected CEM-SS cells showed that the activity was much lower than the parent compound, (S,S)-IsoddA.

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Scheme 1

Experimental Section

General Procedures. Nuclear magnetic resonance spectra were recorded on Bruker Model AC300 and WM 360 systems. Ultraviolet spectra were recorded on a Varian Cary Model 3 spectrophotometer. High-resolution FAB mass spectra were obtained on a VG ZAB-HF mass spectrometer. Flash chromatography used 230-400 mesh silica gel. HPLC analyses were carried out on a Beckman-Coulter instrument with C-18 reversed-phase columns.

4(*S*)-(**6**-Amino-9*H*-purin-9-yl)-2(*S*)-diethylphosphonylmethyltetrahydrofuran (7). A mixture of the mesylate of **6** (178 mg, 0.563 mmole), adenine (152 mg, 1.127 mmole), anhydrous K_2CO_3 (156 mg, 1.127 mmole), and 18-crown-6 (149 mg, 0.563 mmole) in DMF (5 mL) was heated at 75 °C for 24 h. The solvent was then evaporated under reduced pressure and the residue was stirred with CHCl₃ (250 mL) and filtered. Evaporation of the solvent and column chromatography of the residue under reduced pressure (0 to 5% MeOH/ CHCl₃) gave pure coupled product **7** (98 mg, 49%): ¹H NMR (CDCl₃) δ 8.32 (s, 1H, H-2), 8.00 (s, 1H, H-8), 5.86 (br s, 2H, NH₂), 5.29 (m, 1H, H-2'), 4.28 (m, 1H, H-4') 4.17 (dd, 1H, H-1', $J_{\text{H1'-H2'}}$ =2.4 Hz, $J_{\text{H1'-H1''}}$ =10.2 Hz), 4.11 and 4.09 (each dq, 2H, OC H_2 CH₃ × 2, $J_{\text{H-H}}$ =7.2 Hz, $J_{\text{H-P}}$ =10.8 Hz), 4.04

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(dd, 1H, H-1", $J_{\text{H1'-H1"}}$ =10.2 Hz, $J_{\text{H1"-H2'}}$ =6.6 Hz), 2.87 (ddd, 1H, H-3", $J_{\text{H4'-H3"}}$ =7.2 Hz, $J_{\text{H2'-H3''}}$ =8.4 Hz, $J_{\text{H3'-H3''}}$ =13.8 Hz), 2.37 (ddd, 1H, H-5', $J_{\text{H5'-H4'}}$ =5.4 Hz, $J_{\text{H5'-H5''}}$ =15.0 Hz, $J_{\text{H5'-P}}$ =18.6 Hz), 2.13 (ddd, 1H, H-5", $J_{\text{H4'-H5''}}$ =7.8 Hz, $J_{\text{H5'-H5''}}$ =15.0 Hz, $J_{\text{H5'-P}}$ =18.6 Hz) 1.30 and 1.29 (each t, 3H, OCH₂CH₃ × 2, J=7.2 Hz); ¹³C NMR (CDCl₃): δ 155.75 (ethylenic C-4 or C-5), 152.83 (C-2), 149.60 (ethylenic C-4 or C-5), 138.04 (C-8), 119.27 (C-6), 74.22 (C-2'), 72.51 (C-5'), 61.79 (m, P(OCH₂CH₃)₂), 54.22 (C-4'), 40.03 (d, C-3', $J_{\text{C-P}}$ =7.1 Hz), 31.95 (d. P-CH₂, $J_{\text{C-P}}$ =140.5 Hz), 16.31 and 16.23 (P(OCH₂CH₃)₂); ³¹P NMR (CDCl₃): δ 26.44 (s).

4(*S*)-(6-Amino-9*H*-purin-9-yl)tetrahydrofuran-2(*S*)-ylmethyl phosphonic acid (8). To a solution of **7** (70 mg, 0.197 mmole) in CH₃CN (3 mL) containing a catalytic amount of dry pyridine (< 0.1 mL) was added Me₃ SiBr (453 mg, 2.96 mmole) and the reaction mixture was stirred for 96 h at 25 °C. Pyridine (0. 4 mL) and water (4 mL) was added and stirring was continued for an additional 2 hours. The reaction mixture was washed with ether (2 x) and the residue was purified by HPLC to give **8** as a crystalline solid (50 mg, 85%): ¹H NMR (D₂O) δ 8.43 and 8.40 (each s, 1H, H-2 and H-8), 5.41 (m, 1H, H-4'), 4.45-4.10 (m, 3H, H-2', H-5' and H-5'') 3.05-2.90 (m, 1H, H-3''), 2.35-1.95 (m, 3H, H-3' and P-CH₂); ¹³C NMR (D₂O): δ 148.77 (ethylenic C-4 or C-5), 146.18 (C-2), 143.55 (ethylenic C-4 or C-5), 140.16 (C-8), 116.09 (C-6), 73.86 (C-2'), 69.15 (C-5'), 53.59 (C-4'), 36.99 (d, C-3', $J_{\text{C-P}}$ =6.5 Hz), 31.75 (d. P-*C*H₂, $J_{\text{C-P}}$ =131.1 Hz); ³¹P NMR (D₂O): δ 21.92 (s); HRMS (ESI) calcd for C₁₀H₁₄N₅O₄P: (M – H)⁻ 298.0704, found: 298.0706.

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