Imide and isatin derivatives as γ-lactam mimics of β-lactam antibiotics

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Dedicated to Professor Charles Rees on his 75th birthday
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Abstract
Activated γ-lactams, which are derivatives of succinimide, phthalimide and isatin with suitable elements of molecular recognition, have been synthesised as mimics of the β-lactam antibiotics and their chemical and biological reactivity determined.

Keywords: γ-Lactams, imides, isatins, antibacterials, β-lactamase, hydrolysis

Introduction

The traditional antibiotics which interact with the bacterial enzymes, the DD-peptidases and the β-lactamases, are β-lactams, for example, benzyl penicillin, 1. The strain energy inherent in the four-membered ring and the non-planarity of the β-lactam were, for many years, thought to be vital for this activity which involves the acylation of an active site serine residue.1 However, it has been demonstrated that the chemical reactivity of β-lactams is not unusual and that molecular recognition is as equally important as acylating power.2 Furthermore non-β-lactam derivatives such as γ-lactams may show chemical reactivity similar to that of the classical penicillins and cephalosporins3 and consequently offer the potential to act as inhibitors of the bacterial enzymes. We report here the synthesis of activated γ-lactams, e.g. 2 and 3 as structural mimics of the classical β-lactam antibiotics.
Attempts to substitute larger membered rings for the β-lactam ring and yet retain good antibacterial activity have been tried before, for example, using imides and isatins but with little or no success. Other activated γ-lactams which have been synthesised as isosteres of the β-lactams include the pyrazolidinones, imidazolidinones, cycloserines and hydantoins. The antibacterial activity of β-lactam antibiotics results from their acylation of the transpeptidases involved in the synthesis of the bacterial cell wall. Bacterial resistance to these antibiotics is predominantly due to the production of bacterial β-lactamases which catalyse ring opening and hydrolysis of the β-lactam. One of the problems associated with increasing the ring size from four-membered is that the ‘back-reaction’, ie recyclisation, may occur more readily and regenerate the free enzyme (Scheme 1).

\[
\begin{align*}
\text{EnzOH} & \quad \text{(CH)\_n} \quad \text{N} \quad \text{O} \quad \text{R} \quad \text{N} \\
\text{O} \quad \text{OEnz} & \quad \text{(CH\_n)NHR}
\end{align*}
\]

Scheme 1. Reversible ring opening.

For example, we have shown that ring closure can occur by the intramolecular aminolysis of acylenzymes to form γ-lactams but not β-lactams. This paper describes some attempts to discourage the recyclisation whilst retaining features required for good antibiotic activity. Activated structures based on succinimide, phthalimide and isatin which have a potential leaving group (Lg) α-substituted to the incipient amine may not only reduce the probability of reclosure of the ring but also generate a ‘second trap’ for enzyme inactivation (Scheme 2).
Scheme 2. The potential mode of action with serine enzymes of $\gamma$-lactams with a leaving.

Nucleophilic attack at the $\gamma$-lactam carbonyl followed by ring opening and concomitant elimination of the leaving group generates an electrophilic imine. As a result of expulsion of the leaving group the nucleophilicity of the nitrogen is reduced, so the ease of the ‘back-reaction’ and ring closure with regeneration of the free enzyme is discouraged. In addition, the electrophilic imine could also covalently react with any nucleophilic species in the active site, increasing the likelihood of enhanced inactivation, potentially resulting in autolysis and death of the bacterium.

The introduction of a second carbonyl group into a $\gamma$-lactam ring either adjacent to the nitrogen, as in phthalimide and succinimide, or adjacent to the carbonyl as in isatin, activates the amide bond sufficiently to bring its reactivity close to that of the $\beta$-lactam of $\beta$-lactam antibiotics. To be an effective acylating agent it appears that the second order rate constant for hydrolysis, $k_{OH}$, should be in the range exhibited by the biologically active penicillins and cephalosporins, i.e. 0.01 to 1 M$^{-1}$s$^{-1}$. The $\gamma$-lactams of imides and isatins, which have alkaline hydrolysis rates that fall within this range, are activated enough to be considered as starting points for potential antibiotics and/or inactivators of the $\beta$-lactamase enzymes. The synthesis of
the \( \gamma \)-lactam analogues of succinimide, phthalimide and isatin, is reported together with their hydrolytic and antibiotic activity and their inactivation of \( \beta \)-lactamase enzymes.

**Results and Discussion**

The structure-activity relationships of \( \beta \)-lactam antibacterial agents are not very well defined although it is clear that the minimum requirements involve good acylating power combined with the correct geometrical relationship between the acylating centre and the carboxylate anion. The compounds synthesised here represent a variety of different molecular shapes but with reactivities falling into the required range.

The N-\( \alpha \)-hydroxyacetic acid adducts of succinimide 4 (Scheme 3), phthalimide 5 (Scheme 4) and isatin 6 (Scheme 5) were synthesised by condensation with glyoxylic acid. The chemical shifts of the methine hydrogens in the \( ^1 \)H NMR spectra of these structures are particularly diagnostic being at 5.67\( \delta \), 5.88\( \delta \) and 6.01\( \delta \). These adducts are unlikely to be able to expel the hydroxyl group to generate the imine trap, nonetheless they were tested for antibacterial activity, but none was found.

\[
\begin{align*}
4 & \quad \text{i) Glyoxylic acid (50 wt%), THF, reflux; ii) 7; SOCl}_2, \text{THF, 0°C, H}_2\text{O; 9; acetyl chloride, pyridine, 0°C; 11; phenylacetyl chloride, pyridine, 0°C}.
\end{align*}
\]

These adducts were found to be unstable in water and quickly decomposed to their corresponding imide and glyoxylic acid, the half-life of the isatin derivative 6 was only 2-3 minutes at pH 7 and 20°C. The hydroxyl function was consequently converted to a number of more convenient leaving groups. The first leaving groups to be considered were sulphonates but attempts at sulphonation of the hydroxyl group using various sulphonyl halides met with little success. For example, the reaction of 4 with either methanesulphonyl or p-toluenesulphonyl
chloride resulted in sulphonate derivatives which were very labile and immediate decomposition occurred on attempted isolation.

![Chemical structures](image)

**Scheme 4**

*Reagents and conditions*: i) Glyoxylic acid (50 wt%), THF, reflux; ii) 8; SOCl₂, THF, 0 °C, H₂O; 10; acetyl chloride, pyridine, 0 °C; 12; phenylacetyl chloride, pyridine, 0 °C; iii) methyl glyoxylate, THF, reflux; iv) 2,6-dichlorobenzoyl chloride, pyridine, 0 °C; v) 13; LiI.H₂O, pyridine, reflux

Compounds 4, 5 and 6 were reacted with 2.5 equivalents of thionyl chloride to give the corresponding halides as potential leaving groups - 7, 8 and 19, respectively. The free carboxylic acid groups present in these derivatives also reacted to give rise to the formation of the corresponding acyl chlorides and their subsequent reaction with free hydroxy residues formed polymeric esters as side products. Hydrolysis of the acyl chlorides with ice water for 3 h. followed by a work-up and trituration using dichloromethane was the best method found for purifying the products. The characteristic methine protons moved downfield to 6.42δ (7), 6.70δ (8) and 7.18δ (19), respectively.
Reaction of the glycolic acid derivatives 4 and 5 with acid chlorides gave esters 9-12 which, in the active site of the enzymes, could in principle react by expelling their corresponding carboxylate anions. Pyridine was used as both solvent and base in the synthesis but numerous purification procedures using silica gel chromatography were required to obtain pure products in relatively low yield.

In the active sites of the enzymes which recognise β-lactam antibiotics\textsuperscript{15-19} there are a number of different functional groups close enough to the proposed incipient imine intermediate formed by ring opening to be able to react with this ‘second trap’ electrophilic centre. However, such nucleophiles could also attack the carbonyl carbon of the ester “leaving group” in 9-12 and so compete with that at the γ-lactam centre. In order to protect this site from nucleophilic attack a more sterically hindered ester was used. Subramanyam\textsuperscript{20} has reported the use of 2,6-disubstituted benzoates as leaving groups in compounds used to acylate the serine protease Human Leukocyte Elastase (HLE). The inhibition activity was dependent on the nature of the leaving group and the 2,6-disubstituents sterically prevented undesirable nucleophilic attack on the benzoyl carbonyl group. We therefore attempted to prepare the corresponding phthalimide derivative, 13. The
acylation of the free hydroxy acid 5 with 2,6-dichlorobenzoyl chloride proved ineffective probably because of competing anhydride formation. The carboxylic acid residue was consequently protected as its methyl ester before acylation.

The first stage was the reaction of phthalimide with methyl glyoxylate to give N-phthalimido-α-hydroxy-acetic acid methyl ester 14. 1H NMR showed a characteristic methyl ester singlet at 3.77δ and the methine proton of these glyoxylates at 5.93δ. Reaction of 14 with 2,6-dichlorobenzoyl chloride in pyridine to give N-phthalimido-α-2,6-dichlorobenzoyl-acetic acid methyl ester 15, the methine proton of the product now appearing at 7.24δ. When N-phthalimido-α-2,6-dichlorobenzoyl-acetic acid methyl ester 15 was reacted with 1 equivalent of lithium iodide monohydrate in refluxing pyridine to give 13 in high yield. Having established a three step synthesis involving facile purifications and good overall yield, it was decided to try this method to make the phenylacetyl derivative 12, previously synthesised by the much lower yielding, but only two step synthesis, described earlier. The reaction of N-phthalimido-α-hydroxy-acetic acid methyl ester 14 with phenylacetyl chloride gave the diester 16, but in low yield. The phenylacetylation of isatin-N-α-hydroxy-acetic acid methyl ester also gave poor yields compared with that obtained for the 2,6-dichloro derivative. The methine shifted from 5.93δ in the starting material 14 to 7.02δ in the product 16. However, when the lithium iodide demethylation of the methyl ester was attempted degradation occurred with the formation of phenylacetic acid as well as some unidentifiable products. The suggestion that lithium iodide is a specific reagent for ethyl and methyl esters only does not appear universal. Iodide attack on N-phthalimido-α-phenylacetyl-acetic acid methyl ester 16 apparently occurs at the more electron deficient methine carbon (Scheme 6), even though this site is more sterically hindered.

![Scheme 6](image-url)
Presumably the 2,6-dichloro substituted compound 15 has this methine position shielded by the two bulky chloride groups preventing attack at this site. Attempts to selectively remove the methyl ester using pig liver esterase also failed as did other attempts to remove the carboxyl protecting ester group. Using a 4-nitrobenzyl ester protecting group for the carboxylic acid group, 17, followed by acylation of the hydroxyl group led to products 20 and 21 which also could not be selectively deprotected.

It was felt that more enzyme recognition needed to be incorporated into the structures such as an acylamino side chain commonly found in active penicillins. As well as improving the affinity of the substrate for the target enzymes, the reactivity of the amide bond would also be increased. (R/S)-Aspartic acid was N-acylated using phenylacetyl chloride and then converted to the substituted succinic anhydride by the method of Mardle using acetic anhydride. When this anhydride was heated with ammonium carbonate only an 18% conversion to the imide was observed. A series of attempts to heat N-acylated succinic acid with various nitrogen sources gave differing yields of the desired product, ie ammonia (8%), ammonium carbonate (6%) and finally urea (51%). Now that an effective route was available to the imide stage, optically pure (S)-aspartic acid was acylated to give the diacid 30 followed by cyclisation to the imide 31 using urea (Scheme 7). The next step was the previously established reaction of 31 with glyoxylic acid which resulted in formation of 31 with a 50/50 mixture of diastereoisomers. Conversion of the hydroxy acid 32 to the chloro acid 33 by use of thionyl chloride gave a near equal mixture of the required diastereoisomeric product (Scheme 7). The α-carbonyl group of isatin was converted to O-benzyloxime derivatives 27-29 to act also as a potential mimic of the N-phenylacetamido side chain of benzylpenicillin, 1.

![Scheme 7](image-url)
**Reagents and conditions:** i) Phenylacetyl chloride, NaOH, 0°C; ii) Urea, 170°C; iii) Glyoxylic acid.H₂O, THF, reflux; iv) SOCl₂, THF, 0°C, H₂O

**Chemical Reactivity** - The second-order rate constants for the hydroxide-ion catalysed hydrolysis of the γ-lactams are given in Table 1. For comparison the second-order rate constant for the alkaline hydrolysis of benzyl penicillin is 0.15 M⁻¹ s⁻¹ and so the γ-lactams have a similar or even greater reactivity indicating their suitability, on chemical grounds, as potential acylating agents of β-lactamases and DD-peptidases.² It has been suggested that the second order rate constant, kOH, is a good indicator of enzyme acylating power and should have a value of 0.01 to 1.0 M⁻¹ s⁻¹ to maximise inactivation with competing hydrolysis.²²

**Table 1.** The second order rate constants, kOH, for the hydroxide ion catalysed hydrolysis of the γ-lactams in water at 30°C

<table>
<thead>
<tr>
<th>Name</th>
<th>kOH/M⁻¹s⁻¹</th>
</tr>
</thead>
<tbody>
<tr>
<td>n-Phthalimido-α-chloro-acetic acid, 8</td>
<td>68.3</td>
</tr>
<tr>
<td>n-Phthalimido-α-acetoxyacetic acid, 10</td>
<td>33.5</td>
</tr>
<tr>
<td>n-Phthalimido-α-hydroxyacetic acid methyl ester, 14</td>
<td>22.1</td>
</tr>
<tr>
<td>1,7-Dimethylisatin</td>
<td>0.534</td>
</tr>
<tr>
<td>n-Methylisatin</td>
<td>0.465</td>
</tr>
<tr>
<td>7-Methylisatin</td>
<td>1.73</td>
</tr>
<tr>
<td>Benzylpenicillin 1</td>
<td>0.15</td>
</tr>
<tr>
<td>7-Methylisatin-N-acetic acid</td>
<td>0.117</td>
</tr>
</tbody>
</table>

**Inhibition Studies** - The γ-lactams were tested for inhibition²³ of the β-lactamase enzymes from *Bacillus cereus* 569/H class A and class B and the class C enzyme from *Enterobacter cloacae* P99 but no significant activity was found. The γ-lactams were also screened for antibacterial activity against a wide range of micro-organisms but they showed no significant activity up to a concentration of 128 µg/cm³.

**Experimental Section**

**General Procedures.** Melting points were determined on a Gallenkamp melting point apparatus and are uncorrected. 270 MHz ¹H and 67 MHz ¹³C NMR were determined on a Bruker AC-270 spectrometer with tetramethylsilane as internal standard. All J values are given in Hz. IR spectra were recorded on a Perkin Elmer 1600 Series FTIR and FAB MS were performed by Swansea Mass Spectrometry Service and Zeneca Pharmaceuticals. All elemental analyses were performed by MEDAC Ltd., Brunel University. Fluka silica gel 60 was used for all chromatographic separations and thin layer chromatographic separations and thin layer chromatographic techniques used Merck silica gel 60 F₂₅₄ TLC plates. Ether refers to diethyl ether. Tetrahydrofuran was dried by distilling over lithium aluminium hydride under dry nitrogen.
Dichloromethane was dried by passing it through a column of Grade I activated alumina into the reaction flask under argon.

**Method A. N-Succinimido-α-hydroxyacetic acid (4)** Succinimide (50.0 g, 0.51 mol) was dissolved in THF (300 cm³) at room temperature with stirring. To this was added glyoxylic acid (50 wt%) (74.80 g, 0.51 mol) and the reaction mixture was heated to reflux for 3 h. Evaporation to dryness under reduced pressure produced a yellow oil which eventually crystallised. This was recrystallised from ethyl acetate yielding 4 (52.95 g, 60%) as white crystals. M. p. 140-141°C (lit. m.p. 140-141°C) (Found: C, 4.71; H, 4.0; N, 8.0. C₆H₇NO₅ requires C, 41.6; H, 4.0; N, 8.1%); $\nu_{\text{max}}$ (Nujol mull)/cm⁻¹ 3391, 3048, 1778, 1740, 1685, 1455, 1400, 1285, 1170, 743; $\delta_{\text{H}}$ (CD₃OD) 2.75 (4H, s, 2 x CH₂) 5.67 (1H, s, CH); m/z 174 (M+H)+, 156, 128, 100, 82, 72, 56, 44 and 28.

**N-Phthalimido-α-hydroxyacetic acid (5).** Method A was used in the synthesis of 5 using phthalimide (10.0 g, 68.0 mmol), glyoxylic acid (50 wt%) (30.20 g, 0.204 mol) and THF (200 cm³) giving a cream solid on evaporation to dryness under reduced pressure. Recrystallisation from ethyl acetate yielded 5 (13.19 g, 88%) as a white solid. M.p. 190-191°C (lit. m.p. 191-192°C) (Found: C, 54.0; H, 3.1; N, 6.4. C₁₀H₇NO₅ requires C, 54.3; H, 3.2; N, 6.3%); $\nu_{\text{max}}$ (Nujol mull)/cm⁻¹ 3340, 1745, 1709, 1465, 1379, 1290, 1173, 1075, 920, 730; $\delta_{\text{H}}$ (CD₃OD) 5.88 (1H, s, CHOH) 7.90 (4H, m. aryl,; m/z 221 M+. 199, 175, 165, 153 and 147.

**Isatin-N-α-hydroxyacetic acid (6).** Method A was used in the synthesis of 6 using isatin (25.0 g, 0.170 mol), glyoxylic acid monohydrate (47.0 g, 0.612 mol) and THF (500 cm³). Evaporation to dryness give a deep red oil which slowly crystallised. Recrystallisation using ethyl acetate afforded 6 (24.4 g, 65%) as an orange powder. M.p. 167-168°C (Found: C, 54.3; H, 3.2; N, 6.35. C₁₀H₇NO₅ requires C, 54.3; H, 3.2; N, 6.3%); $\nu_{\text{max}}$ (Nujol mull)/cm⁻¹ 3443, 3019, 2927, 2855, 1736, 1726, 1607, 1467, 1353, 1216, 1191, 766; $\delta_{\text{H}}$ (D₆ DMSO) 6.01 (1H, s, CHOCH); m/z (ESP+) 222 (M+H)+, 206, 204, 194, 176, 74 and 73.

**Method B. N-Succinimido-α-chloroacetic acid (7).** N-Succinimido-α-hydroxy-acetic acid 7 (2.00 g, 12.0 mmol) was dissolved in dry THF (50 cm³) with stirring at room temperature under nitrogen. The solution was cooled to 0°C before thionyl chloride (3.44 g, 29.0 mmol) was added dropwise. Evolution of an acidic gas was observed. The reaction mixture was allowed to warm to room temperature overnight. Ice (10 g) was added and the mixture stirred for a further 3 h. The solution was extracted using ethyl acetate (3 x 50 cm³), the organics dried (MgSO₄), filtered and evaporated to give a viscous brown oil. Trituration of this with dichloromethane yielded 6 (0.68 g, 28%) as a white powder. M.p. 182-184°C (Found: C, 37.55; H, 3.2; N, 7.1. C₆H₆ClNO₄ requires C, 37.6; H, 3.2; N, 7.3%); $\nu_{\text{max}}$ (Nujol mull)/cm⁻¹ 3053, 1788, 1749, 1688, 1460, 1413, 1396, 1285, 1173, 745; $\delta_{\text{H}}$ (CD₃OD) 2.90 (4H, s, 2 x CH₂) 6.42 (1H, s, CHCl); m/z 192 (M-H)⁻ (Cl³⁻), 155, 146 and 143.

**N-Phthalimido-α-chloroacetic acid (8).** Method B was used in the synthesis of 8 using N-phthalimido-α-hydroxy-acetic acid 5 (2.00 g, 9.0 mmol), thionyl chloride (3.37 g, 28.0 mmol)
and dry THF (100 cm$^3$). Trituration of the resultant cream solid with hot dichloromethane yielded $8$ (0.30 g, 13%) as a white powder. M.p. 146-148°C (Found: C, 49.7; H, 2.6; N, 5.7. C$_{10}$H$_6$ClNO$_4$ requires C, 50.1; H, 2.5; N, 5.85%). $\nu$$_{\text{max}}$ (Nujol mull)/cm$^{-1}$ 3278, 1788, 1750, 1463, 1277, 1238, 1198, 1174, 1076, 918, 720; $\delta$$_H$ (CD$_3$OD) 6.70 (1H, s, CH$_2$Cl) 7.98 (4H, m, aryl); m/z 240 (M-H)$^-$ (Cl$_{37}$), 238 (M-H)$^-$ (Cl$_{35}$), 219, 204, 194, 159 and 146.

Isatin-N-$\alpha$-chloroacetic acid (19). Method B was used in the synthesis of $19$ using isatin-N-$\alpha$-hydroxy-acetic acid $6$ (10.0 g, 45.0 mmol), thionyl chloride (16.15 g, 0.136 mol), a catalytic amount of DMF (0.5 cm$^3$) and dry CH$_2$Cl$_2$ (150 cm$^3$). The solution was evaporated to dryness and re-dissolved in THF (150 cm$^3$). The mixture was stirred and cooled to 0°C before the addition of ice (50g). The evolution of gases was observed and this stopped after 2 h. The homogenous reaction mixture was again evaporated under reduced pressure until all THF had been removed and the aqueous layer extracted with ethyl acetate (3 x 75 cm$^3$). The solution was evaporated to dryness and the residue was crystallised and was collected by filtration. The first crop of the reaction gave $19$ (1.30 g, 12%) as an orange powder. M.p. 150-151°C (Found: C, 50.1; H, 2.5; N, 5.85. C$_{10}$H$_6$ClNO$_5$ requires C, 50.1; H, 2.5; N, 5.85%). $\nu$$_{\text{max}}$ (CHCl$_3$)/cm$^{-1}$ 3446, 3020, 2927, 2855, 2400, 1747, 1727, 1608, 1471, 1380, 1352, 1216, 1121, 758; $\delta$$_H$ (D$_6$ DMSO) 7.18 (1H, s, CH$_2$Cl) 7.20 (2H, m, 2 aryl protons in the 5 and 6 positions of isatin) 7.78 (1H, s, CH); m/z (FAB+) 241 (M+H)$^+$. 239, 223, 222, 206, 204, 194, 176, 148, 146, 130, 116, 84, 83, 74, 69 and 64.

Method C. N-Succinimido-$\alpha$-acetoxyacetic acid (9). N-Succinimido-$\alpha$-hydroxy-acetic acid $3$ (2.00 g, 12.0 mmol) was dissolved in pyridine (10 cm$^3$) at 0°C with stirring under nitrogen. Acetyl chloride (1.00 g, 13.0 mmol) was added dropwise to the solution over 5 mins. The reaction was left for 4 h. and during this time a brown precipitate was observed. After evaporation to dryness the resultant brown solid was dissolved in a 50/50 ethyl acetate/2M hydrochloric acid mix (100 cm$^3$). The aqueous was extracted with ethyl acetate (3 x 50 cm$^3$) and the combined organics dried using Na$_2$SO$_4$, filtered and evaporated to give a crude yellow oil. Silica gel chromatography (50/50 ethyl acetate/hexane) yielded $9$ (1.38 g, 56%) as a colourless oil which later crystallised. M.p. 120-122°C (Found: C, 44.2; H, 4.4; N, 6.5. C$_8$H$_9$NO$_6$ requires C, 44.6; H, 4.2; N, 6.5%). $\nu$$_{\text{max}}$ (Nujol mull)/cm$^{-1}$ 3284, 1785, 1733, 1727, 1451, 1420, 1374, 1248, 1190, 1049, 741; $\delta$$_H$ (CD$_3$OD) 2.17 (3H, s, CH$_3$) 2.85 (4H, s, 2 x CH$_2$) 6.77 (1H, s, CH); m/z 214 (M-H)$^-$, 165, 151 and 143.

N-Phthalimido-$\alpha$-acetoxyacetic acid (10). Method C was used in the synthesis of $10$ using N-phthalimido-$\alpha$-hydroxy-acetic acid $5$ (2.00 g, 9.0 mmol), acetyl chloride (0.78g, 10.0 mmol) and pyridine (10 cm$^3$). A further 10 cm$^3$ of pyridine was added to help solubilise the thick suspension formed and the reaction left overnight. Recrystallisation of the crude yellow solid from ethyl acetate afforded $10$ (0.72 g, 30%) as a white solid. M.p. 186-188°C (Found: C, 54.3; H, 3.5; N, 5.1. C$_{12}$H$_9$NO$_6$ requires C, 54.7; H, 3.4; N, 5.3%). $\nu$$_{\text{max}}$ (Nujol/mull)/cm$^{-1}$ 3292, 1787, 1736,
1463, 1377, 1265, 1195, 1122, 1053, 745; δH (CD3OD) 2.16 (3H, s, CH3) 6.94 (1H, s, CHO) 7.90 (4H, m, aryl); m/z 262 (M-H)-, 188, 146 and 122.

N-Succinimido-α-phenylacetoxyacetic acid (11). Method C was used in the synthesis of 11 using N-succinimido-α-hydroxy-acetic acid 4 (2.00 g, 12.0 mmol), phenylacetyl chloride (3.58 g, 23.0 mmol) and pyridine (25 cm³). Trituration of the crude brown solid with dichloromethane yielded 11 (1.39 g, 41%) as a white powder. M.p. 149-151°C (Found: C, 58.0; H, 4.6; N, 4.5. C14H13NO6 requires C, 57.7; H, 4.5; N, 4.8%); νmax (Nujol mull)/cm⁻¹ 1787, 1751, 1682, 1496, 1459, 1377, 1233, 1189, 1117, 1055, 915, 721; δH (CD3OD) 2.77 (4H, s, 2 x CH₂) 3.75 (2H, s, CH₂Ph) 6.74 (1H, s, CHO) 7.27 (5H, m, Ph); m/z 290 (M-H)-, 247, 227, 172, 155, 135 and 111.

N-Phthalimido-α-phenylacetoxyacetic acid (12). Method C was used in the synthesis of 12 using N-phthalimido-α-hydroxy-acetic acid 5 (4.59 g, 21.0 mmol), phenylacetyl chloride (6.42 g, 42.0 mmol) and pyridine (30 cm³). Trituration of the crude brown solid with hot 60.80 petroleum ether removed excess phenylacetic acid and silica gel chromatography (gradient elution 25% ethyl acetate/hexane to 100% ethyl acetate) yielded 12 (0.94 g, 13%) as a white powder. M.p. 123-125°C (Found: C, 63.4; H, 3.9; N, 3.9. C18H13NO6 requires C, 63.7; H, 3.9; N, 4.1%); νmax (Nujol mull)/cm⁻¹ 1789, 1730, 1726, 1385, 1265, 1188, 1132, 1030, 741; δH (CD3OD) 3.78 (2H, s, CH₂) 6.95 (1H, s, CHO) 7.23 (5H, m, CH₂Ph) 7.88 (4H, m, aryl); m/z 340 (M+H)+, 294, 277, 204, 177, 171, 167, 154, 137, 118 and 105.

N-Phthalimido-α-hydroxyacetic acid methyl ester (14). Phthalimide (20.0 g, 0.136 mol) was dissolved in THF (150 cm³) at room temperature with stirring. To this was added methyl glyoxylate (17.94 g, 0.204 mol) and the reaction mixture was heated to reflux for 2 h. Evaporation to dryness under reduced pressure produced a white semi-solid. This was recrystallised from ethyl acetate yielding 14 (19.22 g, 60%) as a white solid. M.p. 132-133°C (Found: C, 56.05; H, 3.8; N, 6.0. C11H9NO5 requires C, 56.1; H, 3.8; N, 5.95%); νmax (Nujol mull)/cm⁻¹ 3250, 1784, 1760, 1460, 1378, 1250, 1198, 1120, 722; δH (CDCl3) 3.77 (3H, s, CH3) 5.93 (1H, s, CHO) 7.84 (4H, m, aryl); m/z 234 (M-H)-, 212, 188, 153, 147 and 90.

Method D: N-Phthalimido-α-2,6-dichlorobenzoxyacetic acid methyl ester (15). N-Phthalimido-α-hydroxy-acetic acid methyl ester 14 (1.0 g, 4.3 mmol) was dissolved in pyridine (10 cm³) at 0°C with stirring under nitrogen. To this was added dropwise 2,6-dichlorobenzoyl chloride (1.34 g, 6.4 mmol) and a colour change from yellow to brown was observed following by the appearance of a white precipitate. The reaction mixture was allowed to warm to room temperature over 3 h. The mixture was evaporated to dryness then dissolved in a 50/50 ethyl acetate/2M hydrochloric acid mix (100 cm³). The aqueous was extracted with ethyl acetate (3 x 50 cm³) and the combined organics washed with saturated sodium hydrogen carbonate solution. The organics were dried with Na2SO4, filtered and evaporated to give a dark brown oil. Silica gel chromatography (10% ethyl acetate/petroleum ether (60/80) afforded 15 (1.16 g, 67%) as a white solid. M.p. 152-153°C (Found: C, 52.8; H, 2.7; N, 3.5. C18H11Cl2NO6 requires C, 52.9; H, 2.7; N, 3.4%); νmax (Nujol mull)/cm⁻¹ 1769, 1760, 1709, 1455, 1370, 1256, 1190, 1149, 1080, 722; δH (CDCl3) 3.91 (3H, s, CH3) 7.24 (1H, s, CHO) 7.30 (3H, m, benzyol aromatics) 7.85 (4H, m.
Phthalimide aromatics); δ\text{C} (CDCl₃) 53.86 (CH₃) 69.25 (NCHO) 124.30 (para carbon of benzoyl) 127.96 (2 x CICCH) 131.38 (2 x C(O)C of imide) 131.50 (2 x meta aryl of imide) 131.74 (CIC(C(O)) 132.39 (2 x ClC) 134.93 (2 x C(O)C of imide) 135.44 (ClC(O)) 165.58 (2 x NC(O) of imide) m/z 408 (M+H)⁺, 355, 341, 263, 249, 235, 218 and 190.

**N-Phthalimido-α-2,6-dichlorobenzoxyacetic acid (13).** N-Phthalimido-α-2,6-dichlorobenzoyle acetic acid methyl ester 15 (0.60 g, 1.5 mmol) and lithium iodide monohydrate (0.224 g, 1.5 mmol) were dissolved in pyridine (10 cm³) under nitrogen and the solution heated to reflux for 24 h. The mixture was evaporated to dryness and taken up into ethyl acetate (50 cm³) then washed with saturated sodium hydrogen carbonate solution (50 cm³). The aqueous was extracted using ethyl acetate (3 x 50 cm³) then taken to pH 2 with 2M hydrochloric acid. The aqueous was re-extracted with ethyl acetate (3 x 50 cm³) and the combined organics dried using MgSO₄, filtered and evaporated to dryness to give a yellow solid. This was triturated with dichloromethane then ether to give 12 (0.388 g, 67%) as a white solid. M.p. 181-183°C (Found: C, 51.6; H, 2.35; N, 3.5. C₁₇H₉Cl₂NO₆ requires C, 51.8; H, 2.3; N, 3.55%); ν\text{max} (Nujol mull)/cm⁻¹ 1767, 1762, 1709, 1464, 1377, 1315, 1256, 1198, 1130, 1050, 875, 781, 723; δ\text{H} (CD₃OD) 7.20 (1H, s, CH₂O) 7.42 (3H, s, benzoyl aromatics) 7.83 (4H, m, phthalimide aromatics); m/z 394 (M+H)⁺, 204, 185, 177, 173, 160, 148, 139, 133, 123 and 105.

**N-Phthalimido-α-phenylacetoxyacetic acid methyl ester (16).** Method D was used in the synthesis of 16 using N-phthalimido-α-hydroxy-acetic acid methyl ester 14 (3.0 g, 13.0 mmol), pyridine (25 cm³) and phenylacetyl chloride (5.91 g, 38.0 mmol) and the appearance of a white precipitate was observed. Silica gel chromatography (CH₂Cl₂) afforded 15 (1.27 g, 28%) as a white solid. M.p. 72-73°C (Found: C, 64.2; H, 4.3; N, 3.9. C₁₉H₁₅NO₆ requires C, 64.5; H, 4.25; N, 4.0%); ν\text{max} (Nujol mull)/cm⁻¹ 1785, 1760, 1729, 1457, 1380, 1246, 1188, 1144, 738; δ\text{H} (CDCl₃) 3.81 (3H, s, CH₃) 3.77 (2H, q, JAB 15.8 and 15.8, CH₂Ph) 7.02 (1H, s, CH₂O) 7.29 (5H, m, CH₂Ph) 7.85 (4H, m, phthalimide aromatics); m/z 354 (M+H)⁺, 308, 291, 218, 191, 171, 162 and 132.

**Attempted de-esterification of N-phthalimido-α-phenylacetoxyacetic acid methyl ester (15).** N-Phthalimido-α-phenylacetyl-acetic acid methyl ester 16 (0.75 g, 2.1 mmol) and lithium iodide monohydrate (0.355 g, 2.3 mmol) were dissolved in pyridine (10 cm³) under nitrogen and the solution heated to reflux for 24 h. The mixture was evaporated to dryness and taken up into ethyl acetate (50 cm³) then washed with saturated sodium hydrogen carbonate solution (50 cm³). The aqueous was extracted using ethyl acetate (3 x 50 cm³) then taken to pH2 with 2M hydrochloric acid. The aqueous was re-extracted with ethyl acetate (3 x 50 cm³) and the combined organics dried using MgSO₄, filtered and evaporated to dryness to give a brown solid. Analysis of the crude reaction material revealed by-product phenylacetic acid and further degradation products of N-Phthalimido-α-iodo-acetic acid methyl ester.

**Isatin-N-α-hydroxyacetic acid 4-nitrobenzyl ester (17).** p-Nitrobenzyl glyoxylate (10.0 g, 44.1 mmol) was dissolved in toluene (150 cm³) and the solution refluxed for 1 h. with a Dean and Stark apparatus attached. After cooling, isatin (3.23 g, 22.0 mmol) was added and the
solution refluxed overnight. On cooling with ice, a precipitate appeared which when filtered gave 17 (5.83 g, 74%) as an orange crystalline solid. M.p. 160-163°C (Found: C, 57.1; H, 3.4; N, 8.1. C₁₇H₁₂N₂O₇ requires C, 57.3; H, 3.4; N, 6.9%); ν_max (Nujol mull)/cm⁻¹ 3280, 1734, 1520, 1350; δ_H (D₆ DMSO) 5.40 (2H, s, CH₂) 6.25 (1H, d, J 6.4, CH) 7.12-7.62 (4H, m, aryl) 7.54 (2H, d, J 8.8, para aryl) 8.17 (2H, d, J 8.8, para aryl).

**Isatin-N-α-acetoxyacetic acid 4-nitrobenzyl ester (20).** Method D was used for the synthesis of 20 using isatin-N-α-hydroxy-acetic acid 4-nitrobenzyl ester 17 (2.0 g, 5.62 mmol) pyridine (10 cm³) and acetyl chloride (0.49 g, 6.18 mmol). After reaction for 3 h. the solution was then poured onto ice water (100 cm³) and the resultant precipitate filtered, washed with cold water and dried to give 20 (1.82 g, 54%) as a dark yellow powder. M.p. 138-140°C (Found: C, 57.4; H, 3.6; N, 7.1. C₁₉H₁₄N₂O₈ requires C, 57.3; H, 3.5; N, 7.0%); ν_max (Nujol mull)/cm⁻¹ 1770, 1740, 1620, 1520; δ_H (D₆ DMSO) 2.15 (3H, s, CH₃) 5.40 (2H, s, CH₂) 7.10-7.25 (2H, m, aryl) 7.28 (1H, s, N-CH) 7.52 (2H, d, J 8.6, para aryl) 7.60-7.70 (2H, m, aryl) 8.15 (2H, d, J 8.6, para aryl); m/z 399 (M+H)+, 398 and 339.

**Isatin-N-α,2,6-dichlorobenzoxyacetic acid 4-nitrobenzyl ester (21).** Method D was used in the preparation of 21 using isatin-N-α-hydroxy-acetic acid 4-nitrobenzyl ester 17 (2.0 g, 5.62 mmol), pyridine (10 cm³) and 2,6-dichlorobenzoyl chloride (1.29 g, 6.18 mmol). The reaction afforded 21 (2.36 g, 52%) as a cream powder. M.p. 105-106°C; ν_max (Nujol mull)/cm⁻¹ 1776, 1736, 1610; δ_H (D₆ DMSO) 5.50 (2H, s, CH₂) 7.25-7.28 (2H, m, aryl) 7.55-7.80 (8H, m, aryl and NCH) 8.20 (2H, d, J 8.7, para aryl); m/z 530 (M)+.

**(R/S)-N-Phenylacetylaspartic acid** Phenylacetyl chloride (58.07 g, 0.376 mol), 4M sodium hydroxide solution (92.5 cm³), (R/S)-aspartic acid (50.0 g, 0.376 mol) and 2M sodium hydroxide solution (375 cm³) were used. The reaction yielded the product (75.05 g, 80%) as a white crystalline solid. M.p. 116-117°C (lit m.p. 117°C) (Found: C, 57.0; H, 5.1; N, 5.6. C₁₂H₁₃NO₅ requires C, 57.4; H, 5.2; N, 5.6%); ν_max (Nujol mull)/cm⁻¹ 3409, 3330, 2960, 1734, 1709, 1646, 1539, 1457, 1344, 1283, 1234, 1191, 1154, 959, 734; δ_H (CD₃OD) 2.92 (2H, dd, J_ABX 6.5 and 5.5, CH₂CH) 3.57 (2H, s, H-5) 4.74 (1H, dd, J_ABX 6.5 and 5.5, CH₂CH) 7.29 (5H, m, aryl); m/z (ESP+) 251 (M+H)+, 234, 134, 87, 55.

**(R/S)-N-Phenylacetylaspartic anhydride** (R/S)-N-Phenylacetylaspartic acid (50.0 g, 0.20 mol) and acetic anhydride (150 cm³) were used as described. The product (30.62 g, 66%) was a white crystalline solid. M.p. 160-162°C (lit. m.p. 162°C) (Found: C, 62.0; H, 4.8; N, 6.0. C₁₂H₁₁NO₄ requires C, 61.8; H, 4.7; N, 6.0%); ν_max (Nujol mull)/cm⁻¹ 3331, 2918, 2853, 1851, 1824, 1780, 1652, 1525, 1456, 1231, 1196, 1068, 974, 904, 737; δ_H (D₆ DMSO) 2.79 (1H, dd, J_ABX 18.4 and 5.7, CHCH₂) 3.22 (1H, dd, J_ABX 18.4 and 10.1, CHCH₂) 3.50 (2H, s, CH₂Ph) 4.65 (1H, m, CHCH₂) 7.27 (5H, m, aryl) 8.96 (1H, d, J 7.1, NH).

**(R/S)-N-Phenylacetylaspartimide**

**Method 1** (R/S)-N-Phenylacetylaspartic anhydride (24.83 g, 0.107 mol) was added to finely ground ammonium carbonate (5.63 g, 59.0 mmol), mixed thoroughly and heated to 180°C for 1 h. During this time, the mixture turned partially molten and more ammonium carbonate (5.63 g,
59.0 mmol) was added until no further gases evolved. The mixture turned from colourless to brown and water droplets were observed condensing on the apparatus. The reaction was allowed to cool and then dissolved in a 50/50 ethyl acetate/THF mix (250 cm³) and water (250 cm³). The aqueous layer was extracted using ethyl acetate (3 x 150 cm³), the combined organics were then washed with saturated sodium hydrogen carbonate solution (150 cm³), brine (150 cm³) and dried with Na₂SO₄. Filtration and evaporation gave a brown solid which when washed with cold methanol afforded the product (4.59g, 18%) as a white powder. M.p. 196-198°C (Found: C, 61.7; H, 5.1; N, 12.1. C₁₂H₁₂N₂O₃ requires C, 62.1; H, 5.2; N, 12.1%); νₘₐₓ (Nujol mull)/cm⁻¹ 3299, 3172, 2925, 2854, 1733, 1722, 1640, 1544, 1464, 1377, 1211, 1199, 1177, 761; δₜ (CD₃OD) 2.39-2.50 (1H, dd, J_ABX 17.6 and 5.4, CHCH₂) 2.79-2.93 (1H, J_ABX 17.6 and 9.5, CH₂) 3.55 (2H, s, CH₂Ph) 4.48 (1H, m, CHCH₂) 7.28 (5H, m, aryl); m/z (GC/MS) 232 (M)+, 207, 141, 118, 98, 91, 65, 51 and 43.

Method 2 (R/S)-N-Phenylacetylaspartic acid (36.65g, 0.146 mol) was partially dissolved in conc. ammonia solution (220 cm³). The reaction mixture was heated to reflux and the vapour allowed to escape from the reaction vessel. All solids eventually went into solution. When all the water had been evaporated, the molten residue was allowed to reach 180°C and was maintained at this temperature for a further 2.5 h. The mixture was cooled and dissolved in ethyl acetate (250 cm³) before the residue hardened. The organic layer was washed with saturated sodium hydrogen carbonate solution (150 cm³), brine (150 cm³) and dried with Na₂SO₄. Filtration and evaporation gave an impure brown solid product. Recrystallisation from ethyl acetate yielded the product (2.70g, 8%) as a white powder. M.p. 195-196°C. All other analyses were identical to those in Method 1.

Method 3 (R/S)-N-Phenylacetylaspartic acid (50.5g, 0.20 mol) was heated with finely ground ammonium carbonate (38.28g, 0.40 mol) as of Method 1. The product, after cooling the reaction mixture, was recrystallised from ethyl acetate yielding the product 21 (3.00g, 6%) as a white solid. M.p. 196-198°C. All other analyses were identical to those in Method 1.

Method 4 (R/S)-N-Phenylacetylaspartic acid (100.0g, 0.40 mol) was added to finely ground urea (47.9g, 0.80 mol) and the mixture heated with stirring to 170°C for 2 h. During this time, all solids dissolved and water vapour was observed condensing on the apparatus. The mixture was cooled and dissolved in a 50/50 water/ethyl acetate mix (500 cm³), separated, and the aqueous layer extracted with ethyl acetate (3 x 250 cm³). The combined organics were dried (MgSO₄), filtered and evaporated to give a light brown solid. Trituration using ether yielded the product (46.80g, 51%) as a white solid. M.p. 197-198°C. All other analyses were identical to those in Method 1.

(S)-N-Phenylacetylaspartic acid (30). The synthesis of 30 followed the same procedure as described for the racemate using phenylacetyl chloride (107.70g, 0.70 mol), 4M sodium hydroxide solution (171.5 cm³), (S)-aspartic acid (92.71g, 0.70 mol) and 2M sodium hydroxide solution (695 cm³). The reaction yielded 30 (104.41g, 59%) as a white crystalline solid. M.p. 116-117°C (Found: C, 57.5; H, 5.3; N, 5.7. C₁₂H₁₃NO₅ requires C, 57.4; H, 5.2; N, 5.6%); νₘₐₓ (Nujol mull)/cm⁻¹ 3412, 3332, 2965, 1738, 1706, 1647, 1545, 1467, 1345, 1289, 1244, 1193,
1155, 989, 732; δH (D6 DMSO) 2.65 (2H, s, CH2Ph) 3.48 (2H, s, CH2Ph) 4.55 (1H, JABX 7.0 and 6.3, CHCH2) 7.24 (5H, m, CH2Ph) 8.46 (1H, d, J 7.0, NH) 12.60 (2H, bs, 2 x CO2H).

(S)-N-Phenylacetylaspartimide (31). The synthesis of 31 followed the same procedure as for the racemate using method 4 and (S)-N-phenylacetylaspartic acid 30 (100.0g, 0.40 mol) and urea (47.9g, 0.80 mol). The impure product was triturated with hot methanol and filtered to give 31 (47.94g, 52%) as a white solid. M.p. 196-197°C (Found: C, 61.8; H, 5.1; N, 12.2. C 12H12N2O3 requires C, 62.1; H, 5.2; N, 12.1%); νmax (Nujol mull)/cm-1 3300, 3178, 3069, 2925, 2857, 1731, 1724, 1549, 1496, 1460, 1357, 1332, 1248, 1129, 1177, 776, 742, 637; δH (D6 DMSO) 2.40-2.50 (1H, dd, JABX 17.8 and 5.7, CHCH2) 2.80-2.93 (1H, dd, JABX 17.8 and 9.4, CHCH2) 3.45 (2H, s, CH2Ph) 4.41 (1H, m, CHCH2) 7.26 (5H, m, CH2Ph) 8.68 (1H, d, J 7.6, NH of amide) 11.23 (1H, bs, NH of imide); δC (D6 DMSO) 36.04 (CHCH2) 41.59 (CH2Ph) 49.59 (CH2) 126.36 (para aryl) 128.15 (2 x meta aryl) 128.29 (2 x ortho aryl) 135.59 (quaternary aryl) 170.40 (PhCH2CO); m/z (GC/MS) 232 (M)+, 208, 141, 91, 65, 51 and 43.

(S)-N-Phenylacetylaspartimido-α-hydroxyacetic acid (32). (S)-N-Phenylacetylaspartimido-α-hydroxyacetic acid 32 (25.00g, 108 mol) was dissolved in THF (250 cm3) at room temperature with stirring. To this was added glyoxylic acid (50 wt%) (31.91g, 0.216 mol) and the reaction mixture was heated to reflux for 3.5 h. Evaporation to dryness under reduced pressure produced a yellow oil which was crystallised from ethyl acetate by the dropwise addition of ether. This was recrystallised from methylated spirits yielding 32 (10.10g, 31%) as a white powder. M.p. 169-171°C (Found: C, 54.6; H, 4.5; N, 9.3. C14H14N2O6 requires C, 54.9; H, 4.6; N, 9.15%); νmax (Nujol mull)/cm-1 3380, 2922, 2853, 2721, 2603, 2531, 1786, 1745, 1720, 1714, 1614, 1600, 1541, 14657, 1392, 1348, 1236, 1109, 972, 735; δH (D6 DMSO) 2.42-2.52 (1H, dd, JABX 17.5 and 5.8, CHCH2) 2.95-3.10 (1H, dd, JABX 17.5 and 9.4, CHCH2) 3.49 (2H, s, CH2Ph) 4.65 (1H, d, J 7.6, NH); m/z (FAB+) 307 (M+H)+, 289, 261 and 233.

(S)-N-Phenylacetylaspartimido-α-chloroacetic acid (33). (S)-N-Phenylacetylaspartimido-α-chloroacetic acid 32 (2.5g, 8.2 mmol) was dissolved in dry THF (50 cm3) with stirring at room temperature under argon. The solution was cooled to 0°C before thionyl chloride (2.37g, 20.0 mmol) was added slowly over 20 mins. Evolution of an acidic gas was observed and the mixture was allowed to warm to room temperature overnight. Ice (10g) was added and the mixture stirred for a further 3 h. The solution was extracted using ethyl acetate (3 x 50 cm3), the organics dried (Na2SO4) filtered and evaporated to give a brown semi solid. Purification of this was achieved by recrystallisation from ethyl acetate/hexane to give 33 (0.80g, 30%) as a white solid. M.p. 189-191°C (Found: C, 51.7; H, 4.0; N, 8.75. C14H14ClN2O5 requires C, 51.8; H, 4.0; N, 8.6%); νmax (Nujol mull)/cm-1 2913, 2867, 2700, 1783, 1750, 1724, 1715, 1620, 1599, 1535, 1456, 1423, 1380, 1237, 1105, 973, 734; δH (D6 DMSO) 2.42-2.52 (1H, dd, JABX 17.5 and 5.8, CHCH2) 2.95-3.10 (1H, JABX 17.5 and 9.4, CHCH2) 3.49 (2H, s, CH2Ph) 4.63 (1H, m, CH2CH) 6.25 (1H, s, CHCl) 7.28 (5H, m, Ph) 8.82 (1H, d, J 7.8, NH); m/z (FAB+) 325 (M+H)+, 289, 280, 197 and 179.
Isatin-N-α-hydroxyacetic acid methyl ester (18). Isatin (14.7g, 0.10 mol) was dissolved in THF (200 cm³). Methyl glyoxylate (17.6g, 0.20 mol) was added and the solution was refluxed for 1.5 h. under nitrogen. The solution was cooled, the volume reduced and the resultant oil triturated under ether to give 18 (20.4g, 87%) as bright orange crystals. M.p. 135-138°C (Found: C, 56.5; H, 3.9; N, 6.1. C₁₁H₉NO₅ requires C, 56.2; H, 3.9; N, 6.0%); νₘₐₓ (Nujol mull)/cm⁻¹ 3450, 1760, 1730, 1620; δ_H (D₆ DMSO) 3.70 (3H, s, CH₃) 6.15 (1H, d, J 6.3, CHOH) 7.0-7.75 (4H, m, aromatics); m/z 235 (M⁺) 176 and 147.

Method D: Isatin-N-α-acetoxyacetic acid methyl ester (22). Isatin-N-α-hydroxyacetic acid methyl ester 18 (2.0g, 8.5 mmol) was dissolved in pyridine (10 cm³) with stirring and cooled to 0°C. Acetyl chloride (0.73g, 9.35 mmol) was added dropwise and stirred for 30 mins before being allowed to warm to room temperature for 3 h. The solution was then poured into ice water (50 cm³) which resulted in the precipitation of the product. This was filtered, washed with water and dried to give 22 (1.53g, 65%) as a dark yellow solid. M.p. 140-141°C. (Found: C, 56.1; H, 4.1; N, 4.9. C₁₃H₁₁NO₆ requires C, 56.3; H, 4.0; N, 5.05%); νₘₐₓ (Nujol mull/cm⁻¹ 1770, 1740, 1620, 1205; δ_H (CDCl₃) 2.15 (3H, s, OC(O)CH₃) 3.85, s, CO₂CH₃) 7.10 (1H, d, J 8.1, aryl) 7.12 (1H, s, NCH) 7.25 (1H, t, J 6.9, aryl) 7.64 (1H, d, J 7.65, aryl) 7.72 (1H, t, J 8.1, aryl); m/z 278 (M+H)⁺, 2.77, 218, 176 and 146.

Isatin-N-α-2,6-dichlorobenzoxyacetic acid methyl ester (23). Method D was used in the synthesis of 23 using isatin-N-α-hydroxy-acetic acid methyl ester 18 (2.0g, 8.5 mmol), pyridine (10 cm³) and 2,6-dichlorobenzoyl chloride (1.96g, 9.35 mmol) yielding 23 (2.25g, 62%) as a yellow powder. M.p. 105-109°C (Found: C, 53.3; H, 2.8; N, 3.6. C₁₈H₁₁Cl₂NO₆ requires C, 53.0; H, 2.7; N, 3.4%); νₘₐₓ (Nujol mull)/cm⁻¹ 1795, 1760, 1740, 1620; δ_H (D₆ DMSO) 3.90 (3H, s, CH₃) 7.0-7.90 (8H, m, aromatics and NCH); m/z 408 (M+H)⁺, 218, 190 and 146.

Isatin-N-α-phenylacetoxyacetic acid methyl ester (24). Method D was adopted using isatin-N-α-hydroxy-acetic acid methyl ester 18 (2.0g, 8.5 mmol), pyridine (10 cm³) and phenylacetyl chloride (1.45g, 9.35 mmol) yielding 24 (0.22g, 7%) as a pale yellow powder. M.p. 100-104°C; νₘₐₓ (Nujol mull)/cm⁻¹ 1775, 1740, 1675, 1620; δ_H (D₆ acetone) 3.75 (3H, s, CH₃) 3.90 (2H, s, CH₂) 7.10-7.70 (10H, m, aromatics and NCH); m/z 354 (M+H)⁺, 353, 294, 218 and 146.

Isatin-N-α-chloroacetic acid methyl ester (25). Isatin-N-α-hydroxy-acetic acid methyl ester 18 (0.50g, 2.13 mmol) was dissolved in THF (50 cm³) and cooled with stirring to -15°C. To this was added thionyl chloride (0.30g, 2.52 mmol) and triethylamine (0.26g, 2.52 mmol) dropwise over 10 mins. The solution was stirred for a further 3 h. until a white precipitate formed. The solution was diluted with CH₂Cl₂ (200 cm³) and washed with 0.1M HCl (2 x 50 cm³) and saturated brine (50 cm³). The organics were dried using MgSO₄, filtered and evaporated to dryness to leave an orange sticky solid. Trituration using chloroform gave 22 (0.16g, 30%) as bright orange crystals. M.p. 114-117°C; νₘₐₓ (Nujol mull)/cm⁻¹ 1760, 1740, 1620; δ_H (D₆ DMSO) 3.80 (3H, s, CH₃) 6.80-7.80 (7H, m, aromatics and NCH); m/z 255 (M⁺) (Cl³⁷), 253 (M⁺) (Cl³⁵), 218 and 146.

Isatin-N-α-bromoacetic acid methyl ester (26). Isatin-N-α-hydroxy-acetic acid methyl ester 18 (0.50g, 2.13 mmol) was dissolved in glacial acetic acid (10 cm³) with stirring at room
temperature. Bromine (0.34g, 2.13 mmol) then hydrogen bromide (0.17g, 2.13 mmol) were added dropwise to the solution and this was left to stir overnight. (Water (50 cm³) was added to the deep red solution and this extracted with chloroform (3 x 50 cm³). The organic layer was washed with dilute sodium metabisulphite solution (2 x 50 cm³), dried over MgSO₄, filtered and evaporated to dryness to give 26 (0.26g, 415) as an orange powder. M.p. 130-132°C; νₘₐₓ (Nujol mull)/cm⁻¹ 1740, 1620; δᵥ (D₆ DMSO) 3.70 (3H, s, CH₃) 7.10-7.90 (4H, m, aromatics and NCH); m/z 299 (M⁺ (Br₈¹), 297 (M⁺ (Br₇⁹), 240, 238, 227 and 225.

**Isatin-3-O-benzyl oxime** Isatin (14.7g, 0.1 mol) was dissolved in water (500 cm³) and O-benzylhydroxylamine hydrochloride (17.56g, 0.11 mol) was added and the solution refluxed for 1 h. A precipitate appeared in cooling which was filtered and recrystallised from toluene to give the product (14.83g, 59%) as bright yellow crystals on filtration. M.p. 160-161°C; νₘₐₓ (Nujol mull)/cm⁻¹ 1736, 1694, 1609; δᵥ (D₆ DMSO) 5.50 (2H, s, CH₂) 6.85 (1H, d, J 7.7, aryl) 7.00 (1H, t, J 7.7, aryl) 7.35-7.50 (6H, m, aromatics) 7.75 (1H, d, J 7.5, aryl) 10.85 (1H, s, NH); m/z 253 (M+H)+, 252 and 145.

**Isatin-N-α-hydroxyacetic acid-4-nitrobenzyl-ester-3-O-benzyl oxime** (27). p-Nitrobenzyl glyoxylate (10.0g, 44.1 mmol) was dissolved in toluene (150 cm³) and the solution refluxed for 1h. with a Dean and Stark apparatus attached. After cooling, isatin-3-benzyloxime (5.56g, 22.0 mmol) was added and the solution refluxed overnight. After cooling, the volume was reduced and the resultant oil triturated to give 27 (4.62g, 46%) as a yellow powder. M.p. 135-137°C (Found: C, 62.4; H, 4.2; N, 9.0. C₂₄H₁₉N₃O₇ requires C, 62.6; H, 4.1; N, 9.1%); νₘₐₓ (Nujol mull)/cm⁻¹ 3286, 1759, 1714, 1606, 1515, 1347; δᵥ (D₆ DMSO) 5.35 (2H, s, NO₂PhCH₂) 5.50 (2H, s, PhCH₂) 6.30 (1H, d, J 5.3, NCH) 7.10 (1H, d, J 8.0, aryl) 7.20 (1H, t, J 7.4, aryl) 7.30-7.60 (8H, m, aromatics) 7.95 (1H, d, J 7.0, aryl) 8.15 (2H, d, J 8.75, para aryl).

**Isatin-N-α-acetoxyacetic acid-4-nitrobenzyl-ester-3-O-benzyl oxime** (28). Method D was used in the formation of 28 using isatin-N-α-hydroxy-acetic acid-4-nitrobenzyl-ester-3-O-benzyl oxime (1.00g, 2.17 mmol), acetyl chloride (0.19g, 2.39 mmol) and pyridine (10 cm³) to give 28 (0.52g, 48%) as a pale yellow solid. M.p. 152-155°C (Found: C, 62.1; H, 4.3; N, 8.3. C₂₆H₂₁N₃O₈ requires C, 62.0; H, 4.2; N, 8.35%); νₘₐₓ (Nujol mull)/cm⁻¹ 1780, 1620, 1515, 1347; δᵥ (D₆ DMSO) 2.20 (3H, s, CH₃) 5.40 (2H, s, CO₂CH₂) 5.55 (2H, s, CO₂CH₂ON) 7.05 (1H, d, J 7.9, aryl) 7.25 (1H, t, J 7.25, aryl) 7.25 (1H, s, NCH) 7.35-7.55 (8H, m, aromatics) 7.95 (1H, d, J 7.7, aryl) 8.15 (2H, d, J 8.7, para aryl); m/z 504 (M+H)^+.

**Isatin-N-α-2,6-dichlorobenzoxyacetic acid-4-nitrobenzyl-ester-3-O-benzyl oxime** (29). Method D was used in the formation of 29 using isatin-N-α-hydroxy-acetic acid-4-nitrobenzyl-ester-3-O-benzyl oxime (1.00g, 2.17 mmol), 2,6-dichlorobenzoyl chloride (0.50g, 2.39 mmol) and pyridine (10 cm³) to give 29 (0.35g, 25%) as a yellow solid. M.p. 110-112°C (Found: C, 58.4; H, 3.6; N, 6.5. C₃₁H₂₁Cl₂N₃O₈ requires C, 58.7; H, 3.3; N, 6.6%); νₘₐₓ (Nujol mull)/cm⁻¹ 1736, 1606, 1516; δᵥ (D₆ DMSO) 5.40 (2H, s, CO₂CH₂ON) 6.99 (1H, d, J 7.8, aryl) 7.05 (1H, t, J 7.5, aryl) 7.25 (1H, t, J 6.9, aryl) 7.35-7.80 (11H, m, aromatics) 7.95 (1H, d, J 7.8, aryl) 8.15 (2H, d, J 8.0, para aryl).
Kinetic Studies. The second-order rate constants for alkaline hydrolysis of the \( \gamma \)-lactams were obtained as previously described.\textsuperscript{12} The inhibition studies were also undertaken as described elsewhere.\textsuperscript{24}

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References