

# Ultrasound-assisted synthesis of imidazo[1,2-*a*]pyridines and sequential one-pot preparation of 3-selanyl-imidazo[1,2-*a*]pyridine derivatives

Beatriz M. Vieira,<sup>a</sup> Nathalia Padilha,<sup>a</sup> Nelson M. Nascimento,<sup>a</sup> Gelson Perin,<sup>a</sup> Diego Alves,<sup>a</sup> Ricardo F. Schumacher,<sup>\*a,b</sup> and Eder J. Lenardão<sup>\*a</sup>

<sup>a</sup> LASOL - CCQFA, Universidade Federal de Pelotas, UFPel-P.O. Box 354,96010-900, Pelotas, RS, Brazil <sup>b</sup> Departamento de Química, Universidade Federal de Santa Maria, UFSM, 97105-900, Santa Maria, RS, Brazil E-mail: <u>lenardao@ufpel.edu.br</u>, <u>ricardo.schumacher@ufsm.br</u>

### Dedicated to Professor Lorenzo Testaferri in the occasion of his 75<sup>th</sup> birthday

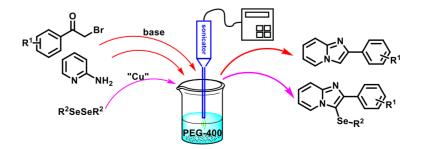
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#### Abstract

A simple and rapid method to synthesize imidazo[1,2-*a*]pyridines starting from 2-aminopyridine and 2bromoacetophenone derivatives under ultrasonic irradiation was developed. This protocol tolerates a wide range of 2-bromoacetophenone derivatives to produce a variety of imidazo[1,2-*a*]pyridines in good to excellent yields. Additionally, the one-pot preparation of 3-(organylselanyl)imidazo[1,2-*a*]pyridines via a sequential method is presented. In this case, different diorganyl diselenides were used as starting materials to afford the corresponding coupling products in excellent yields and short reaction times under sonication. The reactions were conducted in PEG-400, a cheap and nontoxic solvent, compatible with the ultrasound conditions in an environmentally benign process.



Keywords: Aminopyridine, diorganyl diselenides, imidazo[1,2-a]pyridines, PEG-400, ultrasound irradiation

#### Introduction

N-Heterocycles are structures found in natural and synthetic organic compounds relevant in many chemical and biological aspects.<sup>1</sup> Among them, the imidazo[1,2-*a*]pyridines are one of the most studied class of heterocycles, due to their therapeutic use as antiviral,<sup>2</sup> antituberculotic,<sup>3</sup> antibacterial,<sup>4</sup> antipsychotic<sup>5</sup> and antitumor agents.<sup>6,7</sup> In this way, the imidazo[1,2-*a*]pyridine scaffold is present in several established leading drugs, such as Alpidem, Necopidem, Olprinone, Saripidem, Zolimidine and Zolpidem.

In recent years, the synthesis and incorporation of different groups to the imidazo[1,2-*a*]pyridine unity, such as halogen,<sup>8,9</sup> aryl,<sup>10,11</sup> formyl<sup>12,13</sup> and chalcogen<sup>14-19</sup> have attracted the attention of synthetic organic chemists, aiming to prospect for new drug candidates. Most of these methodologies, however, present drawbacks, such as tedious multistage synthesis and workup, the generation of large amounts of waste and a high effective cost. Some of these problems have been met, at least in part, by the use of multicomponent protocols, alternative energy sources and recyclable catalyst systems.<sup>20-24</sup> For instance, Heibel and co-workers described a one-pot sequential microwave-assisted synthesis of 2-aryl-imidazo[1,2-*a*]pyridines in PEG-400,<sup>21</sup> while Wei and co-workers used CeCl<sub>3</sub>.7H<sub>2</sub>O/NaI as a catalytic system to produce 3-sulfenylimidazo[1,2-*a*]pyridines in ethanol under oxygen atmosphere.<sup>22</sup> More recently, Allahabadi and co-workers developed a copper-catalyzed four-component synthesis of 2,3-disubstituted-imidazo[1,2-*a*]pyridines,<sup>23</sup> while Zhang and Jiang described a solvent-free microwave-assisted multicomponent synthesis of 3-substituted-imidazo[1,2-*a*]pyridines.<sup>24</sup> Although the important improvement achieved by these methods, there is a research gap in the development of efficient and environmentally friendly protocols to access imidazo[1,2-*a*]pyridines directly attached to an organoselenium group.

Selenium-containing heterocycles are relevant in biochemical and pharmacological processes since several organoselenium compounds are known to exhibit interesting biological properties.<sup>25,26</sup> Moreover, many organoselenium compounds have been used in a wide spectrum of organic reactions as precursors in the construction of functional materials.<sup>27,28</sup> Thus, the synthesis and biological evaluation of selenium containing imidazo[1,2-*a*]pyridines are interesting fields of research, which combine at least two bioactive moieties in one molecule as an effective strategy for designing new pharmacologically promising drugs.<sup>4,29-32</sup>

In the last years, sonochemistry has established itself as an indispensable tool in drug discovery and green organic synthesis.<sup>33-37</sup> Reactions promoted by ultrasound are faster due to the turbulent flow of the liquid phase caused by the cavitation, which enhances the mass transfer in heterogeneous systems.<sup>38,39</sup> In this sense, due to our interest in the development of green procedures for the synthesis of biologically active organochalcogen-containing *N*-heterocycles,<sup>18,29,31,32,40,41</sup> we describe here the ultrasound-promoted syntheses of imidazo[1,2-*a*]pyridines and 3-selanyl-imidazo[1,2-*a*]pyridines via a sequential one-pot reaction.

#### **Results and Discussion**

The initial experiments to optimize the reaction conditions were performed using 2-aminopyridine **1** and 2-bromoacetophenone **2a** as model substrates to establish the best reaction conditions to prepare 2-phenylimidazo[1,2-*a*]pyridine **3a** under ultrasound irradiation. In this study we have evaluated the influence of different bases and solvents, as well as the reaction time, as described in Table 1.

We started our studies by the reaction of 2-aminopyridine **1** (0.5 mmol) with 2-bromoacetophenone **2a** (0.5 mmol) using NaHCO<sub>3</sub> (1 equiv) as base in PEG-400 (1.0 mL) as the solvent. After sonication for 30 min

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(60% of amplitude, 20 kHz), 2-phenyl-imidazo[1,2-*a*]pyridine **3a** was obtained in 78% isolated yield (Table 1, entry 1). With this result in hand, we turned our attention to testing different environmentally friendly solvents, such as glycerol, dimethylsulfoxide and ethanol (entries 2-4). PEG-400 proved to be the most suitable solvent for this reaction.

Aiming to check the necessity of base, a base-free reaction was performed, but in this case, only 39% yield of the expected product **3a** was obtained after 30 min (Table 1, entry 5). From this point, a screening through different bases was conducted (Table 1, entries 6-12), which revealed that  $K_2CO_3$  was superior under the ultrasound conditions (entry 7). An improvement in the reaction yield was achieved when the amount of  $K_2CO_3$  was raised from 1.0 to 1.5 equiv, giving **3a** in 92% yield (Table 1, entry 14). To our delight, when the reaction was sonicated for only 15 min, a similar yield of **3a** was obtained (94%), proving that the efficient energy-transfer by the ultrasonic irradiation can promote a fast reaction.

	$ \begin{array}{c}                                     $	US, time	a
Entry	Base (equiv)	Solvent	Yield (%)
1	NaHCO <sub>3</sub> (1)	PEG-400	78
2	NaHCO <sub>3</sub> (1)	Glycerol	36
3	NaHCO₃ (1)	DMSO	67
4	NaHCO <sub>3</sub> (1)	Ethanol	11
5	-	PEG-400	39
6	KOH (1)	PEG-400	28
7	K <sub>2</sub> CO <sub>3</sub> (1)	PEG-400	81
8	Cs <sub>2</sub> CO <sub>3</sub> (1)	PEG-400	33
9	Et <sub>3</sub> N (1)	PEG-400	22
10	K <sub>3</sub> PO <sub>4</sub> (1)	PEG-400	63
11	<sup>t</sup> BuOK (1)	PEG-400	27
12	KF/Al <sub>2</sub> O <sub>3</sub> (1)	PEG-400	NR
13	K <sub>2</sub> CO <sub>3</sub> (1.2)	PEG-400	87
14	K <sub>2</sub> CO <sub>3</sub> (1.5)	PEG-400	92
15 <sup>b</sup>	K <sub>2</sub> CO <sub>3</sub> (1.5)	PEG-400	94

<sup>a</sup> Reactions performed using 2-aminopyridine **1** (0.5 mmol), 2-bromoacetophenone **2a** 

(0.5 mmol) and base in 1 mL of solvent, under ultrasonic irradiation (60% amplitude,

20 kHz) for 30 min. <sup>b</sup> Reaction performed for 15 min. NR = No Reaction

Next, with the best reaction conditions found, the substrate scope was evaluated with respect to differently substituted 2-bromoacetotophenones **2a**-**h**. These reactions led to the corresponding imidazo[1,2-*a*]pyridines **3a**-**h** (Table 2).

An inspection in the results of Table 2 shows that the expected imidazo[1,2-*a*]pyridines **3a-h** were obtained in good to excellent yields at reaction times no longer than 30 min. It can also be observed that the presence of electron-withdrawing or electron-releasing groups directly attached to the aryl moiety of the

**Table 2.** Imidazo[1,2-*a*]pyridines **3** synthesized under US <sup>*a*</sup>

	$NH_2 + R\frac{1}{1}$	O Br <u>K<sub>2</sub>CO<sub>3</sub> (1.5 equiv)</u> PEG-400, US <b>2a-h</b>	N 3a-h	Ĩ ∕∕R
Entry	Bromoacetophenone <b>2</b>	Imidazo[1,2- <i>a</i> ]pyridine <b>3</b>	Time (min)	Yield (%)
1	Br 2a	Sa	15	94
2	CI Br 2b	Sb	30	78
3	F 2c	Sc North	30 (15)	70 (88) <sup>b</sup>
4	CI 2d	CI Sd	25	79
5	O <sub>2</sub> N Br	$ \begin{array}{c}                                     $	30 (30)	55 (80) <sup>ь</sup>
6	H <sub>3</sub> C Br 2f	Grand Sf	30	85
7	H <sub>3</sub> CO Br		30 (30)	67 (90) <sup>ь</sup>
8	O OCH <sub>3</sub> 2h	$ \begin{array}{c}                                     $	30	98

<sup>a</sup> Reactions were performed using **1** (0.5 mmol), **2** (0.5 mmol), K<sub>2</sub>CO<sub>3</sub> (1.5 equiv, 0.75 mmol) in PEG-400 (1 mL) under ultrasonic irradiation (60% of amplitude). <sup>b</sup> Number in parenthesis refers to the reaction using 2 equiv of **1** (1.0 mmol).

Vieira, B. M. et al.

For example, the presence of fluorine or nitro groups in the phenyl ring of the 2-bromoacetophenones **2c** (R = 4-R) and **2e** ( $R = 4-NO_2$ ), negatively influenced the reaction, and the respective products **3c** and **3e** were obtained in 70% and 55% yields (Table 2, entries 3 and 5). A similar decrease in reactivity was observed starting from the electron-rich 2-bromo-4'-methoxyacetophenone (**2g**, R = 4-OMe), and the expected product **3g** was isolated in 67% yield using the optimal conditions. Fortunately, better yields were obtained from these recalcitrant ketones when an excess of 2-aminopyridine **1** (2 equiv) was used, raising the yields of **3c**, **3e** and **3g** to 88%, 80% and 90%, respectively, while reducing the reaction time to 15 min in the case of **3c** (Table 2, values in parenthesis). Notably, these reactions do not suffer from steric effects of the substituents at 2-bromoacetophenone counterpart **2**. For instance, the reactions using the sterically constrained 2-bromo-1-(2-chlorophenyl)ethan-1-one **2d** and 2-bromo-1-(2-methoxyphenyl)ethan-1-one **2h**, afforded the corresponding products **3d** and **3h** in 79% and 98% yields, which are close to those obtained starting from the unsubstituted 2-bromoacetophenone **2a** (Table 2, entries 4 and 8). The possibility of obtaining chloro-substituted imidazo-[1,2-*a*]pyridines **3b** and **3d** is noteworthy, once it allows the possibility of future reactions like TM-catalyzed couplings.

As mentioned before, the combination of N-heterocycles with organochalcogen moieties is an interesting strategy to prospect for new compounds with increased pharmacological activities.<sup>29-32</sup> In this line, the development of alternative, green methods based in ultrasound synthesis of organochalcogen-containing N-heterocycles under non-conventional reaction media is an interesting field of research, in consideration of economic, environmental and health issues.<sup>42</sup> Still, the growing interest in these compounds as promising new drugs, functional materials, catalysts and synthetic intermediates collaborate to reinforce the importance of this class of compounds.<sup>25,26</sup> Thus, to extend the application of our US-assisted protocol, we decided to study the possibility of performing an one-pot sequential direct selanylation reaction of the pre-formed imidazo[1,2-*a*]pyridines **3**, aiming to prepare selanylimidazopyridines **5** (Table 3). We started our investigations using the reaction to form imidazo[1,2-*a*]pyridine **3a** (Table 2, entry 1), with diphenyl diselenide **4a** as the selenylating species. After the preformation of **3a** (15 min under sonication), diphenyl diselenide **4a** (0.6 equiv) and the catalyst were added in the same vessel, without isolation of **3a**, and the sonication was continued for additional 2 h (Table 3).

	NH NH	+    <sub>Br</sub>	CO <sub>3</sub> (1.5 equiv) PEG-400, US 15 min	N N 3a not isolated	PhSeSePh <b>4a</b> Conditions	5a SePh	
Entry	Additive	Amount (equiv)	Yield (%) <sup>b</sup>	Entry	Additive	Amount (equiv)	Yield (%) <sup>b</sup>
1	none <sup>c</sup>	-	NR <sup>d</sup>	6	Cul	2	42
2	NBS	1.2	46	7	KI/CuSO <sub>4</sub>	2/2	95
3	FeCl₃	1	31	8	KI/CuSO <sub>4</sub>	1/1	41
4	I <sub>2</sub>	2	80	9	KI	2	NR <sup>d</sup>
5	CuBr	2	57	10	CuSO <sub>4</sub>	2	38

Table 3. Optimization of the reaction	conditions for the	direct selenylation o	f <b>3a</b> ª
	Г	Г	

<sup>*a*</sup> Reactions were performed using initially **1** (0.5 mmol), **2a** (0.5 mmol),  $K_2CO_3$  (0.75 mmol, 1.5 equiv.) in PEG-400 (1.0 mL) under ultrasonic irradiation (60% of amplitude) for 15 min. After that, in the same vessel, **4a** (0.3 mmol) and the additive were added. The ultrasonic irradiation was continued for additional 2 h. <sup>*b*</sup> Yield is given for isolated product after the sequential two step one-pot procedure. <sup>*c*</sup> An additional 1 equiv (0.5 mmol) of  $K_2CO_3$  was added with the diselenide **4a**. <sup>*d*</sup> At the end of the reaction, **3a** and **4a** were recovered. NR = no reaction.

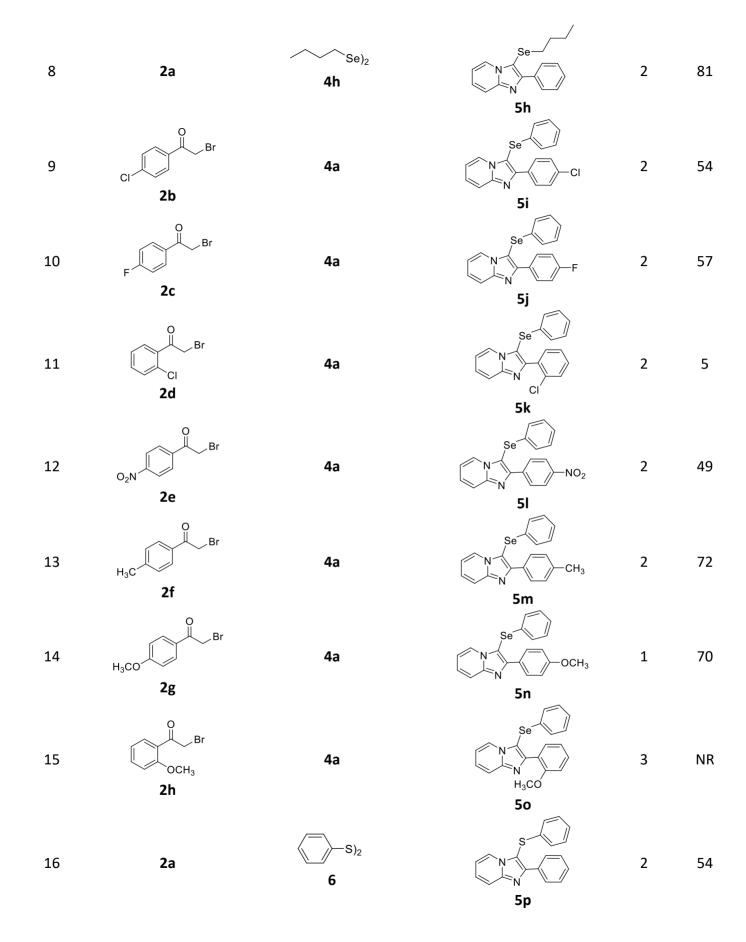
Initially, the reaction was tested just by adding PhSeSePh **4a** and an additional amount of base (K<sub>2</sub>CO<sub>3</sub>; 0.5 mmol) in the reaction vessel containing the preformed imidazo[1,2-*a*]pyridine **3a**. Unfortunately, none of the expected product **5a** was obtained after 2 h of sonication and the imidazo[1,2-*a*]pyridine **3a** and diselenide **4a** were recovered (Table 3, entry 1). Then, we decided to investigate the influence of different well-known reagents able to produce *in situ* electrophilic selenium species such as NBS, FeCl<sub>3</sub>, I<sub>2</sub>, KI and copper salts (Table 3, entries 2-7). Analysis of the preliminary reactions showed that moderate to excellent yields of the expected product **5a** were obtained under US in PEG-400. Remarkably, when the reaction was carried out using a mixture of KI/CuSO<sub>4</sub> (2/2 equiv), the expected **5a** was obtained in 95% yield after 2 h (Table 3, entry 7). In order to evaluate the optimal amount of KI/CuSO<sub>4</sub>, a reaction was conducted using 1 equiv of this mixture of salts. However, the yield of **5a** was dramatically reduced to 41% (Table 3, entry 8). When the reaction was performed using only KI, consumption of the starting materials was not observed (Table 3, entry 9), while using CuSO<sub>4</sub> alone produced only 38% yield of product **5a** (Table 3, entry 10). Thus, the best conditions in the one-pot reaction to prepare 3-phenylselanyl-imidazo[1,2-*a*]pyridine **5a** requires sonication of a mixture of the preformed imidazo[1,2-*a*]pyridine **3a** in PEG-400 with diphenyl diselenide **4a** (0.6 equiv) in the presence of KI/CuSO<sub>4</sub> (2 equiv) for 2 h (Table 3, entry 7).

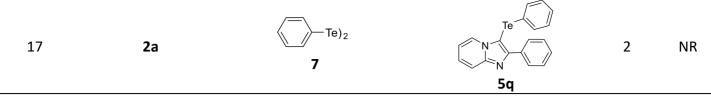
The versatility and usefulness of our ultrasound-activated one-pot protocol was then extended to different diorganyl dichalcogenides **4** and imidazo[1,2-*a*]pyridine derivatives **3** (Table 4). As a general trend, it was observed that most of the reactions proceeded smoothly with good results. The direct chalcogenation showed to be general with respect to a wide array of diorganyl diselenides **4a**-**h**, regardless their substitution patterns (Table 4, entries 1-8).

# Table 4. Synthesis of 3-selenyl-imidazo[1,2-a]pyridines 5a-q.<sup>a</sup>

NH N + 1		K <sub>2</sub> CO <sub>3</sub> PEG-400, US 15-30 min <b>3a-h</b>	$\begin{bmatrix} (R^2Se)_2 4a-h \\ CuSO_4/KI \\ US \end{bmatrix}$	SeR <sup>2</sup>	- ≫ <sub>R1</sub>
Entry	2-Bromoketone <b>2</b>	dichalcogenide 4	Product <b>5</b>	Time (h)	Yield (%)ª
1	Br 2a	Se) <sub>2</sub> 4a	Se N N Sa	2	95
2	2a	CI-Se) <sub>2</sub> 4b	Se CI	1	90
3	2a	F-Se) <sub>2</sub> 4c	Se F Se Se F Sc Sc	1	71
4	2a	F <sub>3</sub> C <b>4d</b>	Se CF <sub>3</sub> 5d	2	70
5	2a	H <sub>3</sub> CO-Se) <sub>2</sub> <b>4e</b>	Se OCH <sub>3</sub>	2	73
6	2a	H <sub>3</sub> C-Se) <sub>2</sub> 4f	Se N N Sf	2	54
7	2a	$H_3C - Se)_2$ CH <sub>3</sub> CH <sub>3</sub>	$H_3C$ $H_3C$ $H_3C$ K K K K K K K K K K	2	38

5g





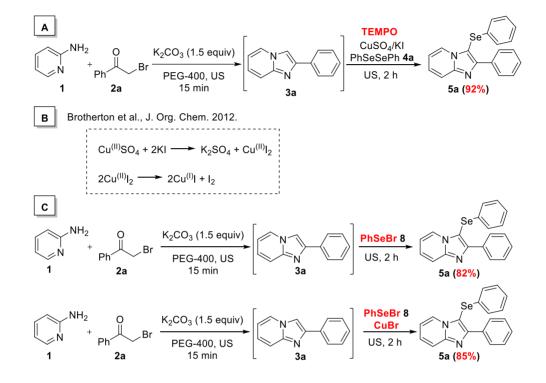
<sup>*a*</sup> Reaction performed using **2** (0.50 mmol), **4** (0.50 mmol),  $K_2CO_3$  (0.75 mmol) in PEG-400 (1 mL) under US for 15-30 min. Then, diorganyl diselenide **4** (0.3 mmol) and the mixture CuSO<sub>4</sub>/KI (1 mmol each) were added. The reactions were monitored by TLC. NR = no reaction.

The electron-deficient diaryl diselenides **4b** (R = 4-Cl), **4c** (R = 4-F) and **4d** (R = 3-CF<sub>3</sub>) reacted under the optimal conditions with **3a** (generated *in situ*) to afford the expected products **5b**-**d** 70-90% yields. The same reactivity was showed by electron-rich diaryl diselenides **4e** (R = 4-OMe) and **4f** (R = 4-CH<sub>3</sub>), that delivered the respective products **5e** and **5f** in 73% and 54% yields (Table 4, entries 2-6). On the other hand, the bulkier dimesityl diselenide **4h** was less reactive, due to the strong steric hindrance effect; the 3-mesitylselanyl-imidazo[1,2-*a*]pyridine **5h** was obtained in only 38% yield (Table 4, entry 7). The methodology was successfully extended to the aliphatic dibutyl diselenide **4h**, affording the product **5h** in 81% after 2 hours (Table 4, entry 8).

We evaluated the scope of the reaction regarding to a range of imidazo[1,2-a]pyridines **3**, prepared in situ from different 2-bromoacetophenone derivatives 2. It was observed that the reaction is very sensitive to steric effects due to substituents in the 2-bromoacetophenone counterpart **2**. This low reactivity was not observed in the synthesis of the respective imidazo [1,2-a] pyridines **3d** (R = 2-Cl) and **3h** (R = 2-OMe); therefore it cannot be attributed to this step of the one-pot synthesis. For instance, the ortho-chloro-substituted acetophenone 2d gave only 5% yield of the expected product 5k, while the reaction using ortho-methoxy derivative 2h failed completely (Table 4, entries 11 and 15). The presence of electron-withdrawing group at the para-position in the phenyl ring of the 2-bromoacetophenone negatively affects the reaction, and products **5i** (R = 4-Cl), **5j** (R = 4-Cl), **5j** (R = 4-Cl), **5j** (R = 4-Cl), **5i** (R = 4-Cl), 4-F) and **5I** (R = 4-NO<sub>2</sub>) were obtained in 54%, 57% and 49% yields, respectively (Table 4, entries 9, 10 and 12). These yields are slightly lower than those observed for 2-bromoacetophenone derivatives 2f (R = 4-CH<sub>3</sub>) and 2g (R = 4-OMe), which afforded the respective selenylated products 5m and 5n in 72% and 70% yields (Table 4, entries 13 and 14). Besides the diorganyl diselenides 4, some reactions were also conducted using diphenyl disulfide 6 and diphenyl ditelluride 7 as chalcogen sources. Diphenyl disulfide 6 led to the formation of 3phenylsulfanyl-imidazo[1,2-a]pyridine **5p** in 54% yield. The ditelluride analogue **7**, however, did not react under the optimal reaction conditions (Table 4, entries 16 and 17). Interesting, the reactions to obtain products 5c and 5n were interrupted after 1 h of sonication, once it was observed decomposition of the products after a longer reaction time (Table 4, entries 3 and 14).

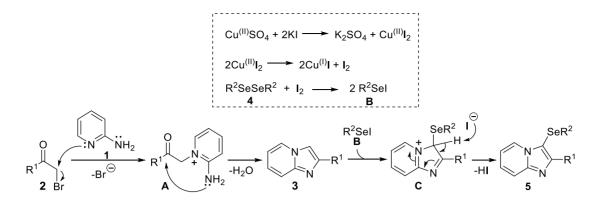
To investigate the possible mechanism involved in this reaction, some control experiments were performed, as depicted in Scheme 2. In order to rule out the hypothesis that the reaction could proceed through a radical mechanism, we performed the reaction under the standard conditions but in the presence of the radical scavenger 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) (Scheme 2-A). After 2 h of reaction, the product **5a** was obtained in 92% yield, hinting that a radical pathway is not involved in the reaction. In addition, and according to the conditions described by Brotherton and coworkers,<sup>43</sup> KI is believed to react with CuSO<sub>4</sub> to generate K<sub>2</sub>SO<sub>4</sub> and Cul<sub>2</sub>. Copper(II) iodide is very unstable and disproportionates to Cul and I<sub>2</sub> (Scheme 2-B). We believe that these transformations could be happening in a similar way under the sonication process. Plausibly, both iodine and copper(I) iodide could be responsible for generate electrophilic species of selenium, as demonstrated by the results from Table 3, entries 4 and 6. Following, two additional control

reactions were performed to investigate the possible formation of  $R^2Se-X$  (X = halogen) in the reaction medium and the influence of the generated copper salts in the reactivity (Scheme 2-C). Firstly, the commercially available phenylselanyl bromide **8** was used instead of PhSeSePh/CuSO<sub>4</sub>/KI under the standard US conditions, giving the product **5a** in 82% yield. Then, to investigate the possible participation of the copper salts present in the reaction media, CuBr was joined to the reagents. After 2 h of sonication, the reaction exhibited virtually the same efficiency, indicating that the halogenated species of selenium (Scheme 3, intermediate **B**) is the main active selenium species in the present protocol.



Scheme 2. Control experiments.

In this way, according to the results obtained and with the support of the literature, we can propose the following mechanism for the formation of the selenylated product **5** by the one-pot reaction (Scheme 3). Initially, 2-aminopyridine **1** nucleophilicaly attacks 2-bromoacetophenone **2** to generate the pyridinium intermediate **A**. Then, intermediate **A** undergoes an intramolecular attack of the NH<sub>2</sub> to produce imidazo[1,2-*a*]pyridine **3**, with water release. In the next step, molecular iodine, formed in the medium,<sup>43,44</sup> reacts with diorganyl diselenide **4** to form the electrophilic species of selenium **B**. Finally, **B** undergoes a nucleophilic attack by an electron pair of imidazo[1,2-*a*]pyridine **3** to afford intermediate **C**, that releases a proton to afford the product **5**.



Scheme 3. Plausible mechanism.

#### Conclusions

In conclusion, a series of imidazo[1,2-*a*]pyridines were synthesized via ultrasound-mediated reaction of 2aminopyridine with 2-bromoacetophenone derivatives. This strategy was efficiently extended to the one-pot synthesis of 3-selanylimidazo[1,2-*a*]pyridines, by the reaction of imidazo[1,2-*a*]pyridines formed in situ with diorganyl diselenide in the presence of copper(II) sulfate/potassium iodide under sonication. The advantages in using ultrasound include short reaction times, mild reaction conditions and ease of operation. All the reactions were conducted in air atmosphere using PEG-400 as a non-toxic and environmentally benign solvent.

#### **Experimental Section**

General. Reactions were monitored by TLC which was carried out on Merck silica gel (60 F<sub>254</sub>) by using UV light as visualizing agent and 5% vanillin in 10% H<sub>2</sub>SO<sub>4</sub> and heat as developing agents. Column chromatography was performed using silica gel (70-230 mesh). Proton nuclear magnetic resonance spectra (<sup>1</sup>H NMR) were obtained at 400 MHz on Bruker DPX 400 spectrometer. Spectra were recorded in CDCl<sub>3</sub> and CDCl<sub>3</sub>/DMSO- $d_6$  solutions. Chemical shifts are reported in ppm, referenced to tetramethylsilane (TMS) as the external reference. Proton coupling patterns are described as singlet (s), doublet (d), triplet (t), double doublet (dd), double triplet (dt) and multiplet (m). Coupling constants (J) are reported in Hertz. Carbon-13 nuclear magnetic resonance spectra (<sup>13</sup>C NMR) were obtained at 100 MHz on a Bruker DPX 400 spectrometer. Chemical shifts are reported in ppm, referenced to the solvent peak of CDCl<sub>3</sub>. Low-resolution mass spectra (MS) were obtained with a Shimadzu GC-MS-QP2010P mass spectrometer. GC analysis were conducted on a RESTEC RTX-5MS capillary column (30 m, 0.25 mm id, 0.25 µm film thickness) using the products dissolved in ethyl acetate with the following conditions: Injected sample volume was 1.0 µL; He constant flow, 54.1 mL/min; initial inlet temperature, 40 °C ramped to 72 °C at 10 °C/min followed by a 5 °C/min ramp to 100 °C (held for 10 min) and 10 °C/min to 280 °C and held for 20 min (total run time: 56.8 min). The ultrasound-promoted reactions were performed using a Cole Parmer-ultrasonic processor Model CPX 130, with a maximum power of 130 W, operating at amplitude of 60% and a frequency of 20 kHz. The temperature of the reaction under US was monitored using an Incoterm digital infrared thermometer Model Infraterm (Brazil). High-resolution mass spectra (HRMS) were obtained for all compounds on a LTQ Orbitrap Discovery mass spectrometer (Thermo Fisher Scientific). This hybrid system meets the LTQ XL linear ion trap mass spectrometer and an Orbitrap mass analyzer. The experiments were performed *via* direct infusion of sample (flow: 10  $\mu$ L/min) in the positive-ion mode using electrospray ionization. Elemental composition calculations for comparison were executed using the specific tool included in the Qual Browser module of Xcalibur (Thermo Fisher Scientific, release 2.0.7) software. Melting point (mp) values were measured in a Marte PFD III instrument with a 0.1 °C precision.

**General procedure for the synthesis 2-arylimidazo[1,2-***a***]pyridines 3a-h.** To a 10 mL vial was added a mixture of 2-aminopyridine **1** (0.5 mmol), 2-bromoacetophenone derivative **2** (0.5 mmol) and K<sub>2</sub>CO<sub>3</sub> (0.103 g, 0.75 mmol) in PEG-400 (1 mL). Then, the ultrasonic probe was introduced in the flask and the mixture was sonicated at 20 kHz and 60% of amplitude.The progress of the reaction was monitored by TLC and after the time indicated in Table 2, the reaction mixture was received in water (10 mL), extracted with ethyl acetate (3 x 5 mL), dried over MgSO<sub>4</sub> and concentrated under vacuum. The residue was purified by column chromatography using silica gel and a mixture of ethyl acetate/hexanes as the eluent.

**2-Phenylimidazo[1,2-***a***]pyridine (3a).** White solid, mp 137-139 °C; lit.<sup>45</sup> 136-137 °C. Yield: 0.091 g (94%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz): δ = 8.1 (d, *J* 6.8 Hz, 1H), 8.0 – 7.9 (m, 2H), 7.9 (s, 1H), 7.7 (d, *J* 9.1 Hz, 1H), 7.5 (t, *J* 7.5 Hz, 2H), 7.4 – 7.3 (m, 1H), 7.2 – 7.1 (m, 1H), 6.8 (dt, *J* 6.8, 1.2, 0.1 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz): δ = 145.7, 145.6, 133.6, 128.8, 128.0, 126.1, 125.6, 124.8, 117.5, 112.5, 108.1 .MS: *m/z* (rel. int.) 194 (100), 167 (7.5), 116 (3.3), 78 (25).

**2-(4-Chlorophenyl)imidazo[1,2-***a***]pyridine (3b)**. White solid, mp 213-115 °C; Lit.:<sup>1</sup> 207-209 °C. Yield: 0.088 g (78%); NMR <sup>1</sup>H (CDCl<sub>3</sub>/DMSO-*d*<sub>6</sub>, 400 MHz): δ 8.4 (t, *J* 5.6 Hz, 1H), 8.2 (d, *J* 7.7 Hz, 1H), 7.9 (dd, *J* 11.6, 7.9 Hz, 2H), 7.5 (t, *J* 10.0 Hz, 1H), 7.4 (dd, *J* 11.4, 7.8 Hz, 2H), 7.2 (dt, *J* 10.3, 5.1 Hz, 1H), 6.8 (q, *J* 6.0, 5.0 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>/DMSO-*d*<sub>6</sub>, 100 MHz): δ = 143.6, 142.0, 131.2, 131.0, 127.1, 125.6, 125.0, 123.6, 115.1, 110.9, 107.6. MS: *m/z* (rel. int.) 228 (100), 193 (13), 115 (3.5), 78 (38).

**2-(4-Fluorophenyl)imidazo[1,2-***a***]pyridine (3c**). White solid, mp 164-165; Lit.:<sup>21</sup> 163.1-163.4. Yield: 0.093 g (88%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.1 (dt, *J* 6.8, 1.2 Hz, 1H), 7.9 – 7.9 (m, 2H), 7.8 (s, 1H), 7.6 (dq, *J* 9.1, 1.0 Hz, 1H), 7.2 – 7.1 (m, 3H), 6.7 (td, *J* 6.7, 1.2 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 162.6 (d, *J* 246.6 Hz), 145.6, 144.8, 129.8, 127.6 (d, *J* 8.5 Hz), 125.5, 124.8, 117.3, 115.6 (d, *J* 21.4 Hz), 112.4, 107.7. MS: *m/z* (rel. int.) 212 (100), 193 (1.9), 96 (3.5), 78 (36).

**2-(2-Chlorophenyl)imidazo[1,2-***a***]pyridine (3d)**. White solid, mp 78.6-79.9 °C; Yield: 0.090 g (79%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.4 – 8.3 (m, 2H), 8.1 (dt, *J* 6.8, 1.2 Hz, 1H), 7.6 (dq, *J* 9.2, 1.0 Hz, 1H), 7.5 (dd, *J* 8.0; 1.3 Hz, 1H), 7.4 (td, *J* 7.6, 1.3 Hz, 1H), 7.3 – 7.2 (m, 1H), 7.2 (ddd, *J* 9.2, 6.7, 1.3 Hz, 1H), 6.8 (td, *J* 6.8, 1.2 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 144.5, 141.8, 132.3, 131.7, 131.0, 130.3, 128.6, 127.1, 125.8, 124.9, 117.5, 112.5 .MS: *m/z* (rel. int.) 228 (100), 193 (14.6), 115 (2.8), 78 (21).

**2-(4-Nitrophenyl)imidazo[1,2-a]pyridine (3e**). Yellow solid, mp 264-266 °C Lit.:<sup>46</sup> 265-267 °C; Yield: 0.0956 g (80%); NMR <sup>1</sup>H (DMSO-*d*<sub>6</sub>, 400 MHz): δ = 8.7 – 8.5 (m, 2H), 8.3 (d, *J* 8.9 Hz, 2H), 8.2 (d, *J* 8.9 Hz, 2H), 7.6 (d, *J* 9.1 Hz, 1H), 7.3 (t, *J* 7.9 Hz, 1H), 6.9 (t, *J* 6.7 Hz, 1H). NMR <sup>13</sup>C (DMSO-*d*<sub>6</sub>, 100 MHz): δ = 144.8, 143.6, 140.4, 138.8, 125.3, 124.5, 123.9, 122.2, 115.2, 111.0, 109.7. MS: *m/z* (rel. int.) 239 (100), 193 (57), 115 (2.3) 78 (24).

**2-**(*p*-Tolyl)imidazo[1,2-*a*]pyridine (3f). White solid, mp 145-146 °C; Lit.:<sup>45</sup> 144-145 °C. Yield: 0,088 g (85%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400MHz):  $\delta$  = 8.1 (dt, *J* 6.8, 1.2 Hz, 1H), 7.9 – 7.8 (m, 3H), 7.6 (dq, *J* 9.1, 1.0 Hz, 1H), 7.3 – 7.2 (m, 2H), 7.2 (ddd, *J* 9.1, 6.8, 1.3 Hz, 1H), 6.8 (td, *J* 6.8, 1.2 Hz, 1H), 2.4 (s, 3H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 75 MHz):  $\delta$  = 145.8, 145.5, 137.8, 130.8, 129.4, 125.9, 125.5, 124.5, 117.3, 112.3, 107.7, 21.3. MS: *m/z* (rel. int.) 208 (100), 193 (1.7), 91 (5.8), 78 (16).

**2-(4-Methoxyphenyl)imidazo[1,2-***a***]pyridine (3g)**. Yellow solid; mp. 132-134 °C; Lit.:<sup>45</sup> 135-136 °C; Yield: 0.109 g (90%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz): δ = 8.1 (d, *J* 6.8 Hz, 1H), 7.9 (d, *J* 8.9 Hz, 2H), 7.7 (d, *J* 2.3 Hz, 1H), 7.6 (d, *J* 

9.1 Hz, 1H), 7.2 – 7.1 (m, 1H), 7.0 – 6.9 (m, 2H), 6.7 (tq, *J* 6.6, 1.2 Hz, 1H), 3.8 (d, *J* 1.1 Hz, 3H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 159.5, 145.5, 145.5, 127.2, 126.3, 125.4, 124.4, 117.1, 114.1, 112.2, 107.2, 55.2. MS: *m/z* (rel. int.) 224 (100), 209 (58), 181 (65), 78 (56).

**2-(2-Methoxyphenyl)imidazo[1,2-a]pyridine** (**3h**). Brown solid; mp 94-96 °C; Lit.:<sup>47</sup> 95.5-97 °C. Yield: 0.109 g (98%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.3 (dd, *J* 7.7, 1.8 Hz, 1H), 8.1 (s, 1H), 8.0 (dt, *J* 6.8, 1.2 Hz, 1H), 7.5 (dd, *J* 9.1, 1.0 Hz, 1H), 7.2 – 7.2 (m, 1H), 7.1 – 7.0 (m, 2H), 6.9 (dd, *J* 8.3, 1.1 Hz, 1H), 6.6 (td, *J* 6.8, 1.2 Hz, 1H), 3.9 (s, 3H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 156.8, 144.3, 141.1, 128.9, 128.7, 125.6, 124.5, 122.3, 121.0, 117.2, 112.5, 112.0, 110.9, 55.4. MS: *m/z* (rel. int.) 224 (100), 193 (1.5), 209 (54.2), 78 (21.4).

**General procedure for the synthesis of 3-selanylimidazo[1,2-***a***]pyridines 5a-q.** A mixture of 2-aminopyridine **1** (0.5 mmol), 2-bromoacetophenone derivative **2** (0.5 mmol), K<sub>2</sub>CO<sub>3</sub> (0.103g, 0,75 mmol) and PEG-400 (1 mL), was added to a 10 mL vial. Then, the ultrasonic probe was introduced in the flask and the mixture was sonicated at 20 kHz and 60% of amplitude. The progress of the reaction was monitored by TLC as indicated in Table 2. Then diorganyldiselenide **4** (0.3 mmol) and CuSO<sub>4</sub>/KI (2 equiv.) were added and the reaction mixture was sonicated again at 20 kHz and 60% of amplitude by the time indicated in Table 4. The progress of the reaction was monitored by TLC. After that, the reaction mixture was received in water (10 mL), extracted with ethyl acetate (3 x 5 mL), dried over MgSO<sub>4</sub> and concentrated under vacuum. The residue was purified by column chromatography using silica gel and a mixture of ethyl acetate/hexanes as the eluent.

**2-Phenyl-3-(phenylselanyl)imidazo[1,2-***a***]pyridine (5a**). Yellow solid, mp 91.1-93.1 °C. Yield: 0.166 g (95%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ = δ 8.31 (d, *J* 6.8 Hz, 1H), 8.16 (d, *J* 7.2 Hz, 2H), 7.69 (d, *J* 9.0 Hz, 1H), 7.42 (t, *J* 7.5 Hz, 2H), 7.34 (t, *J* 7.3 Hz, 1H), 7.27 – 7.23 (m, 1H), 7.13 – 7.07 (m, 5H), 6.78 (t, *J* 6.7 Hz, 1H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): δ =151.7, 147.7, 133.8, 130.8, 129.6, 128.7, 128.3, 128.25, 128.18, 126.6, 126.3, 125.5, 117.5, 112.8, 102.9. MS: *m/z* (rel. int.) 350 (37.5), 270 (100.0), 193 (9.8), 77 (9.9).

**3-[(4-Chlorophenyl)selanyl]-2-phenylimidazo[1,2-***a***]<b>pyridine** (**5b**). White solid, mp 135.1-136.4; Yield: 0.173 g (90%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.28 (d, *J* 6.9 Hz, 1H), 8.15 – 8.13 (m, 2H), 7.69 (d, *J* 9.0 Hz, 1H), 7.42 (t, *J* 7.4 Hz, 2H), 7.35 (t, *J* 7.3 Hz, 1H), 7.29 – 7.24 (m, 1H), 7.10 (d, *J* 8.6 Hz, 2H), 6.99 (d, *J* 8.6 Hz, 2H), 6.81 (td, *J* 6.8, 0.8 Hz, 1H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 151.9, 147.7, 133.6, 132.8, 129.7, 129.5, 129.0, 128.6, 128.4, 128.2, 126.4, 125.3, 117.5, 113.0, 102.4. MS: *m/z* (rel. int.) 384 (46.5), 304 (100.0), 194 (23.6), 78 (98.6).

**3-[(4-Fluorophenyl)selanyl]-2-phenylimidazo[1,2-***a***]<b>pyridine** (**5c**). Yellow solid, mp 116.5-118.1°C Yield: 0.130 g (71%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.31 (d, *J* 6.8 Hz, 1H), 8.16 (d, *J* 7.4 Hz, 2H), 7.70 (d, *J* 9.0 Hz, 1H), 7.45 – 7.34 (m, 3H), 7.29 – 7.25 (m, 1H), 7.07 (dd, *J* 8.5, 5.3 Hz, 2H), 6.87 – 6.80 (m, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 162.0 (d, <sup>1</sup>*J*<sub>C-F</sub> = 246.4 Hz), 151.5, 147.5, 133.5, 130.2 (d, <sup>3</sup>*J*<sub>C-F</sub> = 7.6 Hz), 128.6, 128.4, 128.2, 126.4, 125.3, 125.0 (d, <sup>4</sup>*J*<sub>C-F</sub> = 3.3 Hz), 117.5, 116.7 (d, <sup>2</sup>*J*<sub>C-F</sub> = 21.9 Hz), 113.0, 103.1. MS: *m/z* (rel. int.) 368 (44.0), 288 (100.0), 194 (11.3), 78 (75.3).

**2-Phenyl-3-{[3-(trifluoromethyl)phenyl]selanyl}imidazo[1,2-***a***]<b>pyridine** (**5d**). Yellow solid, mp 93.1-95.7 °C Yield: 0.146 g (70%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.31 (d, *J* 6.9 Hz, 1H), 8.12 (d, *J* 7.2 Hz, 2H), 7.74 (d, *J* 9.0 Hz, 1H), 7.48 – 7.30 (m, 6H), 7.22 (t, *J* 7.8 Hz, 1H), 7.10 (d, *J* 7.9 Hz, 1H), 6.89 – 6.85 (m, 1H); <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 152.2, 147.8, 133.3, 132.3, 131.9 (q, <sup>2</sup>*J*<sub>C-F</sub> = 32.5 Hz), 131.1, 130.0, 128.7, 128.6, 128.3, 126.8, 125.3, 124.9 (q, <sup>3</sup>*J*<sub>C-F</sub> = 3.8 Hz), 123.5 (q, <sup>3</sup>*J*<sub>C-F</sub> = 3.6 Hz), 123.0 (q, <sup>1</sup>*J*<sub>C-F</sub> = 273.4 Hz), 117.6, 113.3, 101.8. MS: *m/z* (rel. int.) 418 (50.8), 338 (100.0), 193 (11.7), 78 (79.9).

**3-[(4-Methoxyphenyl)selanyl]-2-phenylimidazo[1,2-***a***]pyridine (5e). Yellow oil, yield: 0.138 g (73%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** *δ* **= 8.36 (d,** *J* **6.7 Hz, 1H), 8.20 (d,** *J* **8.1 Hz, 2H), 7.65 (d,** *J* **8.8 Hz, 1H), 7.42 (t,** *J* **7.5 Hz, 2H), 7.36 – 7.32 (m, 1H), 7.23 – 7.19 (m, 1H), 7.07 (d,** *J* **8.5 Hz, 2H), 6.77 (t,** *J* **6.8 Hz, 1H), 6.68 (d,** *J* **8.5 Hz, 2H), 3.65** 

(s, 3H); <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 159.1, 151.2, 147.4, 134.0, 130.8, 128.8, 128.2, 128.1, 126.0, 125.5, 120.5, 117.4, 115.4, 112.7, 104.1, 55.2. MS: *m/z* (rel. int.) 380 (21.3), 273 (61.4), 194 (100.0), 78 (70.5).

**2-Phenyl-3-**(*p*-tolylselanyl)imidazo[1,2-*a*]pyridine (5f). White solid, mp 126.5-127.8 °C. Yield: 0.098 g (54%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  =8.4 (d, *J* 6.9 Hz, 1H), 8.2 – 8.1 (m, 2H), 7.7 (dt, *J* 9.1, 1.1 Hz, 1H), 7.5 – 7.4 (m, 2H), 7.4 – 7.3 (m, 2H), 7.1 – 7.0 (m, 4H), 6.8 (td, *J* 6.8, 1.1 Hz, 1H), 2.3 (s, 3H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 151.5, 147.6, 136.7, 133.8, 130.4, 128.8, 128.5, 128.4, 128.3, 126.9, 126.4, 125.6, 117.5, 112.9, 29.7, 20.9. MS: *m/z* (rel. int.) 364 (33.9), 284 (100), 193 (5.3), 78 (65.3). HRMS: Calcd. for C<sub>20</sub>H<sub>16</sub>N<sub>2</sub>Se [M+H]<sup>+</sup>: 365.0552 found: 365.0551.

**3-(MesityIseIanyI)-2-phenyIimidazo[1,2-***a***]pyridine (5g)**. White solid, mp 49.1-51.6 °C. Yield: 0.074 g (38%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.1 – 8.0 (m, 3H), 7.7 (dt, *J* 9.0, 1.2 Hz, 1H), 7.5 – 7.4 (m, 2H), 7.4 – 7.4 (m, 1H), 7.2 – 7.2 (m, 1H), 6.8 – 6.7 (m, 3H), 2.2 (s, 6H), 2.2 (s, 3H); <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 150.0, 146.8, 141.3, 138.0, 134.1, 129.4, 129.0, 128.1, 128.1, 126.2, 125.4, 125.4, 117.3, 112.6, 104.6, 23.6, 20.7 . MS: *m/z* (rel. int.) 392 (28), 312 (100), 193 (8.2), 78 (39). HRMS: Calcd. for C<sub>22</sub>H<sub>20</sub>N<sub>2</sub>Se: [M+H]<sup>+</sup>: 393.0865 found: 393.0865.

**3-(Butylselanyl)-2-phenylimidazo[1,2-***a***]pyridine (5h)**. **B**rown oil. Yield: 0.139 g (81%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.52 (d, *J* 6.8 Hz, 1H), 8.23 (d, *J* 7.5 Hz, 2H), 7.66 (d, *J* 9.0 Hz, 1H), 7.45 (t, *J* 7.6 Hz, 2H), 7.35 (t, *J* 7.3 Hz, 1H), 7.26 – 7.22 (m, 1H), 6.87 (t, *J* 6.7 Hz, 1H), 2.64 (t, *J* 7.3 Hz, 2H), 1.45 (quint, *J* 7.2 Hz, 2H), 1.27 (sext, *J* 7.3 Hz, 2H), 0.74 (t, *J* 7.3 Hz, 3H).<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 150.1, 147.0, 134.0, 128.6, 128.0, 127.9, 125.7, 125.4, 117.3, 112.5, 104.2, 31.9, 29.1, 22.5, 13.3.MS: *m/z* (rel. int.) 330 (44.0), 288 (100.0), 194 (11.3), 78 (75.3).

**2-(4-Chlorophenyl)-3-(phenylselanyl)imidazo[1,2-***a***]<b>pyridine** (5i). Yellow solid, mp 81.1-83.4 °C; Yield: 0,103 g (54%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.4 (dt, *J* 6.9, 1.2 Hz, 1H), 8.2 – 8.1 (m, 2H), 7.7 (dt, *J* 9.0, 1.1 Hz, 1H), 7.4 – 7.4 (m, 2H), 7.4 – 7.3 (m, 1H), 7.2 – 7.1 (m, 3H), 7.1 – 7.1 (m, 2H), 6.9 (td, *J* 6.8, 1.2 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 150.6, 147.8, 134.5, 132.3, 130.6, 130.0, 129.8, 128.6, 128.2, 126.8, 126.7, 125.6, 117.5, 113.2, 103.0. MS: *m/z* (rel. int.) 384 (41.8), 304 (100.0), 193 (1.8), 78 (83.8).

**2-(4-Fluorophenyl)-3-(phenylselanyl)imidazo[1,2-***a***]<b>pyridine** (5**j**). Yellow solid, mp 99.5-101.3 °C Yield: 0.105 g (57%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.36 (dt, *J* 6.9, 1.1 Hz, 1H), 8.21 – 8.16 (m, 2H), 7.71 (dt, *J* 9.0, 1.1 Hz, 1H), 7.33 – 7.28 (m, 1H), 7.20 – 7.10 (m, 7H), 6.85 (td, *J* 6.8, 1.2 Hz, 1H).<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 163.0 (d, <sup>1</sup>*J*<sub>C-F</sub> = 248.1 Hz), 150.8, 147.7, 130.7, 130.5 (d, <sup>3</sup>*J*<sub>C-F</sub> = 8.1 Hz), 130.0 (d, <sup>4</sup>*J*<sub>C-F</sub> = 3.2 Hz), 129.7, 128.2, 126.7, 126.4, 125.5, 117.4, 115.2 (d, <sup>2</sup>*J*<sub>C-F</sub> = 21.3 Hz), 113.0, 102.7. MS: *m/z* (rel. int.) 368 (35.5), 288 (100.0), 211 (7.5), 78 (77.2).

**2-(4-Nitrophenyl)-3-(phenylselanyl)imidazo[1,2-***a***]pyridine (5l). Yellow solid, mp 154.4-156.4 °C. Yield: 0,0967 g (49%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz): \delta = 8.4 – 8.3 (m, 3H), 8.2 – 8.1 (m, 2H), 7.6 (dt,** *J* **9.0, 1.1 Hz, 1H), 7.3 – 7.2 (m, 1H), 7.1 – 7.1 (m, 3H), 7.1 – 7.0 (m, 2H), 6.8 (td,** *J* **6.8, 1.2 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz): \delta = 148.9, 147.9, 147.5, 140.2, 130.1, 129.9, 129.2, 128.3, 127.2, 127.1, 125.7, 123.5, 117.8, 113.6, 104.6. MS:** *m/z* **(rel. int.) 395 (16.5), 315 (69), 193 (2.8), 78 (100). HRMS: Calcd. For C<sub>19</sub>H<sub>14</sub>N<sub>3</sub>O<sub>2</sub>Se: [M+H]<sup>+</sup>: 396.0246 found: 396.0380.** 

**3-(Phenylselanyl)-2-(p-tolyl)imidazo[1,2-***a***]pyridine (5m)**. White solid, mp 65-66 °C; Yield: 0.131 g (72%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz): δ = 8.4 (dt, *J* 6.8, 1.2 Hz, 1H), 8.1 (d, *J* 8.2 Hz, 2H), 7.7 (d, *J* 9.0 Hz, 1H), 7.4 – 7.2 (m, 3H), 7.2 – 7.2 (m, 3H), 7.1 (dt, *J* 6.8, 2.4 Hz, 2H), 6.9 (td, *J* 6.8, 1.2 Hz, 1H), 2.4 (s, 3H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 100 MHz): δ = 151.8, 147.6, 138.3, 130.9, 130.8, 129.6, 129.0, 128.5, 128.1, 126.6, 126.3, 125.5, 117.3, 112.8, 102.4, 21.3 .MS: *m/z* (rel. int.) 364 (34.2), 284 (100), 193 (1.1), 78 (64.3).

**2-(4-Methoxyphenyl)-3-(phenylselanyl)imidazo[1,2-***a***]pyridine (5n). Yellow oil, yield: 0.133 g (70%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** *δ* **= 8.30 (d,** *J* **6.8 Hz, 1H), 8.13 (d,** *J* **8.8 Hz, 2H), 7.67 (d,** *J* **9.0 Hz, 1H), 7.26 – 7.22 (m, 1H),** 

7.16 – 7.07 (m, 5H), 6.95 (d, J 8.8 Hz, 2H), 6.79 – 6.75 (m, 1H), 3.80 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 159.8, 151.6, 147.6, 130.9, 129.9, 129.5, 128.1, 126.5, 126.3, 126.2, 125.4, 117.2, 113.7, 112.7, 101.9, 55.1.MS: *m/z* (rel. int.) 380 (32.9), 300 (100.0), 150 (4.3), 78 (45.6).

**2-Phenyl-3-(phenylthio)imidazo[1,2-***a***]pyridine (5p**). White solid, mp 106-108 °C; Yield: 0.081 g (54%); NMR <sup>1</sup>H (CDCl<sub>3</sub>, 400 MHz):  $\delta$  = 8.3 – 8.2 (m, 3H), 7.8 (dt, *J* 9.0, 1.1 Hz, 1H), 7.5 – 7.1 (m, 7H), 7.1 – 7.0 (m, 2H), 6.9 (td, *J* 6.8, 1.2 Hz, 1H). NMR <sup>13</sup>C (CDCl<sub>3</sub>, 75 MHz):  $\delta$  = 151.3, 147.0, 135.1, 133.2, 129.4, 128.5, 128.4, 128.3, 126.6, 126.0, 125.4, 124.4, 117.6, 113.0, 106.2 .MS: *m/z* (rel. int.) 302 (41.5), 225 (100.0), 193 (15.1), 77 (9.7).

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#### **Supplementary Material**

Copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra of compounds **3** and **5**.

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