

Singlet oxygen addition to cyclo-1,3-hexadienes from natural sources and from organocatalytic enal dimerization

Axel G. Griesbeck,* Alan de Kiff, Jörg M. Neudörfl, and Sarah Sillner

Department of Chemistry, Greinstr. 4, D-50939 Köln / Cologne, Germany

E-mail: griesbeck@uni-koeln.de

Dedicated to Michael Orfanopoulos on the occasion of his retirement
and his 67th birthday

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Abstract

The photooxygenation reactions of the natural monoterpene aldehyde safranal (**1**) and its reduction product safranol (**4**) were studied and the endoperoxide **5** from safranol was isolated and characterized by single crystal X-ray analysis. A dyade synthesis using DCC coupling of safranol with artesunic acid delivered the reactive substrate **7** for 1,3-diene photooxygenation. Singlet oxygen reaction of the substrate **7** enabled the synthesis of the trioxane-endoperoxide dyade **8**. The diastereoisomeric 1,3-cyclohexadienes **10** were obtained from the proline-catalyzed Diels-Alder dimerization of the Michael aldehyde **9**. From the singlet oxygen addition with the diastereoisomeric substrates **10**, only the kinetically preferred *cis*-endoperoxide **11** was formed.

Keywords: Singlet oxygen, Diels-Alder reaction, cyclohexadienes, organocatalysis

Introduction

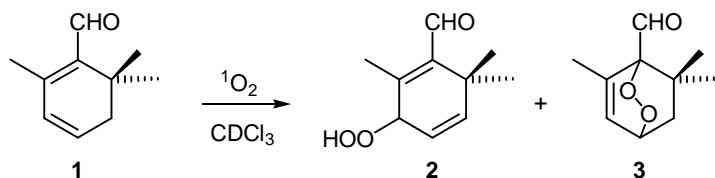
The singlet oxygen cycloaddition to conjugated cyclodienes is a highly controlled and reliable route to 1,4-oxyfunctionalized products.¹ The primary reaction event is the formation of bicyclic 1,2-dioxenes (*i.e.* endoperoxides) by a concerted but asynchronous 4+2 cycloaddition step. Ene reactions can sometimes compete with this cycloaddition route and for heterocyclic substrates such as thiophenes, pyrroles, oxazoles, and especially furans, the endoperoxide formation is exclusive.² There are also numerous naturally occurring 1,2-dioxenes, that are most probably not formed biosynthetically by singlet oxygen reactions, that exhibit interesting and diverse pharmacological properties.³ A well-known natural peroxide that can also be generated by singlet oxygen addition is ascaridole.⁴ This compound was used as an anthelmintic drug and was among the first targets for the Schenck investigations on photooxygenations using sunlight

irradiation of dye solutions which were very early examples of sustainable photocatalysis in synthesis.⁵

Our interest in this field of photochemistry arose from the search for new and effective antimalarial peroxides, an intensively investigated area in medicinal chemistry.⁶ The natural sesquiterpene artemisinin (qinhaosu), used for centuries in Chinese folk medicine as a plant extract, has initiated and stimulated this field of research.⁷ The enormous potency of the natural product that involves also anti-cancer drug activity just recently emerged.^{8,9} At the same time, however, reports appeared that describe the appearance of plasmodium species resistant against artemisinin and artemisinin-derivatives.^{10,11} Three approaches are currently under investigation to deal with this fatal trend: a) artemisinin combination therapy (ACT, with other non-peroxidic antimalarial drugs),¹² b) new peroxidic substances following the structural prototype,^{13,14} c) dyade concepts¹⁵ involving structure combinations of two artemisinin monomers,¹⁶ artemisinin and quinolines,¹⁷ or artemisinin derivatives with synthetic 1,2,4-trioxane structures.^{18,19} In this context, ascaridole was also described as one potential dyade partner molecule.²⁰ To explore new structural motifs, we envisaged other natural occurring terpenoids with pre-endoperoxide structures. One attractive example is the trimethylated cyclohexadiene safranal **1**. Safranal constitutes the major spice component of crocus flowers together with crocin, crocetin and picrocrocetin, and is crucial for the aroma of saffron.²¹ We envisaged the singlet oxygen reaction with the aldehyde **1** as a route to an electrophilic building block for dyade coupling.

Results and Discussion

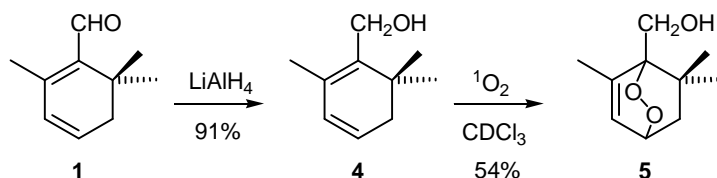
The photooxygenation of the aldehyde **1** in CDCl_3 delivered after long reaction a mixture of the ene product **2** and the desired endoperoxide **3** (Scheme 1) together with unreacted starting material. Column chromatography allowed the isolation of **2**²² but unfortunately led to decomposition of the endoperoxide **3**. Therefore, we reversed the dyade concept and allocated the nucleophilic part to the endoperoxide component, *i.e.* conversion of the aldehyde safranal to the corresponding alcohol.²³



Scheme 1. Photooxygenation of natural safranal (**1**).

Near quantitative reduction of safranal (**1**) was achieved with LiAlH_4 , and the subsequent photooxygenation of the alcohol **4** in CDCl_3 resulted in the endoperoxide **5** in a chemoselective singlet oxygen 4+2 cycloaddition (Scheme 2). No ene products were detected in the crude NMR

of the reaction mixture indicating that the hydroxymethyl group strongly activates the diene system towards 4+2 cycloaddition.



Scheme 2. Synthesis and photooxygenation of safranols (**4**).

The endoperoxide **5** was crystallized in racemic form from acetone and analyzed by crystal structure analysis (Figure 1).²⁴ In contrast to the substrate **4** that is described as unstable, the endoperoxide **5** is completely stable at room temperature.

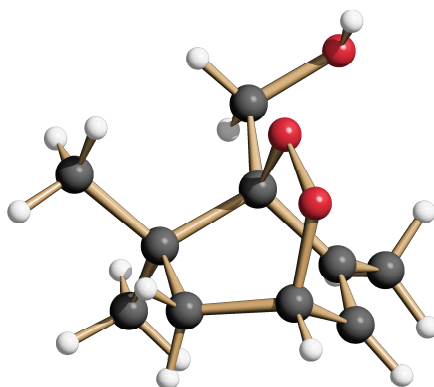
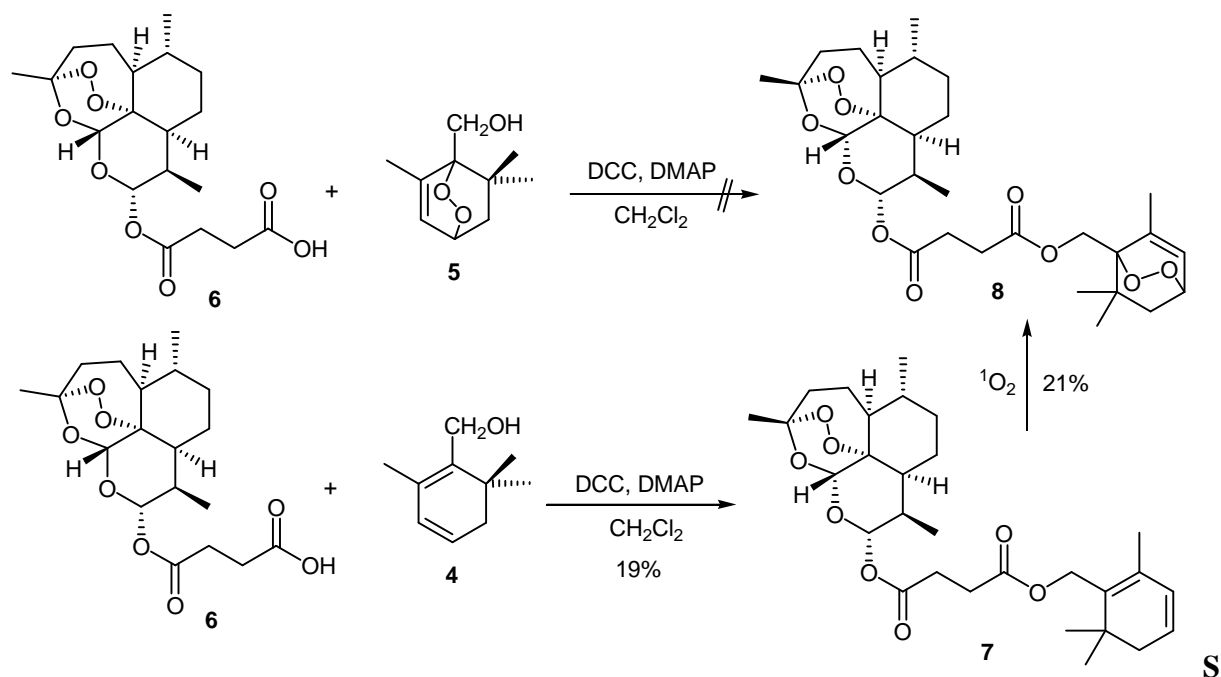


Figure 1. Structure of the endoperoxide **5** in the crystal.²³

To accomplish the dyade synthesis from endoperoxide **5** and an artemisinin skeleton, the Steglich esterification of **5** with artesunate (**6**) was investigated; a well-known process with numerous alcohols.²⁵ In contrast to the literature reports coupling was not successful and led to Brønsted-acid catalyzed decomposition of the alcohol component. Consequently, safranols was used as the alcohol component and esterified with artesunate using DCC/DMAP to give the ester **7** in 19% isolated yield. Photooxygenation under standard conditions led to the desired artesunate-endoperoxide couple in 21% yield (Scheme 3).



Scheme 3. Synthesis and photooxygenation of the artesunate-safranol dyade 7.

In the NMR spectra of the dyade **8** (Figure 2), the characteristic signals for the artemisinin skeleton – acetal H-11 and epimeric center H-12 - clearly show that **8** exists in an epimerically pure form with the ester substituent at C-12 in anti-position to the methyl group at C-13. The safranol endoperoxide part that is linked by the succinate chain appears as one set of signals in the ¹H as well as ¹³C NMR (significant for carbon: the methylene group C-20 and the stereogenic carbon center C-21). No single signal doubling was observed neither in the ¹³C nor in the ¹H-NMR spectra indicating that only one diastereoisomer of the dyade **8** was formed in the final photooxygenation process (**7**→**8**).

During this study, we discovered an alternative approach to 1,3-cyclohexadiene-1-carbaldehydes as substrates for dyade coupling. We performed these experiments in the context of organocatalytic photooxygenation reactions following the α -hydroxylation route developed by Cordova *et al.* from aliphatic ketones and aldehydes, respectively, with singlet oxygen and amino acids as chiral organocatalysts.^{26,27} When these conditions were applied to α,β -unsaturated aldehydes, complex mixtures of peroxides were observed that could not be separated. We expected the formation of dieneamines, as described in the work by Jørgensen *et al.*^{28,29} Obviously, we were not able to trap these dienes with singlet oxygen. Thus, the reaction conditions were modified and the photooxygenation was designed as a delayed step. When the Michael aldehyde **9** was treated with proline in the absence of any (additional) dienophile, the 1,3-cyclohexadiene carbaldehyde **10** was isolated as a mixture of *trans/cis*-isomers (3:2, enantioselectivity was not determined) in good yields. The rationale for this reaction is shown in Scheme 4 following the sequence that was described for intramolecular cycloaddition by Christmann and coworkers.³⁰ During the subsequent reaction with singlet oxygen, one new set of

signals appeared in the NMR spectra with concurrent disappearance of the signals of the *cis*-isomer. After completion of the reaction, a complete conversion of *cis*-**10** was observed with less than 5% of the *trans*-endoperoxide **11** observable. The endoperoxide **11** with *cis* configuration, that could not be isolated from the reaction mixture, appeared in the ^{13}C NMR with a new indicative set of signals at 73.2 (C-4), 98.6 (C-1), 127.1 (C-5), and 135.1 ppm (C-6). We are currently investigating this new synthetic approach to 1,3-cyclohexadienes³¹ together with the unusual high stereoselective singlet oxygen cycloaddition.

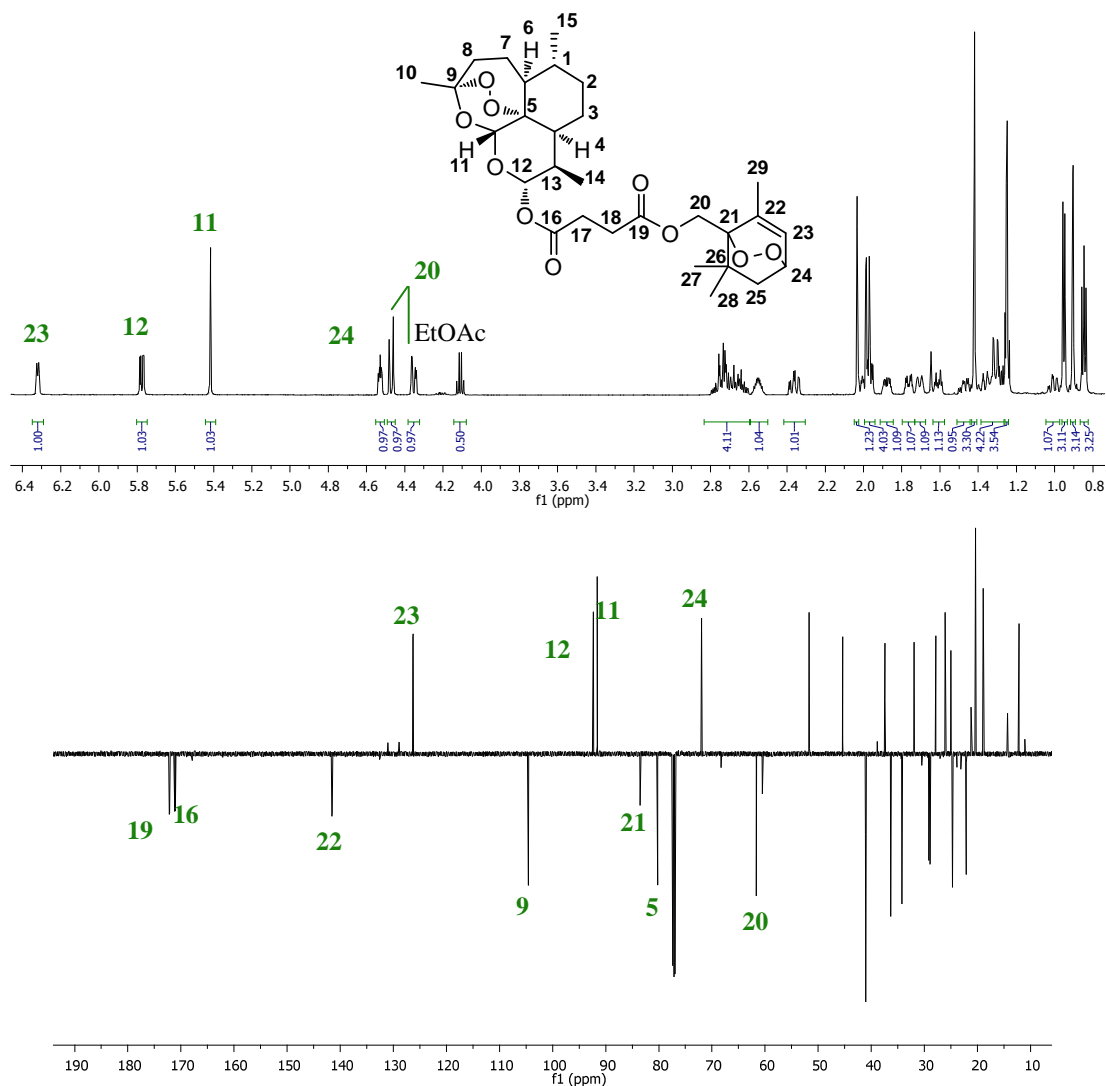
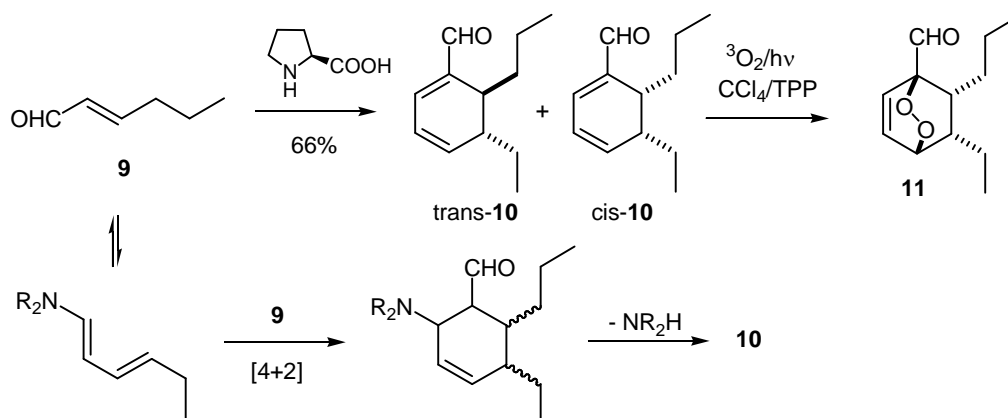


Figure 2. ^1H - and ^{13}C -NMR of the trioxane-dioxane-dyade **8**.



Scheme 4. Organocatalytic cycloaddition/elimination synthesis and diastereoselective photooxygenation of cyclohexadienes **10**.

Experimental Section

General. *Meso*-tetraphenylporphyrin (TPP) was purchased from Porphyrin Systems, Bremen. The solvents for solution photooxygenation were puriss. and used as purchased. NMR spectra were recorded on Bruker DPX 300 and DPX 600 spectrometers, chemical shifts are given in δ (ppm) versus 0.0 (TMS for ^1H) and 77.0 (CDCl_3 for ^{13}C), multiplicities were determined by DEPT; IR spectra were obtained from a Perkin-Elmer 1600 series FTIR spectrometer; melting points were determined with a Büchi melting point apparatus (type Nr. 535) and are uncorrected; CHN-combustion analyses were measured using an Elementar Vario EL instrument.

1-Hydroxymethyl-2,6,6-trimethylcyclohexa-1,3-diene (safranols, 4). To a suspension of 320 mg (8.32 mmol, 2.5 equiv.) of lithium aluminium hydride in dry diethyl ether (30 mL) externally cooled $0\text{ }^\circ\text{C}$ was slowly added a solution of safranols **1** (500 mg, 3.33 mmol) in diethyl ether (20 mL). After stirring for 1 h at $0\text{ }^\circ\text{C}$ and 1 h at room temperature, the reaction was quenched carefully with water and filtrated, extracted with diethyl ether ($3 \times 20\text{ mL}$) and dried (MgSO_4). After filtration and evaporation, safranols (**4**) (460 mg, 91%) was obtained as colorless oil that was directly used for further reactions. ^1H NMR (300 MHz, CDCl_3): 1.05 (s, 6H, $\text{C}(\text{CH}_3)_2$), 1.81 (s, 3H, CH_3), 2.06 (d, 2H, J 2.6 Hz, 5-H), 4.21 (s, 2H, CH_2OH), 5.69-5.79 (m, 2H, 3-H, 4-H); ^{13}C NMR (75 MHz, CDCl_3): 18.0 (q, CH_3), 26.7 (q, $\text{C}(\text{CH}_3)_2$), 33.4 (s, C-6), 40.1 (t, C-5), 58.7 (t, CH_2OH), 126.3 (d, $\text{HC}=\text{CH}$), 129.1 (d, $\text{HC}=\text{CH}$), 129.3 (s, $\text{C}=\text{C}$), 137.4 (s, $\text{C}=\text{C}$).

1-Hydroxymethyl-6,7,7-trimethyl-2,3-dioxabicyclo[2.2.2]oct-5-ene (5). A solution of safranols **4** (228 mg, 1.5 mmol) in ($2 \times 10^{-4}\text{ M}$ in TPP) deuteriochloroform (20 mL) was irradiated at room temperature by means of a halogen metal vapour lamp under purging with dry oxygen and under TLC and NMR control for 8 h. After evaporation of the solvent and column chromatography (silica, EtOAc/cyclohexane 1:4), the title compound **5** (150 mg, 54%) was obtained as a colorless

solid, mp 101-102 °C. ^1H NMR (300 MHz, CDCl_3): 0.76 (s, 3H, $-(\text{CH}_3)_2$), 1.09 (s, 3H, $-(\text{CH}_3)_2$), 1.19 (dd, 1H, J 13.1, 2.2 Hz, 8-H), 1.81 (dd, 1H, J 13.1, 3.5 Hz, 8'-H), 1.99 (d, 3H, J 1.6 Hz, CH_3), 3.78-3.89 (m, 2H, CH_2OH), 4.38-4.43 (m, 1H, 4-H), 6.18 (dd, 1H, J 5.9, 1.0 Hz, 5-H); ^{13}C NMR (75 MHz, CDCl_3): 18.6 (q, C-CH_3), 24.7 (q, $-(\text{CH}_3)_2$), 27.3 (q, $-(\text{CH}_3)_2$), 33.6 (s, C-7), 41.0 (t, C-8), 59.2 (t, CH_2OH), 71.9 (d, C-4), 85.3 (s, C-1), 125.7 (d, C-5), 142.3 (s, C-6); HRMS (ESI, $\text{M}+\text{Na}$) Found: 207.0986, $\text{C}_{10}\text{H}_{16}\text{O}_3$ requires 207.0992.

Dihydroartemisinyl(2,6,6-trimethylcyclohexa-1,3-dien-1-yl)methylsuccinate (7). A solution of artesunate (7.69 g, 20.0 mmol) and DCC (4.33 g 21.0 mmol) in methylene chloride (100 mL) was cooled to 0 °C. Over a period of 30 min, a solution of safranol (**4**) (3.04 g, 20 mmol) and DMAP (320 mg, 2.6 mmol) were added. The mixture was stirred for 12 h at room temp., then treated with a small amount of water and the separated organic was washed several times with water, separated and dried (MgSO_4). After evaporation of the solvent and column chromatography (silica, EtOAc/cyclohexane 1:8), the title compound **7** (1.94 g, 19%) was obtained as a colorless oil. ^1H NMR (600 MHz, CDCl_3): 0.82 (d, 3H, J 7.2 Hz, 14-H), 0.94 (d, 3H, J 6.2 Hz, 15-H), 0.97-1.03 (m, 1H, 2-H), 1.00 (s, 6H, 27-H, 28-H), 1.80 (s, 3H, 29-H), 1.23-1.49 (m, 4H, 6-H, 1-H, 3-H, 7-H), 1.41 (s, 3H, 10-H), 1.58-1.62 (m, 1H, 4-H), 1.67-1.72 (m, 1H, 2-H), 1.73-1.78 (m, 1H, 3-H), 1.84-1.90 (m, 1H, 7-H), 1.98-2.04 (m, 1H, 8-H), 2.07 (d, 2H, 25-H), 2.35 (dt, 1H, J 14.4, 3.9 Hz, 8-H), 2.51-2.57 (m, 1H, 13-H), 2.61-2.75 (m, 4H, 17-H, 18-H), 4.72 (s, 2H, 20-H), 5.42 (s, 1H, 11-H), 5.77 (d, 1H, J 9.9 Hz, 12-H), 5.75-5.81 (m, 3H, 12-H, 23-H, 24-H); ^{13}C NMR (150 MHz, CDCl_3): 12.1 (q, C-14), 18.2 (q, C-29), 20.3 (q, C-15), 22.1 (t, C-3), 24.7 (t, C-7), 26.0 (q, C-10), 26.3 (q, Me), 28.8 (t, $-\text{CH}_2\text{COOR}$), 29.0 (t, $-\text{CH}_2\text{COOR}$), 29.1 (q, Me), 31.9 (d, C-13), 33.1 (s, C-26), 34.2 (t, C-2), 36.3 (t, C-8), 37.3 (d, C-1), 39.8 (t, C-25), 45.3 (d, C-4), 51.6 (d, C-6), 61.5 (t, C-20), 80.2 (s, C-5), 91.6 (d, C-11), 92.4 (d, C-12), 104.6 (s, C-9), 127.0 (d, $\text{HC}=\text{CH}$), 128.9 (d, $\text{HC}=\text{CH}$), 131.8 (s, C_q), 131.9 (s, C_q), 171.1 (s, C-16), 172.2 (s, C-19), 171.3 (s, $-\text{COOR}$), 178.1 (s, $-\text{COOR}$).

Dihydroartemisinyl[(6,7,7-trimethyl-2,3-dioxabicyclo[2.2.2]oct-5-en-1-yl)methyl]-succinate (8). A solution of the dihydroartemisine **7** (780 mg, 1.5 mmol) in (2×10^{-4} M in TPP) of deuteriochloroform (20 mL) and pyridine (10 μL) was irradiated at room temperature by means of a halogen metal vapour lamp under purging with dry oxygen and under TLC and NMR control for 12 h. Evaporation of the solvent and column chromatography (silica, EtOAc/cyclohexane 1:6) afforded the title compound **8** (170 mg, 21%) as a colorless solid, mp 167-168 (decomp.); ^1H NMR (600 MHz, CDCl_3): 0.82 (d, 3H, J 7.1 Hz, 14-H), 0.90 (d, 3H, J 1.9 Hz, 27-H), 0.94 (d, 3H, J 6.2 Hz, 15-H), 0.97-1.03 (m, 1H, 2-H), 1.23-1.49 (m, 4H, 6-H, 1-H, 3-H, 7-H), 1.24-1.25 (m, 3H, 28-H), 1.27-1.39 (m, 4H, 6-H, 25-H, 1-H, 3-H), 1.41 (s, 3H, 10-H), 1.58-1.62 (m, 1H, 4-H), 1.67-1.72 (m, 1H, 2-H), 1.73-1.78 (m, 1H, 3-H), 1.84-1.90 (m, 1H, 7-H), 1.95-2.00 (m, 4H, 25-H, 29-H), 1.98-2.04 (m, 1H, 8-H), 2.35 (dt, 1H, J 14.4, 3.9 Hz, 8-H), 2.51-2.57 (m, 1H, 13-H), 2.61-2.75 (m, 4H, 17-H, 18-H), 4.35 (dd, 1H, J 12.6, 2.4 Hz, 20-H), 4.52-4.54 (m, 1H, 24-H), 5.42 (s, 1H, 11-H), 5.77 (d, 1H, J 9.9 Hz, 12-H), 6.32 (d, 1H, J 6.1 Hz, 23-H); ^{13}C NMR (150 MHz, CDCl_3): 12.1 (q, C-14), 18.9 (q, C-29), 20.3 (q, C-15), 22.1 (t, C-3), 24.7 (t, C-7), 25.0 (q, C-28), 26.0 (q, C-10), 27.9 (q, C-27), 28.8 (t, $-\text{CH}_2\text{COOR}$), 29.0

(t, $-\underline{\text{C}}\text{H}_2\text{COOR}$), 31.9 (d, C-13), 34.2 (t, C-2), 34.3 (s, C-26), 36.3 (t, C-8), 37.3 (d, C-1), 41.1 (t, C-25), 45.3 (d, C-4), 51.6 (d, C-6), 61.1 (t, C-20), 72.0 (d, C-24), 80.2 (s, C-5), 83.6 (s, C-22), 91.6 (d, C-11), 92.4 (d, C-12), 104.6 (s, C-9), 126.3 (d, C-23), 140.6 (s, C-21), 171.1 (s, $-\underline{\text{C}}\text{OOR}$), 171.2 (s, C-16), 172.2 (s, C-19), 178.1 (s, $-\underline{\text{C}}\text{OOR}$); HRMS (ESI) found: m/z 573.26688, $\text{C}_{29}\text{H}_{40}\text{O}_{10}$ requires 573.26701.

5-Ethyl-6-propylcyclohexa-1,3-dienecarbaldehyde (10). A solution of *trans*-2-hexenal (**9**) (1.0 g, 10 mmol) in methanol (30 mL) of was treated with (*S*)-proline (115 mg, 1.00 mmol) and stirred at room temp. for 24 h. The solvent was subsequently evaporated and the residue extracted with EtOAc (20 mL), washed with water (20 mL) and extracted again with EtOAc (2 \times 20 mL). Column chromatography (cyclohexane/EtOAc 3:1, R_f 0.48) gave a mixture of diastereoisomeric cyclohexadienes **10** (400 mg, 45%, *trans:cis* 65:35) as a colorless oil. ^1H NMR (300 MHz, CDCl_3 , *trans*-**10**): 0.81-0.96 (m, 6H, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3/\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 1.16-1.42 (m, 6H, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3/\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 2.18 (q, 1H, J 6.6 Hz $\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 2.69 (t, 1H, J 6.6 Hz $\underline{\text{C}}\text{H}-\underline{\text{C}}-\underline{\text{C}}=\text{O}$), 6.11 (dd, 1H, J 9.6, 5.2 Hz, $\text{CH}-\text{C}\text{H}_{\text{sp}2}-\underline{\text{C}}\text{H}_{\text{sp}2}$), 6.23 (dd, 1H, J 9.3, 5.9 Hz, $\text{CH}-\underline{\text{C}}\text{H}_{\text{sp}2}-\underline{\text{C}}\text{H}_{\text{sp}2}$), 6.66 (d, 1H, J 5.2 Hz, $\underline{\text{C}}\text{H}_{\text{sp}2}-\underline{\text{C}}-\underline{\text{C}}=\text{O}$), 9.46 (s, 1H, CHO); ^{13}C NMR (75 MHz, CDCl_3 , *trans*-**10**): 11.2 (q, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 14.2 (q, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 19.9 (t, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 26.4 (t, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 33.2 (d, $\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 34.7 (t, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 39.7 (d, 1C, $\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 122.0 (d, 1C, $\text{C}\text{H}_{\text{sp}3}-\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}$), 141.0 (d, 1C, $\text{C}\text{H}_{\text{sp}3}-\underline{\text{C}}\text{H}$), 141.2 (s, 1C, $\underline{\text{C}}-\text{CHO}$), 141.3 (d, 1C, $\text{CH}-\underline{\text{C}}-\text{CHO}$), 193.5 (d, 1C, CHO); ^1H NMR (300 MHz, CDCl_3 , *cis*-**10**): 0.81-0.96 (m, 6H, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3/\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 1.16-1.42 (m, 6H, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3/\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 2.33-2.46 (m, 1H, $\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 2.74-2.84 (m, $\underline{\text{C}}\text{H}-\underline{\text{C}}-\text{CHO}$), 6.00 (d, 1H, J 9.5 Hz, $\text{CH}-\text{C}\text{H}_{\text{sp}2}-\underline{\text{C}}\text{H}_{\text{sp}2}$), 6.10-6.20 (m, 1H, J 9.3, 5.9 Hz, $\text{CH}-\underline{\text{C}}\text{H}_{\text{sp}2}-\underline{\text{C}}\text{H}_{\text{sp}2}$), 6.70 (d, 1H, J 5.1 Hz, $\underline{\text{C}}\text{H}_{\text{sp}2}-\underline{\text{C}}-\underline{\text{C}}=\text{O}$), 9.52 (s, 1H, CHO); ^{13}C NMR (75 MHz, CDCl_3 , *cis*-**10**): 12.0 (q, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 14.8 (q, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 20.7 (t, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 23.3 (t, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 31.1 (d, $\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 28.0 (t, 1C, $\text{CH}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 41.0 (d, 1C, $\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_2-\underline{\text{C}}\text{H}_3$), 123.8 (d, 1C, $\text{C}\text{H}_{\text{sp}3}-\underline{\text{C}}\text{H}-\underline{\text{C}}\text{H}$), 142.1 (d, 1C, $\text{C}\text{H}_{\text{sp}3}-\underline{\text{C}}\text{H}$), 141.1 (d, 1C, $\underline{\text{C}}-\text{CHO}$), 143.7 (s, 1C, $\underline{\text{C}}-\text{CHO}$), 193.2 (d, 1C, CHO). GC-MS: τ_R (*trans*-**10**) = 9.82 min; m/z (%) 178 (35; $[\text{M}]^+$), 149 (13), 145 (10), 135 (12), 119 (10), 107 (40), 91 (31), 79 (100), 65 (9), 51 (7); τ_R (*cis*-**10**) = 10.16 min; m/z (%) 178 (21; $[\text{M}]^+$), 149 (9), 141 (12), 133 (13), 119 (11), 107 (30), 94 (42), 79 (100), 65 (25), 55 (19).

Acknowledgements

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