Enantioselective synthesis of the ester side chain of homoharringtonine

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Abstract
From D-Malic acid as chiral starting material, an efficient synthesis of the ester side chain of homoharringtonine has been developed. A cross-metathesis reaction leads to the formation of the key intermediate, which can be converted later by selective hydrogenation to the methyl ester side chain of homoharringtonine and deoxy-homoharringtonine in a total of six steps with 24.5% and 23.5% in yields, respectively.

Keywords: Homoharringtonine, cross-metathesis, cephalotaxine, self-retention of chirality

Introduction
Homoharringtonine (HHT) has been proved to be the most potent member of the cephalotaxus esters in treating leukemia.1-3 Research on the biological properties of this alkaloid has shown that HHT inhibits protein biosynthesis in the cell via the breakdown of polyribosomes, the release of completed globin chain, and the inhibition of the initiation of protein synthesis without effecting chain elongation.4-5 In 2012, omacetaxine mepesuccinate (Synribo), a semisynthetic HHT compound (99.7% purity) was used in studies of chronic myeloid leukemia (CML) and approved by the FDA for the treatment of CML. Later approval from the FDA in 2013 for this drug to be self-administered by patients further demonstrated its efficacy and safety.6

Figure 1. Cephalotaxine and Homoharringtonine.
Limited availability of HHT together with the higher purity of semisynthetic HHT over natural sources have engendered much interest in the semisynthesis of HHT from the more abundant but biologically inactive cephalotaxine. A number of synthetic approaches to the ester side chain of HHT have been described. In 2001, Jean d’Angelo and co-workers reported a ten-step stereoselective approach to the methyl ester derivative of the HHT side chain via a regio- and stereoselective Michael addition on a chiral amine. Starting from a chiral citric acid derivative, Russell described an approach to HHT ester side chain acid via Rosenmund reduction of homocitrate to form the corresponding aldehyde. This aldehyde was then oxidized to form Robin’s acid, which could be attached to the cephalotaxine moiety via Robin’s first semisynthesis of HHT. More recently, Yang reported the [2,3]-Meisenheimer rearrangement in the construction of chiral tertiary alcohols in the ester side chain. However, there is still a need for a better and more convenient synthesis of the side chain in order to perform an efficient transformation from cephalotaxine to homoharringtonine.

Herein we reported a synthetic approach to the ester side chain that is enantioselective, convenient, and distinct from the previous methods. The strategy is based on the Seebach’s procedure of alkylation of D-Malic acid with “self-retention of chirality” and the cross metathesis reaction of the resulted allyl acid with a tertiary alkenyl alcohol. The success of this synthetic approach enables an efficient pathway to HHT side chain synthesis as well as to that of other Cephalotaxus side chain ester derivatives.

Results and Discussion

Our methodology started with the application of Seebach’s concept of “self-reproduction of chirality” as a means to introduce the stereocenter of the ester side chain. Starting with d-Malic acid as a readily available chiral starting material, dioxolanone 2 was synthesized as previously reported. The cis-product was predominant due to its being favored thermodynamically.

Scheme 1. Synthesis of the key intermediate 5.
The allylation of the dioxolanone 2 with allyl bromide in the presence of two equivalents of LiHMDS in THF at -78 °C gave the desired compound in 75% yield. Due to the strong steric hindrance by the t-butyl group, the allyl electrophile attacks from the opposite face, thereby securing the required R-configuration.

The cross metathesis of the acid 3 with 2-methylbut-3-en-2-ol was next undertaken using Grubbs’s 2 catalyst in DCM with a large excess of the tertiary alcohol. Unfortunately, only starting materials together with the dimerization product of the alcohol were observed. However, the transformation of acid 3 into its methyl ester 4 by DCC/DMAP in the presence of catalytic amount of PTSA, permitted the cross metathesis to succeed in high yield. $^1$H NMR analysis showed that the product 5 exists mainly as the trans-isomer.

With the key intermediate 5 in hand, hydrogenation of the double bond with H$_2$-Pd/C in methanol gave surprisingly a quantitative dehydroxy product 7. This result can be explained by the fact that the allylic alcohol position in 5 is slightly acidic, thus making it more favourable to be removed under the reaction conditions. Addition of an amine, to neutralize the alcohol and afford a selective hydrogenation, was then applied. After several trials, we found that one equivalent of triethylamine in ethanol permitted a complete conversion of 5 into the desired product 6. The final transformations of the protected dioxalanes 6 and 7 were performed smoothly using MeOH/MeONa affording the methyl ester side chains of homoharringtonine 8 and deoxy-homoharringtonine 9 in very high yields. The NMR and optical rotation data obtained were identical with the reported data. 18

**Scheme 2.** Synthesis of the methyl ester side chains.
Conclusions

An enantioselective synthesis of homoharringtonine ester side chain and its deoxy-derivative have been completed in six steps with 24.5% and 23.5% overall yields, respectively. The key tactical elements of this synthesis include the use of chiral malic acid as starting material and the cross metathesis with available methylbut-3-en-2-ol. These enabled an efficient access to the protected intermediate 5, which can be converted easily into methyl ester of the side chain. This strategy can be considered as a potential pathway to the semi-synthesis of homoharringtonine and its derivatives by the coupling of cephalotaxine and the ester side chain. Studies directed towards the esterification of the α-hydroxy acid with cephalotaxine to produce enantiopure HHT are currently under investigation.

Experimental Section

General. All the reactions were carried out under a nitrogen atmosphere. Unless otherwise noted, all the reagents obtained from commercial sources were used without further purification. All solvents were dried by standard methods. THF were dried with sodium and benzophenone and used immediately after distillation. DCM were dried with diphosphorus pentoxide (P2O5). Pentane was distilled and then dried with sodium. Analytical thin-layer chromatography (TLC) was performed on 0.25 mm Merck precoated silica gel plates (60-F254). Column chromatography was carried out with silica gel (60-F254). The TLC plates were visualized with a UV lamp (254 nm and 366 nm) and/or with TLC visualizing solutions activated with heat, including: p-anisaldehyde solution and potassium permanganate solution. Mass spectral analyses were performed with a VARIAN 920-MS at VAST. The specific optical rotation data were measured with a JASCO P-2000 Polarimeter instrument (wavelength of the light used was 589 nm). 1H NMR and 13C NMR were recorded on BRUKER 300 and 500 MHz instruments using TMS as the internal standard and CDCl3 as the solvent.

2-((2R, 4R)-2-tert-Butyl-5-oxo-1,3-dioxolan-4-yl)acetic acid (2). To a suspension of D-(+)-malic acid (10 g, 74.58 mmol) and pivalaldehyde (13.2 mL, 116.1 mmol) in pentane (150 mL), PTSA (1.1 g, 6.39 mmol) and concentrated H2SO4 (2 drops) were added. The mixture was heated under reflux for 40 h with azeotropic removal of water. The resulting suspension was filtered. The solid cake was dissolved in CH2Cl2, and washed with 8% aqueous H3PO4 (2×40 mL). The combined organic phases were dried with Na2SO4. The solvent was removed under vacuum, giving 9.554 g of the product as a white solid (yield 64%). [α]D24 +2.20 (c 1.00, MeOH). 1H NMR (300 MHz, CDCl3): δH 5.33 (m, 1H), 4.77 – 4.54 (m, 1H), 3.08 – 2.77 (m, 2H), 0.98 (s, 9H). 13C NMR (75 MHz, CDCl3): δc 1745.0, 172.4, 111.4, 71.4, 35.9, 35.4, 23.4.

2-((2R,4R)-2-tert-butyl-4-allyl-5-oxo-1,3-dioxolan-4-yl)acetic acid (3). To a stirred solution of dioxolanone 2 (1.78 g, 8.83 mmol) in THF (110 mL) a solution of LiHMDS (0.5 M in THF,
17.65 mL, 8.83 mmol, 1 eq.) was slowly added at –78 °C through a dropping funnel. After 15 min, another portion of 17.65 mL LiHMDS 0.5M in THF (1 eq.) was added slowly to the mixture and it was stirred for 20 min. After the dropwise addition of the allyl bromide (1.53 mL, 17.65 mmol, 2 eq) over a period of 20 min the temperature was raised up to –10 °C over a period of 5.5 h. The resulting solution was partitioned between EtOAc and 1 M HCl, and extracted with EtOAc. The combined organic phases were dried with Na₂SO₄. The solvent was removed under vacuum and the residue purified by column chromatography (hexane/EtOAc, 9:1 – 1:1) to give 1.67 g pure product as yellow oil, yield 75%. 

\[ \alpha_D^{24} -18 \ (c \ 1.1, \text{MeOH}) \]

1H NMR (300 MHz, CDCl₃): δ H 5.90 – 5.73 (m, 1H), 5.32 – 5.18 (m, 3H), 2.85 (d, J=1.8 Hz, 2H), 2.57 (m, J=7.6, 5.1, 1.1 Hz, 2H), 0.95 (s, 9H). 13C NMR (75 MHz, CDCl₃): δ c 173.5, 173.4, 130.0, 121.3, 108.5, 79.8, 39.4, 38.16, 34.4, 23.6.

**Methyl 2-((2R,4R)-2-tert-butyl-4-allyl-5-oxo-1,3-dioxolan-4-yl)acetate (4).** To a solution of acid 3 (158 mg, 0.654 mmol) in 3.5 mL of MeOH (86.5 mmol, 132 eq) was added DMAP (6 mg, 0.0491 mmol, 7.5 mol%) and p-TsOH (5.7 mg, 0.03 mmol, 4.6 mol%). The mixture was cooled to 0 °C, then, DCC (170 mg, 0.824 mmol, 1.26 eq.) was added. After 18 h, the reaction was diluted with 20 mL MeOH. The solution was filtered through Celite and the solvent was removed in vacuum. The product was purified by column chromatography (hexane: EtOAc, 9:1 – 6:1) to give 137 mg ester as a colorless oil, yield 84%. 

\[ \alpha_D^{24} -14 \ (c \ 0.5, \text{MeOH}) \]

1H NMR (500MHz, CDCl₃): δ H 5.82 (m, J=7.2, 9.7, 17.6 Hz, 1H), 5.26-5.20 (m, 3H), 3.66 (s, 3H), 2.82 (dd, J=1.3, 12.1 Hz, 2H), 2.62-2.52 (m, 2H), 0.94 (s, 9H). 13C NMR (125 MHz, CDCl₃): δ c 173.5, 168.7, 130.1, 121.1, 108.7, 79.9, 51.9, 39.7, 38.2, 34.2, 23.5. HRMS m/z calculated for [M+Na]+ C₁₁H₁₆NaO₅ 279.1208 found 279.1201.

**Cross-metathesis product of ester with 2-methyl-but-3-en-2-ol (5).** Ester 4 (27 mg, 0.105 mmol) and Grubbs II catalyst (8 mg, 0.005 mmol) were dissolved in 1 mL of 2-methyl-but-3-en-2-ol (90 eq.). The mixture was refluxed at 45 °C for 18 h. The solvent was then removed and the product was purified by column chromatography (hexane: EtOAc 4:1 – 2:1) to give 30 mg of the product 5 (26 mg, 80% yield). 

\[ \alpha_D^{24} -26.4 \ (c \ 0.5, \text{MeOH}) \]

1H NMR (500 MHz, CDCl₃): δ H 5.78 (d, J=15.5 Hz, 1H), 5.60 (m, J=15.5, 7.4Hz, 1H), 5.18 (s, 1H), 3.65 (s, 3H), 2.79 (dd, J=15.5, 9.5Hz, 2H), 2.52 (m, 2H), 2.02 (m, 2H), 1.30 (s, 6H), 0.94 (s, 9H). 13C NMR (125 MHz, CDCl₃): δ c 173.5, 168.7, 146.0, 144.5, 118.2, 110.8, 108.3, 80.1, 70.5, 51.9, 39.7, 36.7, 34.2, 29.6, 29.6, 29.3, 23.5. HRMS m/z calculated for [M+Na]⁺ C₁₆H₂₆NaO₆ 337.1627 found 337.1620.

**Methyl 2-((2R,4R)-2-tert-butyl-4-(4-hydroxy-4-methylpentyl)-5-oxo-1,3-dioxolan-4-yl)acetate (6).** 46 mg of 5 (0.147 mmol) was dissolved in 3 mL of EtOH. 7.5 mg of Pd/C (17% weight) and 25 μL of TEA (1 eq) were added. The solution was then bubbled with H₂ in 15 min and keeps stirring in H₂ atmosphere for 3 hours. The solvent was then evaporated and the product was purified by column chromatography (hexane: EtOAC 4:1) to give 48 mg of 6 (yield 100%). 

\[ \alpha_D^{24} -26.4 \ (c \ 0.5, \text{MeOH}) \]

1H NMR (500 MHz, CDCl₃): δ H 5.15 (s, 1H), 3.65 (s, 3H), 2.81 (s, 2H), 1.44 – 1.80 (m, 7H), 1.44 (s, 6H), 0.93 (s, 9H). 13C NMR (125 MHz, CDCl₃): δ c 174.5, 169.3, 108.9, 80.8, 71.2, 52.6, 44.1, 40.3, 34.9, 34.9, 29.9, 29.9, 24.2, 18.8. HRMS m/z calculated for [M+Na]⁺ C₁₆H₂₈NaO₆ 339.1784 found 339.1776.
Methyl 2-((2R,4R)-2-tert-butyl-4-(4-methylpentyl)-5-oxo-1,3-dioxolan-4-yl)acetate (7). 50 mg of 5 (0.159 mmol) was dissolved in 5 mL of MeOH. 7 mg of Pd/C (14% weight) was added. The solution was then bubbled with H2 in 15 min and keeps stirring in H2 atmosphere for 3 hours. The solvent was then evaporated and the product was purified by column chromatography (Hexane: EtOAC 5:1) to give 44 mg of 7 (88%). \([\alpha]_D^{24} -20.1 \ (c \ 0.4, \text{MeOH})\) \(R_f \ 0.65\) (Hexane/ EtOAC, 4:1). 

1H NMR (500 MHz, CDCl3): \(\delta_H \ 5.16 \ (s, \ 1H), 3.67 \ (s, \ 3H), 2.83 \ (s, \ 2H), 1.78 \ (m, \ 1H), 1.25 \ (m, \ 4H), 1.20 \ (m, \ 2H), 0.95 \ (s, \ 9H), 0.86 \ (d, \ J \ 6.6 \ Hz, \ 6H). \) 13C NMR (125 MHz, CDCl3): \(\delta_c \ 174.2, 169.0, 108.4, 80.5, 52.1, 39.9, 39.0, 34.5, 34.2, 29.9, 27.9, 23.8, 22.6, 22.6, 21.3. \) HRMS \(m/z\) calculated for \([M+Na]^+\) C16H28NaO5 323.1834 found 323.1831.

General synthetic procedure, exemplified by (R)-dimethyl 2-hydroxy-2-(4-hydroxy-4-methylpentyl)succinate (8). 9.5 mg of 6 was dissolved in 1 mL of MeOH (previously dried and distilled with sodium). Next, 40 μL of NaOMe 1M (1.2 eq) was added. The reaction mixture was stirred for 1 h for a complete conversion. The solvent was then evaporated. The residue was dissolved in EtOAc and washed with HCl 0.1N. The organic layer was dried by Na2SO4, filtered, evaporated under vacuum. The crude was purified by flash chromatography on silica gel to give desired product. \([\alpha]_D^{24} -15\ (c \ 0.2, \text{MeOH})\); \(R_f \ 0.2\) (hexane/EtOAc, 4:1).

(R)-dimethyl 2-hydroxy-2-(4-hydroxy-4-methylpentyl)succinate (8) 1H NMR (500 MHz, CDCl3): \(\delta_H \ 3.80 \ (s, \ 3H), 3.67 \ (s, \ 3H), 2.93 \ (d, \ J \ 16.2 \ Hz, \ 1H), 2.68 \ (d, \ J \ 16.2 \ Hz, \ 1H), 1.81-1.83 \ (m, \ 2H), 1.54-1.60 \ (m, \ 2H), 1.45-1.49 \ (m, \ 2H), 1.22 \ (s, \ 6H). \) 13C NMR (125 MHz, CDCl3): \(\delta_c \ 175.6, 171.3, 75.2, 70.8, 52.9, 51.9, 43.5, 43.4, 39.6, 29.4, 29.1, 18.1. \) HRMS \(m/z\) calculated for \([M+Na]^+\) C12H22NaO6 285.1314 found 285.1319.

(R)-dimethyl 2-hydroxy-2-(4-methylpentyl)succinate (9). Following the general procedure above, 9 was obtained in 85% yield. \([\alpha]_D^{24} -16.7\ (c \ 0.3, \text{MeOH})\). \(R_f \ 0.45\) (hexane/EtOAc, 4:1).

1H NMR (500 MHz, CDCl3): \(\delta_H \ 3.80 \ (s, \ 3H), 3.67 \ (s, \ 3H), 2.93 \ (d, \ J \ 16.2 \ Hz, \ 1H), 2.68 \ (d, \ J \ 16.2 \ Hz, \ 1H), 1.43 – 1.66 \ (m, \ 5H), 1.13 \ 1.15 \ (m, \ 2H), 0.84 \ (d, \ J \ 6.6 \ Hz, \ 6H). \) 13C NMR (125 MHz, CDCl3): \(\delta_c \ 175.7, 171.4, 75.3, 52.9, 51.9, 43.4, 39.5, 38.8, 27.8, 22.6, 22.5, 20.1. \) HRMS \(m/z\) calculated for \([M+Na]^+\) C12H22NaO5 269.1365 found 269.1360

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