

Click chemistry on diterpenes: anti-inflammatory activity of the acetylenic derivatives of levopimaric acid and products of their transformations

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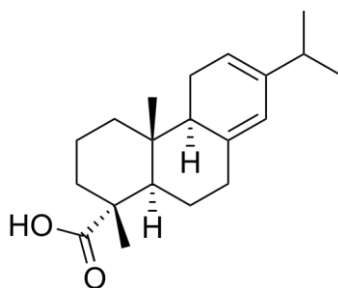
Abstract

Anti-inflammatory activity of amides of levopimaric acid (LpA) with acetylenic moiety and its esters with 4-R-triazolic substitutions was investigated using histamine-induced mice paw edema model. The synthesized LpA derivatives possess anti-inflammatory activity comparable to the activity of indomethacin.

Keywords: Acetylenes, Levopimaric acid, 1,2,3-triazole, Click Chemistry, diterpenoids, anti-inflammatory activity

Introduction

The rational use of plant metabolites as a platform for the discovery of new medicinal agents takes advantage of the structurally unique building blocks provided by nature for the rapid increase of molecular complexity. Diterpene resin acids (DRA), produced by conifers, combine an unusual stereochemically defined hydrocarbon scaffold with the possibility for possible functionalization via reactions at the carboxyl group. Levopimaric acid (LpA), the main component of pine oleoresin, is one of the most important in this family.



The relative content of Levopimaric acid in the resin of Scots pine *Pinus sylvestris* and its relative *Pinus hamata* reaches 27% and 36%, respectively.^{1,2} Along with the other diterpene resin acids, it plays a key role in providing defense against insects and microbial pathogens.³ In general, the abietene types of diterpene resin acid display a broad spectrum of biological activity. In particular, derivatives of Levopimaric acid show antibacterial, cardiovascular and antioxidant activities.¹

The general mode of biological activity of abietene derivatives is related to their anti-inflammatory properties. These properties are based on the suppression of inflammatory cytokines production and COX-2 expression.⁴ Dehydroabietic acid significantly decreases the production of pro-inflammatory mediators (MCP-1, TNF α , NO) in LPS-stimulated RAW264 macrophages and in mixed cell line of macrophages and adipocytes.⁵ Abietic acid was used as a building block for the preparation of heterocyclic derivatives close in activity to prednisolone but less toxic.⁶

Synthetic transformations based on the natural compounds expand medicinal potential of the nature's molecular treasury. For compounds of levopimaric acid family, an important role is played by the Diels-Alder reaction, usually with various benzo- and naphthoquinones. This efficient transformation provides a very convenient method for the isolation of LpA from complex mixture of product derived from the pine oleoresin. Furthermore, the recently reported adducts of LpA with benzoquinones display activity comparable with orthophen.⁷ The derivatives of dihydroquinopimaric acid show a particularly attractive combination of high anti-inflammatory activity with low toxicity.⁸

These examples illustrate the potential of abietic diterpenoids as a structural scaffold in the design of anti-inflammatory agents. In this aspect, LpA remains the less studied in this family of compounds. As the part of our ongoing research program dedicated to the investigation of plant metabolites produced by trees and herbs of Siberia,^{9,10} we present the first synthesis of acetylenic derivatives of LpA, selected transformations of these compounds, and studies of the anti-inflammatory activity with the aim of better understanding of the structure-activity relation.

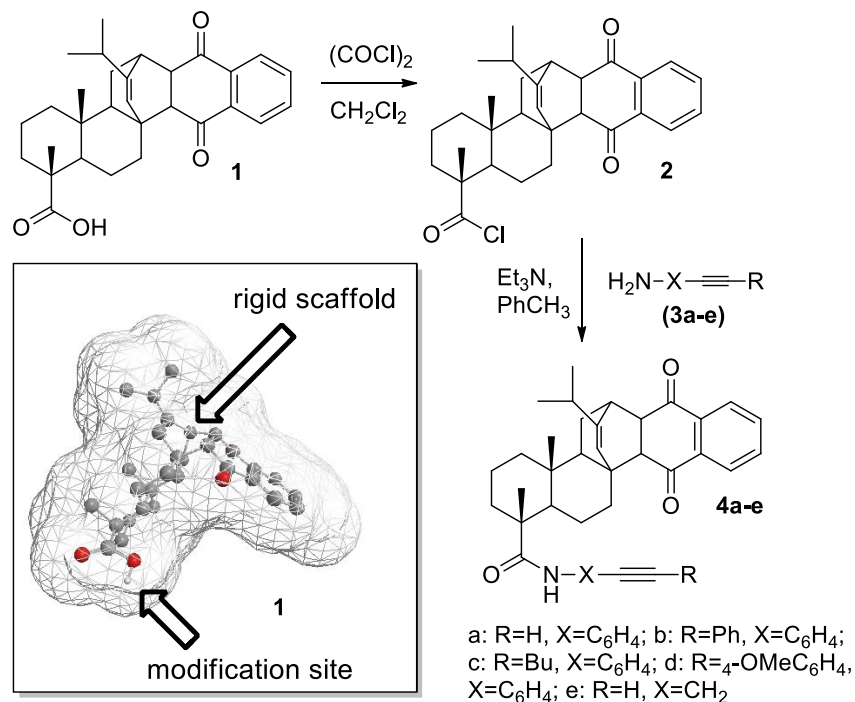
Presence of acetylenic moiety into natural^{11,12} and designed¹³ medicinal agents can play important role in the mechanism of their biological activity. Furthermore, it introduces possibilities for the further structural modifications. This strategy has been successfully applied to natural compounds of triterpenoid family.^{14,15} However, in the latest monograph on the

properties of DRA, where synthesis and properties of levopimaric acid are described in a chapter with 142 references,¹ data on acetylenic derivatives of this diterpene are absent.

Results and Discussion

Based on the above, we had chosen double modification of LpA using the Diels-Alder reaction with 1,4-naphthoquinone¹ with following introduction of alkyne groups via the LpA carboxyl.

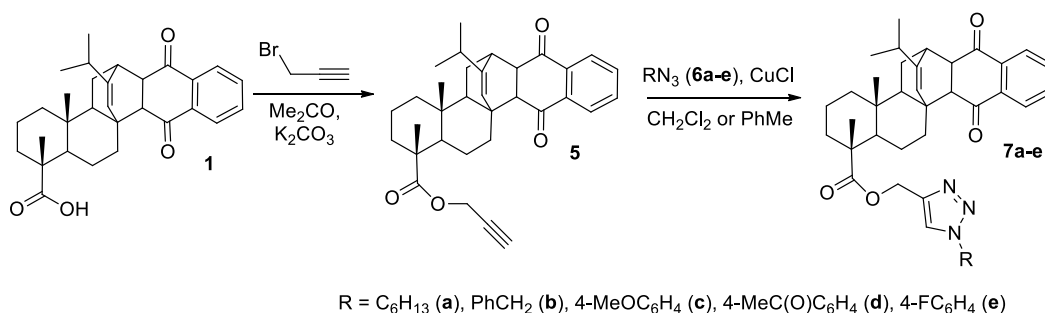
The first step of the sequence proceeds under very mild conditions (one week at the room temperature). The precipitated naphtholevopimaric acid **1** (NqLpA) is filtered to provide the 93% of the product.¹⁶ The NqLpA amides **4a-e** with alkyne substituents were prepared by condensation of NqLpA acid chloride **2** with the library of aminoacetylenes **3a-e** (see the SI part). We had used amides as the first set of targets based on our earlier reports of promising anti-inflammatory and anti-ulcer properties of related NqLpA amides.¹⁷



Scheme 1. Synthesis the NqLpA amides.

The NqLpA acid chloride **2** was synthesized by the reaction of acid **1** with oxalyl chloride in DCM at the room temperature. After removal of solvent from the reaction mixture, it was diluted with toluene and introduced in the reaction with the respective acetylenic amine **3** in triethylamine.

We had also explored the possibility of modifying the acetylenic derivatives of LpA via 1,3-dipolar cycloaddition with aryl and alkyl azides. The Cu-catalyzed version of this process works very well for terminal alkynes.^{18,19} The high yields, attractive regioselectivity and mild conditions account for the growing popularity of this reaction (the most popular of the “click” reactions) in the preparation of bioconjugates,²⁰⁻²⁷ including those of triterpenoid family.¹⁵ This remarkable selective and cost-effective process, leading to 1,2,3-triazoles, possessing anti-inflammatory activity²⁸ prompted us to synthesize the levopimaric acid-1,2,3-triazole hybrid. The key intermediate – the propargyl ester NqLpA **5** – prop-2-ynyl 16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate **5** was obtained by the reaction of acid **1** and propargyl bromide in the presence of K₂CO₃ in 30% isolated yield.



Scheme 2. Synthesis the propargyl ester NqLpA and the 1*H*-1,2,3-triazole derivatives of NqLpA.

In the following steps, the terminal alkyne **5** was introduced in the “click” reactions with alkyl and aryl azides **6a-e** in the presence of Cu(I) salts. Interestingly, alkyl azides are more reactive in comparison to their aromatic analogues. Whereas the reactions with alkyl azides proceed at the room temperature in 84-96 % yields, the reactions with aromatic azides require reflux in toluene (1*H*-1,2,3-triazole yields of ~70%).

The structure of all new acetylenes and the products of their transformations was established on the basis of analytical and spectroscopical methods (IR, ¹H and ¹³C NMR)

Anti-inflammatory activity of levopimaric acid derivatives

Two groups of levopimaric acid derivatives – amides with acetylenic moiety (**4-5**) and triazolyl esters (**7**) – were tested for their anti-inflammatory activities and structure-activity relationships using the histamine-induced paw edema model. Data in Table 1 illustrate that all synthesized derivatives injected intraperitoneally in mice (20 mg/kg dose) decrease paw edema by 23-47 percent relatively to control group. The same effect of indomethacin is 49 percent. The majority of the levopimaric acid derivatives (**4a-c**, **4e**, **7a-d**) have moderate anti-inflammatory activity - 1.5-2.0-fold lower than that of indomethacin. However, the activity of several esters (**4d**, **5**, **7e**) compares favorably with that of reference drug.

Table 1. Anti-inflammatory activity of levopimaric acid derivatives in the histamine-induced paw edema model

Substance	Inflammation index (%)	Anti-inflammatory activity (%)	Anti-inflammatory activity relative to indomethacin (%)
Control	34.2±2.2 ^{§§§}	0	-
4a	24.9±1.9 ^{*§§§}	27.2*	55.4 ^{§§§}
4b	25.2±2.0 ^{*§§§}	26.3*	53.6 ^{§§§}
4c	22.4±2.0 ^{**§}	34.5**	70.3 [§]
4d	19.3±1.8 ^{***}	43.6 ^{***}	88.8
4e	26.3±3.0 ^{*§§}	23.1*	47.0 ^{§§}
5	20.1±1.3 ^{***}	41.2 ^{***}	83.9
7a	25.9±2.1 ^{*§§§}	24.3*	49.5 ^{§§§}
7b	22.0±1.3 ^{***§§}	35.7 ^{***}	72.7 ^{§§}
7c	25.4±2.0 ^{*§§§}	25.7*	52.3 ^{§§§}
7d	23.2±1.3 ^{**§§§}	32.2**	65.6 ^{§§§}
7e	20.9±1.5 ^{***}	38.9 ^{***}	79.2
Indomethacin	17.4±0.9 ^{***}	49.1 ^{***}	100

*P<0.05, **P<0.01, ***P<0.001 relative to control

§ P<0.05, §§ P<0.01, §§§ P<0.001 relative to indomethacin

Comparative analysis of anti-inflammatory potencies of derivatives with acetylenic moiety in side residue (**4a**, **5**, **4e**) indicates that it depends on the structure of the linker at carbon group. Exchange of amide moiety to ester in the side residue causes significant 1.8 fold-increase of the anti-inflammatory effect (compound **5**). Introduction of a methoxy group in the diphenylacetylenic residue causes 1.7- fold increase of activity (**4d**→**4b**).

The activity of triazolic esters is also modulated by the substitution. For example, both change from an aliphatic to an aromatic substituents and fluorine introduction in the pendant aryl increase the activity. These data indicates that anti-inflammatory activity of amides and esters of LpA is sensitive to the nature of substitution and confirms the potential value of this class of biological active substances.

Conclusions

In conclusion, we reported the first synthesis of acetylene derivatives of LpA, their Cu-catalyzed alkyne-azide cycloadditions reactions, and anti-inflammatory activity of the new compounds. The possibility of combining the stereochemically defined polycyclic diterpene scaffold with the flexibility of structural “click” modifications should open new opportunities for the rational design of terpene-based medicinal agents. Encouragingly, the synthesized LpA derivatives possess anti-inflammatory activity comparable to the activity of indomethacin.

Experimental Section

General. Melting points were determined with a Kofler apparatus. Column chromatography was performed on Al₂O₃ (“Aldrich”) and the Silufol UV-254 plates were used for TLC analysis. The IR-spectra were recorded in KBr pellets on a “Bruker IFS 66” instrument. PdCl₂(PPh₃)₂, propargylamine, phenylacetylene, 4-iodaniline, Et₃N, CuCl were commercially available reactants (“Aldrich”). All the organic solvents were of analytical quality. Combustion analysis was performed with CHN-analyzer (Model 1106, “Carlo Erba”, Italy). NMR spectra were recorded on a «Bruker AM-400» 400.13 (¹H) and 100.61 MHz (¹³C) at 25 °C.

16-Isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2H-(6,12b-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylic acid (1) was prepared as described in ref.16.

General procedure of syntheses of aminoacetylenes 3. A mixture of 4-iodaniline (3mmol), corresponding alkyne-1 (3.2 mmol), PdCl₂(PPh₃)₂ (10 mg), CuI (5 mg) and Et₃N (2 mL) in benzene (25 mL) was stirred at 70 °C under the argon atmosphere. When the coupling was complete (TLC control) the products **3** was separated by column chromatography using hexane and then hexane–ethylacetate mixtures as eluent and recrystallized. Spectroscopic data and melting points for 4-(*p*-methoxyphenyl)ethynylaniline (**3d**),²⁹ 4-phenylethynylaniline (**3b**),³⁰ *n*-butylethynylaniline (**3c**),³¹ and *p*-aminophenylacetylene (**3a**)³² were consistent with the literature data.

General procedure of syntheses of amides 4. A mixture of 16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2H-(6,12b-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylic acid **1** (500 mg, 1.0 mmol) and oxalyl chloride (0.3 mL, 4.3 mmol) in of dry dichloromethane (10 mL) was stirred in argon for 3.5 h at room temperature. The solvent was evaporated at a reduced pressure. This crude product **2** without further purification was dissolved in of toluene (10 mL) and added to mixture Et₃N (0.25 mL) and amine **3** (1.1 mmol) in of toluene (5 ml). A mixture was stirred for 1-4 h at 75-80 °C under the argon atmosphere. The solvent was evaporated at a reduced pressure, the product **4** was purified by chromatography on

Al₂O₃ (elution with dichloromethane and mixture ethyl acetate - toluene) and crystallized from hexane.

***N*-(4-Ethynylphenyl)-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo-*[b]*chrysene-7,12-dione-1-carboxamide (4a).** The yield is 224 mg (40%), mp 150-153 °C (hexane); IR (KBr, v, cm⁻¹): 1680, 1726 (C=O); 2106 (C≡C). ¹H NMR (400 MHz, CDCl₃): δ 0.49 (3H, d, *J* 6.7 Hz), 0.58 (3H, s), 0.86 (3H, d, *J* 6.7 Hz), 0.90 (1H, m), 1.26 (3H, s), 1.37 (2H, m), 1.49-1.60 (6H, m), 1.65 (1H, m), 1.76-1.93 (5H, m), 2.85 (1H, d, *J* 8.6 Hz), 3.06 (1H, s), 3.07 (1H, br.s), 3.25 (1H, dd, *J* 3.0, 8.6 Hz), 5.06 (1H, s), 7.44-7.54 (5H, m), 7.60 (2H, m), 7.69 (1H, m), 7.78 (1H, m). ¹³C NMR (100 MHz, CDCl₃): δ 16.37, 17.07, 17.30, 19.13, 20.73, 21.73, 28.19, 33.24, 35.19, 37.15, 37.75, 38.53, 42.21, 42.38, 47.93, 50.33, 52.17, 56.37, 59.35, 77.36, 83.57, 117.82, 119.83, 125.13, 125.92, 125.96, 133.08, 133.24, 134.02, 136.85, 138.47, 138.64, 147.71, 176.83, 198.28, 198.80. Anal. Calcd for C₃₈H₄₁NO₃: C, 81.54; H, 7.38; N, 2.50%. Found: C, 81.23; H, 7.49; N, 2.65%.

***N*-(4-(Phenylethynyl)phenyl)-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo-*[b]*chrysene-7,12-dione-1-carboxamide (4b).** The yield is 204 mg (32%), mp 149-151 °C (hexane); IR (KBr, v, cm⁻¹): 1680 (C=O); 2214 (C≡C). ¹H NMR (CDCl₃, 400 MHz): δ 0.50 (3H, d, *J* 6.7 Hz), 0.58 (3H, s), 0.87 (3H, d, *J* 6.7 Hz), 1.01 (1H, m), 1.27 (3H, s), 1.29-1.58 (8H, m), 1.66 (1H, m), 1.78-1.92 (5H, m), 2.86 (1H, d, *J* 8.6 Hz), 3.07 (1H, br.s), 3.25 (1H, dd, *J* 2.9, 8.6 Hz), 5.07 (1H, s), 7.34 (3H, m), 7.45-7.57 (7H, m), 7.60 (2H, m), 7.70 (1H, m), 7.79 (1H, m). ¹³C NMR (100 MHz, CDCl₃): δ 16.38, 17.09, 17.31, 19.13, 20.74, 21.74, 28.19, 33.24, 35.20, 37.17, 37.76, 38.53, 42.22, 42.39, 47.92, 50.33, 52.18, 56.37, 59.35, 89.15, 89.35, 119.06, 119.91, 123.47, 125.14, 125.92, 125.96, 128.29, 128.48, 131.69, 132.53, 133.24, 134.02, 136.85, 138.01, 138.65, 147.70, 176.80, 198.30, 198.82. Anal. Calcd for C₄₄H₄₅NO₃: C, 83.11; H, 7.13; N, 2.20%. Found: C, 83.11; H, 7.13; N, 2.64%.

***N*-(4-(Hex-1-ynyl)phenyl)-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo-*[b]*chrysene-7,12-dione-1-carboxamide (4c).** The yield is 205 mg (33%), mp 121-123 °C (hexane); IR (KBr, v, cm⁻¹): 1680 (C=O); 2230 (C≡C). ¹H NMR (400 MHz, CDCl₃): δ 0.50 (3H, d, *J* 6.7 Hz), 0.58 (3H, s), 0.86 (3H, d, *J* 6.7 Hz), 0.96 (3H, t, *J* 7.3 Hz), 1.04 (1H, dd, *J* 6.7, 12.4 Hz), 1.25 (3H, s), 1.28-1.68 (13H, m), 1.78-1.91 (5H, m), 2.41 (2H, t, *J* 7.0 Hz), 2.86 (1H, d, *J* 8.6 Hz), 3.06 (1H, br), 3.25 (1H, dd, *J* 2.9, 8.6 Hz), 5.06 (1H, s), 7.37 (2H, m), 7.41 (1H, s), 7.47 (2H, m), 7.60 (2H, m), 7.69 (1H, m), 7.78 (1H, m). ¹³C NMR (100 MHz, CDCl₃): δ 13.82, 16.36, 17.08, 17.31, 19.13, 19.28, 20.73, 21.72, 33.19, 28.21, 31.02, 33.24, 35.20, 37.16, 37.77, 38.53, 42.21, 42.39, 47.85, 50.30, 52.19, 56.37, 59.36, 80.35, 90.10, 119.88, 120.02, 125.18, 125.92, 125.96, 132.37, 133.22, 134.01, 136.86, 137.29, 138.67, 147.69, 176.69, 198.27, 198.82. HRMS: *m/z* [M-2H]⁺ calcd 615.3707, found 613.3550 for C₄₂H₄₉NO₃.

***N*-(4-(4-Methoxyphenylethynyl)phenyl)-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)-benzo-*[b]*chrysene-7,12-dione-1-carboxamide (4d).** The yield is 240 mg (36%), mp 149-151 °C (hexane); IR (KBr,

v, cm^{-1}): 1680 (C=O); 2214 (C≡C). ^1H NMR (400 MHz, CDCl_3): δ 0.50 (3H, d, J 6.7 Hz), 0.58 (3H, s), 0.86 (3H, d, J 6.7 Hz), 1.03 (1H, m), 1.26 (3H, s), 1.28-1.58 (8H, m), 1.65 (1H, m), 1.78-1.91 (5H, m), 2.86 (1H, d, J 8.6 Hz), 3.07 (1H, br), 3.25 (1H, dd, J 2.9, 8.6 Hz), 3.83 (3H, s), 5.06 (1H, s), 6.88 (2H, m), 7.45-7.56 (7H, m), 7.60 (2H, m), 7.69 (1H, m), 7.78 (1H, m). ^{13}C NMR (100 MHz, CDCl_3): δ 16.37, 17.08, 17.31, 19.13, 20.73, 21.74, 28.20, 33.23, 35.21, 37.17, 37.77, 38.53, 42.21, 42.39, 47.89, 50.31, 52.18, 55.44, 56.37, 59.35, 88.02, 89.12, 114.13, 115.59, 119.43, 119.94, 125.16, 125.92, 125.95, 132.33, 133.13, 133.21, 134.00, 136.85, 137.73, 138.65, 147.69, 159.67, 176.76, 198.27, 198.80. Anal. Calcd for $\text{C}_{45}\text{H}_{47}\text{NO}_4$: C, 81.17; H, 7.11; N, 2.10%. Found: C, 81.12; H, 7.03; N, 2.19%.

***N*-Prop-2-ynyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)-benzo[*b*]chrysene-7,12-dione-1-carboxamide (4e).** The yield is 120 mg (24%), mp 110-112 °C (hexane); IR (KBr, v, cm^{-1}): 1661, 1678 (C=O); 2120, 3310 (C≡CH). ^1H NMR (400 MHz, CDCl_3): δ 0.49 (3H, d, J 6.5 Hz), 0.54 (3H, s), 0.85 (3H, d, J 6.5 Hz), 1.04 (1H, dd, J 6.7, 11.6 Hz), 1.13 (3H, s), 1.21-1.71 (9H, m), 1.75-1.89 (5H, m), 2.25 (1H, s), 2.84 (1H, d, J 8.6 Hz), 3.05 (1H, br.s), 3.24 (1H, dd, J 2.7, 8.6 Hz), 4.05 (2H, m), 5.05 (1H, s), 5.87 (1H, br.s), 7.60 (2H, m), 7.70 (1H, m), 7.78 (1H, m). ^{13}C NMR (100 MHz, CDCl_3): δ 16.33, 16.78, 17.22, 19.12, 20.72, 21.53, 28.22, 29.79, 33.22, 35.11, 36.93, 37.76, 38.46, 42.23, 42.38, 46.89, 50.13, 52.20, 56.23, 59.38, 71.80, 80.00, 125.22, 125.92, 125.93, 133.20, 133.98, 136.87, 138.71, 147.60, 178.24, 198.31, 198.86. Anal. Calcd for $\text{C}_{33}\text{H}_{39}\text{NO}_3$: C, 79.64; H, 7.90; N, 2.81%. Found: C, 79.52; H, 8.34; N, 2.78%.

Prop-2-ynyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate (5). A mixture of 16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylic acid **1** (7 g, 15.2 mmol), propargyl bromide (3.6 g, 30.4 mmol) and K_2CO_3 (4.8 g, 45.6 mmol) in acetone (190 mL) was boiled for 7 h. Then mixture was filtered, the solvent was evaporated at a reduced pressure. The crude product was purified by column chromatography on Al_2O_3 (elution with toluene and dichloromethane). The yield is 2.3 g (30%), mp 175-176 °C (hexane); IR (KBr, v, cm^{-1}): 1680, 1722 (C=O); 2131, 3290 (C≡CH). ^1H NMR (400 MHz, CDCl_3): δ 0.49 (3H, d, J 6.7 Hz), 0.53 (3H, s), 0.85 (3H, d, J 6.7 Hz), 0.94 (1H, m), 1.14 (3H, s), 1.25 (2H, m), 1.35-1.58 (6H, m), 1.65 (1H, m), 1.72-1.90 (5H, m), 2.48 (1H, t, J 2.4 Hz), 2.87 (1H, d, J 8.6 Hz), 3.05 (1H, br(m)), 3.25 (1H, dd, J 2.9, 8.6 Hz), 4.68 (2H, m), 5.06 (1H, br.s), 7.61 (2H, m), 7.71 (1H, m), 7.79 (1H, m). ^{13}C NMR (100 MHz, CDCl_3): δ 16.23, 16.82, 17.10, 19.11, 20.73, 22.07, 28.21, 33.24, 35.02, 36.48, 37.82, 38.33, 42.24, 42.45, 47.28, 49.35, 52.23, 52.26, 56.23, 59.41, 74.75, 78.10, 125.19, 125.90, 125.93, 133.26, 134.02, 136.91, 138.70, 147.60, 177.93, 198.57, 198.86. Anal. Calcd for $\text{C}_{33}\text{H}_{38}\text{O}_4$: C, 79.48; H, 7.68%. Found: C, 79.69; H, 7.62%.

General procedure of syntheses of arylazides 6c-e. To a solution of amine in THF (10-20 mL) was added HCl (1-6 mL) and water (8-47 mL). The reaction mixture was stirred for 15 min at 0 °C, and a solution of NaNO_2 (4.4-29 mmol) in of water (3-23 mL) was added dropwise to the solution, keeping the temperature of the reaction mixture below 5 °C. The reaction mixture was

stirred for 15 min at 0 °C, and then to the cooled reaction mixture was added dropwise a solution of NaN₃ (5.5-36 mmol) in of water (4-23 mL). After the addition, the reaction mixture was stirred for 1 h at room temperature. The organic layer was extracted with dichloromethane, and the extract was washed with dilute hydrochloric acid (0.1 M) and dried over Na₂SO₄. The solvent was removed *in vacuo*. Hexane was added and the resulting precipitate was filtered off to afford of the azide.

4-Azidoanisole (**6c**),³³ *p*-azidoacetophenone (**6d**),³⁴ 1-azido-4-fluorobenzene (**6e**),³⁵ *n*-hexyl azide (**6a**),³⁶ benzyl azide (**6b**)³⁷ were prepared as described in the literature.

Syntheses of triazoles. Method A. A mixture of prop-2-ynyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12b-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate **5** (200 mg, 0.4 mmol), CuCl (4 mg, 0.04 mmol) and alkyl azide **6a,b** (0.4 mmol) in dichloromethane (4 mL) was stirred at room temperature for 4-5 h. The crude product was purified by chromatography on silica gel or Al₂O₃ (elution with dichloromethane and ethyl acetate) and crystallized from hexane.

(1-Hexyl-1*H*-1,2,3-triazol-4-yl)methyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12b-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate (7a**).** The yield is 240 mg (96%), mp 52-55 °C (hexane); IR (KBr, *v*, cm⁻¹): 1678, 1722 (C=O). ¹H NMR (400 MHz, CDCl₃) δ 0.48 (3H, d, *J* 6.7 Hz), 0.51 (3H, s), 0.84 (3H, d, *J* 6.7 Hz), 0.86-0.91 (4H, m), 1.11 (3H, s), 1.19-1.96 (22H, m), 2.84 (1H, d, *J* 8.6 Hz), 3.04 (1H, br.s), 3.23 (1H, dd, *J* 2.7, 8.6 Hz), 4.40 (2H, t, *J* 7.2 Hz), 5.04 (1H, s), 5.24 (1H, q, *J* 12.5, 25.7 Hz), 7.60 (3H, m), 7.69 (1H, m), 7.78 (1H, m). ¹³C NMR (100 MHz, CDCl₃): δ 14.12, 16.19, 16.78, 17.05, 19.10, 20.71, 22.07, 22.60, 26.30, 28.18, 30.38, 31.29, 33.23, 35.09, 36.55, 37.77, 38.30, 42.22, 42.38, 47.17, 49.41, 50.58, 52.18, 56.23, 58.02, 59.38, 123.64, 125.11, 125.87, 125.93, 133.26, 133.99, 136.87, 138.65, 143.25, 147.63, 178.73, 198.40, 198.86. Anal. Calcd for C₃₉H₅₁N₃O₄: C, 74.85; H, 8.21; N, 6.71%. Found: C, 74.80; H, 8.11; N, 6.81%.

(1-Benzyl-1*H*-1,2,3-triazol-4-yl)methyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12b-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate (7b**).** The yield is 210 mg (84%), mp 72-74 °C (hexane); IR (KBr, *v*, cm⁻¹): 1678, 1722 (C=O). ¹H NMR (400 MHz, CDCl₃): δ 0.48 (3H, d, *J* 6.9 Hz), 0.50 (3H, s), 0.84 (3H, d, *J* 6.9 Hz), 0.91 (1H, m), 1.08 (3H, s), 1.20-1.89 (14H, m), 2.83 (1H, d, *J* 8.8 Hz), 3.04 (1H, s), 3.23 (1H, dd, *J* 2.7, 8.6 Hz), 5.03 (1H, s), 5.21 (2H, q, *J* 12.7, 15.4 Hz), 5.60 (2H, m), 7.36 (5H, m), 7.52 (1H, s), 7.61 (2H, m), 7.71 (1H, m), 7.79 (1H, m). ¹³C NMR (100 MHz, CDCl₃): δ 16.17, 16.76, 17.04, 19.11, 20.70, 22.02, 28.18, 33.22, 35.10, 36.48, 37.77, 38.28, 42.21, 42.38, 47.15, 49.40, 52.17, 54.35, 56.20, 57.90, 59.38, 123.69, 125.11, 125.87, 125.95, 128.29, 128.92, 129.29, 133.26, 134.00, 134.73, 136.88, 138.65, 143.77, 147.64, 178.63, 198.40, 198.83. Anal. Calcd for C₄₀H₄₅N₃O₄: C, 76.04; H, 7.18; N, 6.65%. Found: C, 76.14; H, 7.47; N, 6.76%.

Method B. A mixture of prop-2-ynyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12b-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate **5** (200 mg, 0.4 mmol), CuCl (4 mg, 0.04 mmol) and aryl azide **6c-e** (0.4 mmol) in 6 mL of toluene was

boiled for 1 h. The product was purified by chromatography on silica gel or Al₂O₃ (elution with dichloromethane and ethyl acetate) and crystallized from hexane.

(1-(4-Methoxyphenyl)-1*H*-1,2,3-triazol-4-yl)methyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate (7c). The yield is 180 mg (70%), mp 91-93 °C (hexane); IR (KBr, v, cm⁻¹): 1678, 1722 (C=O). ¹H NMR (400 MHz, CDCl₃): δ 0.47 (3H, d, *J* 7.0 Hz), 0.52 (3H, s), 0.84 (3H, d, *J* 7.0 Hz), 0.90 (1H, m), 1.12 (3H, s), 1.25-1.86 (14H, m), 2.85 (1H, d, *J* 8.6 Hz), 3.04 (1H, br.s), 3.24 (1H, dd, *J* 2.9, 8.6 Hz), 3.88 (3H, s), 5.04 (1H, s), 5.31 (2H, q, *J* 12.6, 28.5 Hz), 7.07 (2H, d, *J* 9.13 Hz), 7.61 (2H, m), 7.70 (3H, m), 7.78 (1H, m), 7.96 (1H, s). ¹³C NMR (100 MHz, CDCl₃): δ 16.22, 16.82, 17.06, 19.10, 20.71, 22.16, 28.18, 33.23, 35.18, 36.62, 37.79, 38.33, 42.22, 42.43, 47.24, 49.49, 52.21, 55.79, 56.24, 58.02, 59.39, 115.00, 122.16, 122.57, 125.12, 125.89, 125.93, 130.55, 133.25, 134.00, 136.90, 138.69, 143.87, 147.65, 160.08, 178.77, 198.40, 198.84. Anal. Calcd for C₄₀H₄₅N₃O₅: C, 74.16; H, 7.00; N, 6.49%. Found: C, 74.59; H, 7.32; N, 6.43%.

(1-(4-Acetylphenyl)-1*H*-1,2,3-triazol-4-yl)methyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate (7d). The yield is 184 mg (70%), mp 97-99 °C (hexane); IR (KBr, v, cm⁻¹): 1681, 1724 (C=O). ¹H NMR (400 MHz, CDCl₃): δ 0.48 (3H, d, *J* 6.7 Hz), 0.52 (3H, s), 0.85 (3H, d, *J* 6.7 Hz), 0.91 (1H, m), 1.13 (3H, s), 1.20-1.89 (14H, m), 2.67 (3H, s), 2.85 (1H, d, *J* 8.6 Hz), 3.05 (1H, br.s), 3.24 (1H, dd, *J* 2.9, 8.6 Hz), 5.04 (1H, s), 5.33 (2H, q, *J* 12.8, 17.8 Hz), 7.61 (2H, m), 7.68 (1H, m), 7.79 (1H, m), 7.96 (2H, m), 8.13 (1H, s), 8.19 (2H, m). ¹³C NMR (100 MHz, CDCl₃): δ 16.21, 16.84, 17.07, 19.12, 20.71, 22.22, 26.85, 28.20, 33.24, 35.25, 36.66, 37.85, 38.36, 42.19, 42.45, 47.30, 49.53, 52.19, 56.33, 57.87, 59.44, 120.48, 121.77, 125.08, 125.90, 125.99, 130.36, 133.29, 134.01, 136.91, 137.18, 138.65, 140.12, 144.59, 147.78, 178.75, 196.75, 198.37, 198.72. Anal. Calcd for C₄₁H₄₅N₃O₅: C, 74.63; H, 6.87; N, 6.37%. Found: C, 74.89; H, 7.06; N, 6.07%.

(1-(4-Fluorophenyl)-1*H*-1,2,3-triazol-4-yl)methyl-16-isopropyl-1,4a-dimethyl-1,2,3,4,4a,4b,5,6a,7,12,12a,13,14,14a-pentadecahydro-2*H*-(6,12*b*-etheno)benzo[*b*]chrysene-7,12-dione-1-carboxylate (7e). The yield is 180 mg (71%), mp 78-80 °C (hexane); IR (KBr, v, cm⁻¹): 1678, 1722 (C=O); ¹H NMR (400 MHz, CDCl₃): δ 0.48 (3H, d, *J* 6.5 Hz), 0.52 (3H, s), 0.84 (3H, d, *J* 6.5 Hz), 0.92 (1H, m), 1.13 (3H, s), 1.21-1.88 (14H, m), 2.85 (1H, d, *J* 8.5 Hz), 3.05 (1H, br.s), 3.24 (1H, dd, *J* 3.0, 8.5 Hz), 5.04 (1H, s), 5.32 (2H, q, *J* 12.8, 16.8 Hz), 7.29 (2H, m), 7.61 (2H, m), 7.69 (1H, m), 7.80 (3H, m), 8.00 (1H, s). ¹³C NMR (100 MHz, CDCl₃): δ 16.20, 16.82, 17.06, 19.11, 20.71, 22.17, 28.18, 33.24, 35.22, 36.61, 37.81, 38.34, 42.21, 42.42, 47.26, 49.53, 52.19, 56.27, 57.92, 59.41, 116.88, 117.11, 122.16, 122.95, 123.03, 125.06, 125.89, 125.97, 133.30, 134.02, 136.89, 138.63, 144.25, 147.72, 178.76, 198.41, 198.79. Anal. Calcd for C₃₉H₄₂FN₃O₄: C, 73.68; H, 6.66; F, 2.99; N, 6.61%. Found: C, 73.87; H, 7.02; F, 3.08; N, 6.60%.

Pharmacological experiments

The experiments were carried out using outbred female mice in accordance with the Guidelines

for the Care and Use of Laboratory Animals. The animals were given standard granulated food and water ad libitum.

Histamine-induced edema model

Inflammatory edema was induced by subplanar injection of 0.05 ml 0.01% histamine in water solution into the hind paw of female mice. The test compounds were administered intraperitoneally in 20 mg/kg b.w. dose (as aqueous-Tween-80 suspension) one hour before the histamine injection. The reference agent indomethacin ('Fluka BioChemica') was administered in the 20 mg/kg dose. The control group of animals received a water-Tween-80 mixture. The animals were sacrificed by cervical dislocation 5 hours after the phlogogenic agent injection, the mouse paws were cut off at the ankle joint and weighed. The ratio of the difference in weight between the treated and untreated hind paws to the weight of the untreated hind paw was used as an index of inflammation. The results were analyzed using "STATISTICA 6" software. The differences were significant at $p < 0.05$. The anti-inflammatory activity was presented as a difference between 100% and percentage of inflammation index relative to control group.

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References

1. Tolstikov, G. A.; Tolstikova, T. G.; Shults, E. E.; Tolstikov, S. E.; Khvostov, M. V. *In Resin acids from Russian forest conifers. Chemistry and Pharmacology*; Trofimov, B. A.; Ed.; Academic Publishing House "GEO": Novosibirsk, 2011, 394pp (Russ.).
2. Kersten, P. J.; Kopper, B. J.; Raffa, K. F.; Illman, B. L. *J. Chem. Ecol.* **2006**, *32*, 2679.
<http://dx.doi.org/10.1007/s10886-006-9191-z>
3. Trapp, S.; Croteau, R. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **2001**, *52*, 689.
<http://dx.doi.org/10.1146/annurev.arplant.52.1.689>
4. Takahashi, N.; Kawada, T.; Goto, T.; Kim, Chu-S.; Taimatsu, A.; Egawa, K.; Yamamoto, T.; Jisaka, M.; Nishimura, K.; Yokota, K.; Yu, R.; Fushiki, T. *FEBS Lett.* **2003**, *550*, 190.
[http://dx.doi.org/10.1016/S0014-5793\(03\)00859-7](http://dx.doi.org/10.1016/S0014-5793(03)00859-7)
5. Kang, M. S.; Hirai, S.; Goto, T.; Kuroyanagi, K.; Lee, J. Y.; Uemura, T.; Ezaki, Y.; Takahashi, N.; Kawada, T. *Biochem. Biophys. Res. Commun.* **2008**, *369*, 333.
<http://dx.doi.org/10.1016/j.bbrc.2008.02.002>

6. Abdulla M. M. *Monatsh. Chem.* **2008**, *139*, 697.
<http://dx.doi.org/10.1007/s00706-007-0804-1>
7. Flekhter, O. B.; Tretyakova, E. V.; Makara, N.S.; Gabdrakhmanova, S.F.; Baschenko, N.Zh.; Galin, F. Z.; Zarudii, F. S.; Tolstikov, G. A. *Pharm. Chem. J.* **2003**, *37*, 142.
<http://dx.doi.org/10.1023/A:1024538715025>
8. Kazakova, O. B.; Tretyakova, E. V.; Smirnova, I. E.; Spirikhin, I. V.; Tolstikov, G. A.; Chudov, I. V.; Bazekin, G. V.; Ismagilova, A. K. *Russ. J. Bioorg. Chem.* **2010**, *36*, 257.
<http://dx.doi.org/10.1134/S1068162010020160>
9. Shults, E. E.; Raldugin, V. A.; Volcho, K. P.; Salakhutdinov, N. F.; Tolstikov, G. A. *Russ. Chem. Rev.* **2007**, *76*, 655.
<http://dx.doi.org/10.1070/RC2007v076n07ABEH003717>
10. Vasilevsky, S. F.; Govdi, A. I.; Shult's, E. E.; Shakirov, M. M.; Alabugin I. V.; Tolstikov, G. A. *Proc. Russ. Acad. Sci.* **2009**, *424*, 631.
11. Dembitsky, V. M.; Levitsky, D. O. *Nat. Prod. Commun.* **2006**, 405.
12. Galm, U.; Hager, M. H.; Van Lanen, S. G., Ju, J.; Thorson, J. S.; Shen, B. *Chem. Rev.* **2005**, *105*, 739.
<http://dx.doi.org/10.1021/cr030117g>
13. Breiner, B.; Kaya, K.; Roy, S.; Yang, W.-Y.; Alabugin, I. V. *Org. Biomol. Chem.* **2012**, *10*, 3974.
<http://dx.doi.org/10.1039/c2ob00052k>
14. Vasilevsky, S. F.; Govdi, A. I.; Shults, E. E.; Shakirov, M. M.; Sorokina, I. V.; Tolstikova, T. G.; Baev, D. S.; Tolstikov, G. A.; Alabugin, I. V. *Bioorg. Med. Chem.* **2009**, *17*, 516.
<http://dx.doi.org/10.1016/j.bmc.2009.05.059>
15. Vasilevsky, S. F.; Govdi, A. I.; Sorokina, I. V.; Tolstikova, T. G.; Tolstikov, G. A.; Mamatuyk, V. I.; Alabugin, I. V. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 62.
<http://dx.doi.org/10.1016/j.bmcl.2010.11.072>
16. Vafina, G. F.; Fazlyev, R.R. Galin, F.Z.; Spirikhin, L.V. *Russ.J. Org. Chem.* **2009**, *45*, 500.
<http://dx.doi.org/10.1134/S1070428009040058>
17. Kazakova, O. B.; Tretyakova, E. V.; Kukovinets, O. S.; Tolstikov, G. A.; Nazzyrov, T.I.; Chudov, I.V.; Ismagilova, A. F. *Russ. J. Bioorg. Chem.* **2010**, *36*, 762.
<http://dx.doi.org/10.1134/S1068162010060130>
18. Kolb, H. C.; Finn, M. G.; Sharpless, B. K. *Angew. Chem. Int. Ed.* **2001**, *40*, 2004.
[http://dx.doi.org/10.1002/1521-3773\(20010601\)40:11<2004::AID-ANIE2004>3.0.CO;2-5](http://dx.doi.org/10.1002/1521-3773(20010601)40:11<2004::AID-ANIE2004>3.0.CO;2-5)
19. Finn, M. G.; Fokin, V. V. *Chem. Soc. Rev.* **2010**, *39*, 1221.
<http://dx.doi.org/10.1039/c003740k>
20. Sletten, E. M.; Bertozzi, C. R. *Angew. Chem. Int. Ed.* **2009**, *48*, 6974.
<http://dx.doi.org/10.1002/anie.200900942>
21. Debets, M. F.; van Berkel, S. S.; Dommerholt, J.; Dirks, A. J.; Rutjes, F. P. J. T.; van Delft, F.L. *Acc. Chem. Res.* **2011**, *44*, 805.
<http://dx.doi.org/10.1021/ar200059z>

22. Best, M. D.; Rowland, M. M.; Bostic, H. E. *Acc. Chem. Res.* **2011**, *44*, 686.
<http://dx.doi.org/10.1021/ar200060y>
23. Codelli, J. A.; Baskin, J. M.; Agard, N. J.; Bertozzi, C. R. *J. Am. Chem. Soc.* **2008**, *130*, 11486.
<http://dx.doi.org/10.1021/ja803086r>
24. Ning, X.; Guo, J.; Wolfert, M. A. Boons, G.-J. *Angew. Chem. Int. Ed.* **2008**, *47*, 2253.
<http://dx.doi.org/10.1002/anie.200705456>
25. Poloukhine, A. A.; Mbua, N. E.; Wolfert, M. A.; Boons, G.-J.; Popik, V. V. *J. Am. Chem. Soc.* **2009**, *131*, 15769.
<http://dx.doi.org/10.1021/ja9054096>
26. Gold, B.; Schevchenko, N.; Bonus, N.; Dudley, G. B.; Alabugin, I. V. *J. Org. Chem.* **2012**, *77*, 75.
<http://dx.doi.org/10.1021/jo201434w>
27. Gold, B.; Dudley, G. B.; Alabugin, I. V. *J. Am. Chem. Soc.* **2013**, *135*, 1558.
<http://dx.doi.org/10.1021/ja3114196>
28. Porto de Oliveira Assis, Sh.; Targino da Silva, M.; de Oliveira, R. N.; deMenezes Lima, V. L. *The ScientificWorld Journal* **2012**, Article ID 925925, 7 pages.
29. Csekei, M.; Novak, Z.; Kotschy, A. *Tetrahedron* **2008**, *64*, 975.
<http://dx.doi.org/10.1016/j.tet.2007.10.031>
30. Endo, Y.; Songkram, C.; Yamasaki, R.; Tanatani, A.; Kagechika, H.; Takaishi, K.; Yamaguchi, K. *J. Organomet. Chem.* **2002**, *657*, 48.
[http://dx.doi.org/10.1016/S0022-328X\(02\)01322-0](http://dx.doi.org/10.1016/S0022-328X(02)01322-0)
31. Lin, C.-H.; Wang, Y.-J.; Lee, C.-F. *Eur. J. Org. Chem.* **2010**, 4368.
32. Takahashi, S.; Kuroyama, Y.; Sonogashira, K.; Hagihara, N. *Synthesis* **1980**, *8*, 627.
<http://dx.doi.org/10.1055/s-1980-29145>
33. Ugi, I.; Perlinger, H.; Behringer, L. *Chem. Ber.* **1958**, *55*, 2330.
<http://dx.doi.org/10.1002/cber.19580911111>
34. Pokhodylo, N. T.; Savka, R. D.; Matiichuk, V. S.; Obushak, N. D. *Russ. J. Gen. Chem.* **2009**, *79*, 309.
<http://dx.doi.org/10.1134/S1070363209020248>
35. Gilyarov, V. A.; Kabachnik, M. I. *Zh. Obsh. Khim.* **1966**, *36*, 282.
36. Henkel, K.; Weygand, F. *Ber.* **1943**, *76*, 812.
<http://dx.doi.org/10.1002/cber.19430760809>
37. Moulin, Fr. *Helv. Chim. Acta* **1952**, *5*, 167.
<http://dx.doi.org/10.1002/hlca.19520350124>