Ultrasound promoted synthesis of chromeno[2,3-*b*]pyridines and their evaluation as lipid peroxidation inhibitors

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Abstract

A series of 2,3-disubstituted-5-oxochromeno[2,3-*b*]pyridine derivatives were synthesized under ultrasound irradiation, and also under conventional methods, from the reaction of 3-cyanochromones with some active methylene compounds in the presence of a stoichiometric base. The structures of the products were unambiguously elucidated by 1D and 2D NMR experiments. Full assignment of all ¹H and ¹³C NMR chemical shifts has been achieved. In addition, the lipid peroxidation inhibition of the synthesized chromenopyridine derivatives was evaluated.

Keywords: Antioxidant activity, chromeno[2,3-*b*]pyridines, 3-cyanochromones, lipid peroxidation inhibition, sonochemistry

Introduction

In recent decades, ultrasound irradiation has been used more and more frequently in organic synthesis.^{1,2} The use of ultrasound to accelerate reactions has proven to be a particularly important tool for meeting the Green Chemistry goals of minimization of waste and reduction of energy requirements.^{3,4} Applications of ultrasonic irradiation are playing an increasing role in chemical processes, especially in cases where classical methods require drastic conditions or prolonged reaction times. The excellent review of Cella and Stefani⁵ clearly showed the importance of taking advantage of the unique features of ultrasound–assisted reactions to synthesize heterocyclic ring systems.

Heterocyclic compounds are highly ranked among pharmaceutically important natural and synthetic materials. The remarkable ability of heterocyclic nuclei to serve both as biomimetics and active pharmacophores has largely contributed to their unique value as traditional key elements of numerous drugs. As a result, in the last years the preparation of natural-product-like libraries has become a foremost strategy for the discovery of new drugs.⁶ In addition, merging privileged structures constitutes a powerful strategy to reach drug-like bioactive compounds in a fast and productive manner.⁷ In this context, the discovery of core structures having this potential is one of the main challenges for the pharmaceutical industry in its search for rich sources of new molecular entities.⁸

In this way, the chromenopyridine scaffold may be regarded as the fusion of two main fragments: of a chromene and of a pyridine moiety. Both fragments display a wide range of biological activities; they are present in many drugs and have been extensively studied. Chromenes form a group of either naturally or synthetically obtained compounds possessing a broad diversity of biological activities with an impact in the treatment of inflammation and ulcers,⁹ schizophrenia,¹⁰ cancer,¹¹ etc.

On the other hand, pyridines are ubiquitous in nature and pivotal in synthetic and medicinal chemistry constituting a useful core for biomedical research.¹²

Concerning chromenopyridines, a wide array of biological activities has been described for compounds containing this core in their structures. This small, compact scaffold is rich in intermolecular interactions, and can be easily functionalized and diversified. Owing to such interesting properties, the medicinal chemistry around this structural class is emerging as an active subject of research, not only in academia, but especially in the pharmaceutical industry. In this context, some chromenopyridines have been reported to be alpha adrenergic antagonists,¹³ potent bronchodilating agents,¹⁴ and useful in the treatment of bronchitis and asthma.¹⁵

In view of our continuous interest in merging privileged structures containing a chromene moiety¹⁶⁻¹⁸ we decided to pursue a more detailed investigation concerning the synthesis of chromeno[2,3-*b*]pyridines from the reaction of 3-cyanochromones with active methylene compounds in the presence of a stoichiometric base.

Results and Discussion

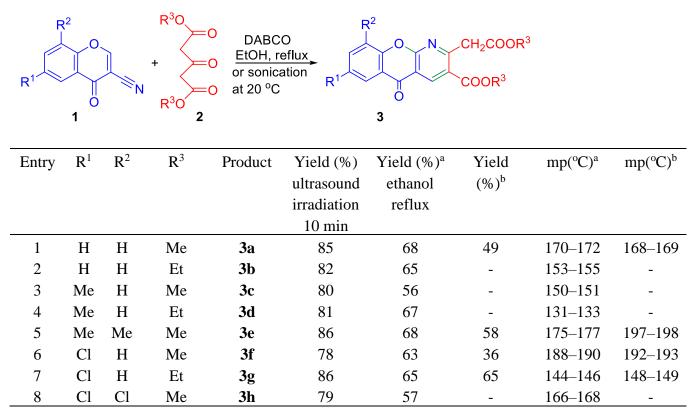
Chemistry

Some early reports^{19–21} concerning the synthesis of chromeno[2,3-*b*]pyridines from 3cyanochromones with some active methylene compounds by refluxing in ethanol in the presence of piperidine are reported in the literature. In addition recently, the reaction between cyanochromones with some active methyl and methylene compounds was studied in the presence of DBU, as a base, whereupon chromenopyridines were isolated in satisfactory yields.²² In the present study we decided to pursue an investigation concerning the possibility of improving the reaction yield by using Green Chemistry conditions, namely ultrasound irradiation, and to provide a structure elucidation based on detailed NMR study since the proposed in the literature structure of chromeno[2,3-*b*]pyridines^{19–22} was based on inadequate ¹H-NMR data. Thus initially, 3cyanochromone, dimethyl acetonedicarboxylate and p-toluidine were refluxed in ethanol for 4 h, whereupon the yellow solid, which was formed upon cooling of the reaction mixture, was filtered off and recrystallized from ethanol to give the chromenopyridine **3a** in 60% yield. The reaction was repeated with other bases and the results are given in Table 1.

Entry	Base	Yield (%)	
1	<i>p</i> -toluidine	60	
2	<i>p</i> -chloroaniline	45	
3	DMAP	35	
4	DABCO	68	
5	DBU	unidentified	
		product	
6	cinchonine	47	
7	DABCO with	85	
	ultrasound		
	irradiation		

Table 1. The effect of different bases on the yield of product 3a

Table 2. Synthesis of chromenopyridines 3



^aIsolated product in this work. ^bRef. 21.

In all cases chromenopyridine **3a** was isolated in varying yields with exception the reaction with DBU, where most probably a three component unidentified product incorporating DBU was isolated. Finally, the reaction was investigated under ultrasound irradiation, using DABCO as a base, since DABCO provided the best results under conventional heating. Analytically, the reaction mixture in a small amount of ethanol was subjected to ultrasound irradiation for 10 min, whereupon after initial solvation of the reactants the chromenopyridine **3a** was crystallized out. After the reaction conditions were optimized, the generality of the reaction was investigated with a variety of cyanochromones, with methyl and ethyl acetonedicarboxylates, and the results summarized in Table 2.

Next, the reaction was investigated by using ethyl acetylacetate and ethyl benzoylacetate and it was found, that in order for the reaction to proceed, the stronger base DBU was necessary. No reaction was observed when *p*-toluidine was used as a base. To the contrary, concerning ultrasound irradiation DABCO was found to be the best base. The results are presented in Table 3.

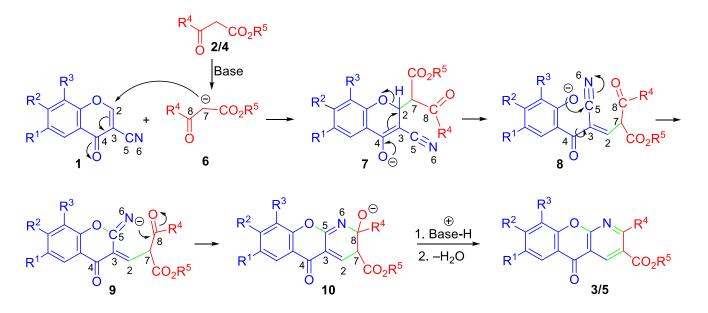
$R^{1} \xrightarrow{0}_{0} N + \frac{R^{2}}{EtO} \xrightarrow{0}_{0} \frac{DBU, EtOH}{or DABCO} \xrightarrow{0}_{0} \frac{1}{20 \circ C} R^{1} \xrightarrow{0}_{0} COOEt$										
Entry	\mathbb{R}^1	\mathbb{R}^2	Product	Yield (%)	Yield (%) ^a	mp (°C)	Yield (%)	mp (°C)		
				ultrasound	ethanol					
				irradiation	reflux					
				DABCO	DBU					
1	Н	Me	5a	76 (40 min)	47	161–163	63 ^b	162 ^b		
2	Н	Ph	5b	70 (15 min) ^c	55	146–148	41 ^d	270 ^d		
3	Me	Me	5c	78 (35 min)	55	165–167	63 ^b	170 ^b		
4	Me	Ph	5d	72 (15 min)	57	174–176	-	-		
5	Cl	Me	5e	76 (40 min)	53	166–168	65 ^b	169 ^b		
6	Cl	Ph	5f	76 (20min)	59	167–169	-			

Table 3. Synthesis of chromenopyridines 5

^aThis work. ^bRef. 19. ^c16 % yield when DBU was used. ^dRef. 22.

Concerning the reaction mechanism, it is conceivable that the initial attack of the anion 6, formed from the active methylene group compound 2 (or 4), to C2 of cyanochromone 1 is accompanied by the chromone ring opening to intermediate 8. After rotation around the C3 – C4 bond and recyclization through attack of the pyran oxygen anion to the cyano-carbon C5 intermediate 9 is formed, which subsequently undergoes a new cyclo-condensation to furnish 10. Finally, after loss of water, the chromenopyridine 3 (or 5) is formed (Scheme 1). For the isolated products structure 11, depicted in Figure 2, could also be proposed. However, this structure was excluded on the basis

of rigorous spectroscopic analysis (mainly 2D NMR spectra) as described in the following session. Moreover, minor products with a structure analogous to **11** were not identified.



Scheme 1. Mechanistic rationalization for the formation of chromenopyridines 3/5. The numbering used is arbitrary for the understanding the skeletal carbon rearrangement.

Structure assignment of the new compounds

The assigned molecular structures of all chromenopyridines **3** and **5** were based on rigorous spectroscopic analysis including IR, MS, NMR (¹H, ¹³C, H–H COSY, HMQC and HMBC) and elemental analysis data.

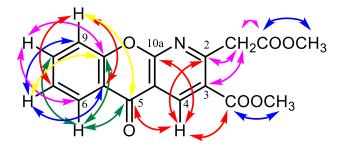


Figure 1. HMBC correlations via ${}^{2}J_{CH}$, ${}^{3}J_{CH}$ and ${}^{4}J_{CH}$ (in yellow color) couplings observed in compound **3a**, and the numbering of its skeleton.

A typical ¹H NMR spectrum of the chromenopyridine derivative **3a** contains a characteristic sequence of four aromatic chromone protons, at δ 7.63 (dd), 7.82 (ddd), 7.49 (ddd) and 8.33 (dd) with the corresponding carbons at 118.6, 136.1, 125.3, and 126.9 ppm, respectively. Very characteristic and of good diagnostic value is the one-proton signal at δ 9.32 corresponding to the 4–H with its carbon at 141.1 ppm correlating in HMBC with carbons at 160.6 (C10a), 161.6 (C2),

165.0 (3-CO), and 176.7 (C5) ppm (Figure 1). The ester methyl protons correlate with the ester carbonyl carbons and the 2-CH₂ protons with carbons at 161.6 (C2), 123.4 (C3), and at 170.1 (2-CO) distinguishing thus their neighbor carbonyl group. In the case of structure **11** (Figure 2) the singlet at δ 9.32 corresponding to the proton 4-H could not have HMBC correlation with 1-CO carbonyl carbon, due to the five intervening bonds, proving thus that the structure of **3a** corresponds beyond doubt to the isolated products.

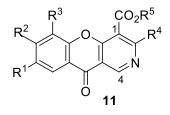


Figure 2. Alternative product not isolated.

Biological evaluation

The synthesized chromenopyridines **3** and **5** contain two pharmacophore groups: i) the chromone ring and ii) the pyridine ring. Thus, they might be able to behave as hybrid molecules in terms of biological activity. Therefore, some of the new compounds were screened for their antioxidant activity according to a previously reported methodology,^{23,24} and more specifically, for their reducing ability (RA) by interaction with the stable free radical 1,1-diphenylpicrylhydrazyl (DPPH) and for their inhibition on lipid peroxidation by interaction with the water soluble azo compound 2,2"-azobis(2-amidinopropane) dihydrochloride (AAPH) and the results are compared to well known antioxidant agents e.g. nordihydroguairetic acid (NDGA), and 6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (trolox). Also, since both pharmacophores have been separately implicated in the inhibition of lipoxygenase a preliminary test *in vitro* for their anti-LOX activity should be of interest.^{18,24} Each in vitro experiment was performed at least in triplicate, the results were averaged and the standard deviation of absorbance was less than 10% of the mean. The results are presented in Table 4.

The tested compounds present limited reducing activity expressed as RA%, through their interaction with DPPH. However, using AAPH as azo initiator of lipid peroxidation, all tested compounds (**3a**, **3c**, **3d**, **3f**, **3g**, and **5f**) except **5c** were found to highly inhibit lipid peroxidation. Concerning the inhibition of soybean lipoxygenase activity compounds **3c**, and **3g** seem to be the most biologically active and their anti-lipid peroxidation activity proceeds in parallel to their enzyme inhibition. Therefore, these two derivatives could be used as lead for the design of new potent antioxidants.

Product		DPPH (RA %)		LOX		
		at 100 µM				
	20 min	60 min		Inh %	$IC_{50} \mu M$	
				100 µM		
3 a	14	9	85	43		
3c	14	12	76	61	82	
3d	15	12	73	35		
3f	16	14	75	30		
3g	12	12	75	59	85	
5c	2	9	19 % at	no		
			100 µM			
5 f	8	no	80	17		
NDGA	83	86		93		
Trolox			55			

Table 4. Reducing activity (RA%) by interaction with DPPH; In vitro inhibition values (IC₅₀) of lipid peroxidation induced by AAPH; In vitro % inhibition of soybean lipoxygenase (LOX %)

Conclusions

In the present study an investigation concerning the optimization of the reaction conditions under normal heating and, alternatively, under sonication leading to the synthesis of chromeno[2,3-b]pyridines **3** and **5** is described. The biological results showed that all tested compounds except **5c** were found to highly inhibit lipid peroxidation. Compounds **3c** and **3g** are promising leads chromenopyridines combining increased anti-lipid peroxidation and lipoxygenase inhibitory activities. Full assignment of all ¹H and ¹³C NMR chemical shifts has been unambiguously achieved. The earlier proposed plausible mechanistic scheme was confirmed.

Experimental Section

General. Melting points were measured on a Kofler hot–stage and are uncorrected. Column chromatography was carried out using Merck silica gel (70–230 mesh). TLC was performed using precoated silica gel glass plates 0.25 mm containing fluorescent indicator UV₂₅₄ purchased from Macherey–Nagel using a 3:1 mixture of petroleum ether–ethyl acetate. Petroleum ether refers to the fraction boiling between 60 and 80 °C. NMR spectra were recorded at 25 °C on a Bruker AVANCE III 300 spectrometer at 300 MHz for ¹H and 75 MHz for ¹³C, using CDCl₃, DMSO-*d*₆ or their mixture as solvent, as indicated. Some samples are measured on a Varian 500 MHz spectrometer, as indicated. The chemical shifts are expressed in δ values (ppm) relative to TMS as internal standard

for ¹H and relative to TMS (0.00 ppm), or to CDCl₃ (77.05 ppm) or to DMSO-*d*₆ (40.0 ppm) for ¹³C NMR spectra. Coupling constants ^{*n*}J are reported in Hz. IR spectra were recorded on a Perkin-Elmer 1600 series FTIR spectrometer and are reported in wave numbers (cm⁻¹). Low resolution LC–MS (ESI, 1.65 eV) mass spectra were recorded on LC–MS–2010 EV system (Shimadzu). Elemental analyses performed with a Perkin–Elmer 2400-II CHN analyzer. Structural assignments of the derived compounds were established by analysis of their IR, MS and NMR spectra (¹H, ¹³C, COSY H-H, HMQC and HMBC). Ultrasound for sonication is generated using the instrument: ELMA TRANSSONIC T310 (Germany); operating frequency 35 kHz, 220 V, 50 Hz, 140 mA; bath dimensions 190×85×55 mm.

5-Oxo-5*H***-chromeno[2,3-***b***]pyridine-carboxylates.** A solution of acetonedicarboxylate (0.5 mmol) and DABCO (0.5 mmol) in ethanol (5 mL) were refluxed for 15 min, whereupon 3-cyanochromone (0.5 mmol) was added and the reaction mixture was refluxed further for 4 h. Upon cooling the product was crystallized, filtered by Gooch funnel, washed on the funnel with diethyl ether and recrystallized from ethanol. Alternatively, the reaction mixture in a small amount of ethanol (2 mL) was subjected to ultrasound irradiation for 10 min using the water bath at 20 °C, whereupon after initial solvation of the reactants the chromenopyridine product was crystallized out.

Methyl 2-(2-methoxy-2-oxoethyl)-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (3a). Yield: 0.139 g (85%), yellow solid, mp 170–172 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1731 (C=O), 1676 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 3.75 (s, 3H, 2-OCH₃), 3.97 (s, 3H, 3-OCH₃), 4.42 (s, 2H, 2-CH₂), 7.49 (ddd,** *J* **8.0, 7.4, 0.5 Hz, 1H, 7-H), 7.63 (dd,** *J* **8.0, 0.5 Hz, 1H, 9-H), 7.82 (ddd,** *J* **8.0, 7.4, 1.2 Hz, 1H, 8-H), 8.33 (dd,** *J* **8.0, 1.2 Hz, 1H, 6-H), 9.32 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 43.8 (2-CH₂), 52.3 (2-OCH₃), 52.7 (3-OCH₃), 115.2 (C4a), 118.6 (C9), 121.6 (C5a), 123.4 (C3), 125.3 (C7), 126.9 (C6), 136.1 (C8), 141.1 (C4), 155.5 (C9a), 160.6 (C10a), 161.6 (C2), 165.0 (3-CO), 170.1 (2-CO), 176.7 (C5). LC–MS (ESI, 1.65 eV):** *m/z* **(%) 382 [98, (M + Na + MeOH)⁺], 366 [70 (M + K)⁺], 350 [100, (M + Na)⁺], 328 [10, (M + H)⁺]. Anal. Calcd for C₁₇H₁₃NO₆ (327.29): C, 62.39; H, 4.00; N, 4.28. Found: C, 62.34; H, 4.12; N, 4.34.**

Ethyl 2-(2-ethoxy-2-oxoethyl)-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (3b). Yield: 0.146 g (82%), yellow solid, mp 153–155 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1724 (C=O), 1673 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 1.26 (t,** *J* **7.1 Hz, 3H, 2-OCH₂CH₃), 1.44 (t,** *J* **7.1 Hz, 3H, 3-OCH₂CH₃), 4.20 (q,** *J* **7.1 Hz, 2H, 2-OCH₂CH₃), 4.41 (s, 2H, 2-CH₂), 4.42 (q,** *J* **7.1 Hz, 2H, 3-OCH₂CH₃), 7.48 (ddd,** *J* **8.0, 7.3, 0.7 Hz, 1H, 7-H), 7.63 (dd,** *J* **8.3, 0.7 Hz, 1H, 9-H), 7.82 (ddd,** *J* **8.3, 7.3, 1.6 Hz, 1H, 8-H), 8.34 (dd,** *J* **8.0, 1.6 Hz, 1H, 6-H), 9.31 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): δ 14.16 (2-CH₂CH₃), 14.24 (3-OCH₂CH₃), 44.1 (2-CH₂), 61.2 (2-OCH₂CH₃), 61.9 (3-OCH₂CH₃), 115.1 (C4a), 118.6 (C9), 121.7 (C5a), 123.9 (C3), 125.3 (C7), 126.9 (C6), 136.0 (C8), 140.9 (C4), 155.6 (C9a), 160.5 (C10a), 161.7 (C2), 164.6 (3-CO), 169.6 (2-CO), 176.8 (C5). LC–MS (ESI, 1.65 eV): m/z (%) 733 [60, (2×M + Na)⁺], 410 [100, (M + Na + MeOH)⁺], 378 [95, (M + Na)⁺], 356 [20, (M + H)⁺]. Anal. Calcd for C₁₉H₁₇NO₆ (355.35): C, 64.22; H, 4.82; N, 3.94. Found: C, 64.44; H, 4.70; N, 4.05.**

Methyl 2-(2-methoxy-2-oxoethyl-7-methyl)-5-oxo-5*H*-chromeno[2,3-*b*]pyridine-3-carboxylate (3c). Yield: 0.136 g (80%), yellow solid, mp 150–151 °C (ethanol); IR (KBr, v_{max} , cm⁻¹): 1739 (C=O), 1680 (C=O), 1670 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 2.49 (s, 3H, 7-CH₃), 3.74 (s, 3H, 2-OCH₃), 3.96 (s, 3H, 3-OCH₃), 4.41 (s, 2H, 2-CH₂), 7.51 (d, *J* 8.6 Hz, 1H, 9-H), 7.61 (dd, *J* 8.6, 1.9 Hz, 1H, 8-H), 8.10 (d, *J* 1.9 Hz, 1H, 6-H), 9.30 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): δ 20.9 (7-CH₃), 43.8 (2-CH₂), 52.3 (2-OCH₃), 52.7 (3-OCH₃), 115.2 (C4a), 118.4 (C9), 121.4 (C5a), 123.3 (C3), 126.3 (C6), 135.4 (C7), 137.3 (C8), 141.1 (C4), 153.9 (C9a), 160.7 (C10a), 161.5 (C2), 165.1 (3-CO), 170.0 (2-CO), 176.8 (C-5). LC–MS (ESI, 1.65 eV): *m/z* (%) 412 [80, (M + K + MeOH)⁺], 396 [78, (M + Na + MeOH)⁺], 380 [31, (M + K)⁺], 364 [100, (M + Na)⁺], 342 [18, (M + H)⁺]. Anal. Calcd for C₁₈H₁₅NO₆ (341.31): C, 63.34; H, 4.43; N, 4.10. Found: C, 63.22; H, 4.51; N, 4.18.

Ethyl 2-(2-ethoxy-2-oxoethyl-7-methyl)-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (3d).Yield: 0.149 g (81%), yellow solid, mp 131–133 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1731 (C=O), 1670 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 1.26 (t,** *J* **7.1 Hz, 3H, 2-CH₂CH₃), 1.43 (t,** *J* **7.1 Hz, 3H, 3-CH₂CH₃), 2.50 (s, 3H, 7-CH₃), 4.19 (q,** *J* **7.1 Hz, 2H, 2-CH₂CH₃), 4.40 (s, 2H, 2-CH₂), 4.41 (q,** *J* **7.1 Hz, 2H, 3-CH₂CH₃), 7.51 (d,** *J* **8.5 Hz, 1H, 9-H), 7.61 (dd,** *J* **8.5, 1.9 Hz, 1H, 8-H), 8.10 (d,** *J* **1.9 Hz, 1H, 6-H), 9.29 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 14.17 (2-CH₂CH₃), 14.26 (3-OCH₂CH₃), 20.9 (7-CH₃), 44.1 (2-CH₂), 61.2 (2-OCH₂CH₃), 61.9 (3-CH₂CH₃), 115.1 (C4a), 118.4 (C9), 121.4 (C5a), 123.8 (C3), 126.3 (C6), 135.3 (C7), 137.2 (C8), 140.9 (C4), 153.9 (C9a), 160.6 (C10a), 161.6 (C2), 164.7 (3-CO), 169.6 (2-CO), 176.9 (C5). LC–MS (ESI, 1.65 eV):** *m/z* **(%) 424 [65, (M + Na + MeOH)⁺], 392 [100, (M + Na)⁺]. Anal. Calcd for C₂₀H₁₉NO₆ (369.37): C, 65.03; H, 5.18; N, 3.79. Found: C, 65.22; H, 5.11; N, 3.88.**

Methyl 2-(2-methoxy-2-oxoethyl-7,8-dimethyl)-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (3e). Yield: 0.153 g (86%), yellow solid, mp 175–177 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1735 (C=O), 1725 (C=O), 1673 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 2.39 (s, 3H, 8-CH₃), 2.44 (s, 3H, 7-CH₃), 3.74 (s, 3H, 2-OCH₃), 3.96 (s, 3H, 3-OCH₃), 4.41 (s, 2H, 2-CH₂), 7.37 (s, 1H, 9-H), 8.04 (s, 1H, 6-H), 9.30 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 19.3 (8-CH₃), 20.8 (7-CH₃), 43.8 (2-CH₂), 52.3 (2-OCH₃), 52.7 (3-OCH₃), 115.3 (C4a), 118.7 (C9), 119.4 (C5a), 123.1 (C3), 126.5 (C6), 134.7 (C7), 141.0 (C4), 147.1 (C8), 154.0 (C9a), 160.7 (C10a), 161.2 (C2), 165.1 (3-CO), 170.1 (2-CO), 176.6 (C-5). LC–MS (ESI, 1.65 eV):** *m/z* **(%) 733 [100, (2×M + Nα)⁺], 410 [60, (M + Na + MeOH)⁺], 394 [31, (M + K)⁺], 378 [65, (M + Na)⁺], 356 [20, (M + H)⁺]. Anal. Calcd for C_{19H17}NO₆ (355.34): C, 64.22; H, 4.82; N, 3.94. Found: C, 63.22; H, 4.51; N, 4.18.**

Methyl 2-(2-methoxy-2-oxoethyl-7-chloro)-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (3f**). Yield: 0.141 g (78%), yellow solid, mp 188–190 °C (ethanol); IR (KBr, v_{max} , cm⁻¹): 1732 (C=O), 1721 (C=O), 1673 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 3.74 (s, 3H, 2-OCH₃), 3.97 (s, 3H, 3-OCH₃), 4.42 (s, 2H, 2-CH₂), 7.58 (d, *J* 8.9 Hz, 1H, 9-H), 7.74 (dd, *J* 8.9, 2.5 Hz, 1H, 8-H), 8.28 (d, *J* 2.5 Hz, 1H, 6-H), 9.29 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): δ 43.6 (2-CH₂), 52.2 (2-OCH₃), 52.6 (3-OCH₃), 114.7 (C4a), 120.3 (C9), 122.5 (C5a), 123.7 (C3), 126.1 (C6), 131.2 (C7), 136.0 (C8), 141.0 (C4), 153.8 (C9a), 160.3 (C10a), 161.8 (C2), 164.7 (3-CO), 169.7 (2-CO), 175.6 (C-5). LC–MS (ESI, 1.65 eV): *m/z* (%) 416/418 [100, (M + Na + MeOH)⁺], 400/402 [28, (M + K)⁺], 384/386 [75, (M + Na)⁺], 362/364 [5, (M + H)⁺]. Anal. Calcd for C₁₇H₁₂ClNO₆ (361.73): C, 56.45; H, 3.34; N, 3.87. Found: C, 56.52; H, 3.41; N, 3.73.

Ethyl 2-(2-ethoxy-2-oxoethyl-7-chloro)-5-oxo-5*H*-chromeno[2,3-*b*]pyridine-3-carboxylate (3g). Yield: 0.168 g (86%), yellow solid, mp 144–146 °C (ethanol); IR (KBr, v_{max} , cm⁻¹): 1728 (C=O),

1667 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 1.26 (t, *J* 7.1 Hz, 3H, 2-CH₂CH₃), 1.44 (t, *J* 7.1 Hz, 3H, 3-CH₂CH₃), 4.19 (q, *J* 7.1 Hz, 2H, 2-CH₂CH₃), 4.40 (s, 2H, 2-CH₂), 4.42 (q, *J* 7.1 Hz, 2H, 3-CH₂CH₃), 7.59 (d, *J* 8.9 Hz, 1H, 9-H), 7.76 (dd, *J* 8.9, 2.5 Hz, 1H, 8-H), 8.27 (d, *J* 2.5 Hz, 1H, 6-H), 9.26 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): δ 14.0 (2-CH₂CH₃), 14.1 (3-CH₂CH₃), 43.9 (2-CH₂), 61.1 (2-CH₂CH₃), 61.9 (3-CH₂CH₃), 114.6 (C4a), 120.3 (C9), 124.1 (C3), 125.8 (C5a), 127.5 (C6), 130.9 (C7), 136.0 (C8), 140.7 (C4), 155.0 (C9a), 160.1 (C10a), 161.8 (C2), 164.2 (3-CO), 169.3 (2-CO), 175.6 (C5). LC–MS (ESI, 1.65 eV): *m*/*z* (%) 444/446 [55, (M + Na + MeOH)⁺], 428/430 [15, (M + K)⁺], 412/414 [50, (M + Na)⁺], 390/392 [5, (M + H)⁺], 313/315 [100, (M – 76)⁺]. Anal. Calcd for C₁₉H₁₆CINO₆ (389.79): C, 58.55; H, 4.14; N, 3.59. Found: C, 58.62; H, 4.32; N, 3.68.

Methyl 2-(2-methoxy-2-oxoethyl-7,9-duchloro)-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (3h**). Yield: 0.157 g (79%), yellow solid, mp 166–168 °C (ethanol); IR (KBr, v_{max} , cm⁻¹): 1725 (C=O), 1677 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 3.73 (s, 3H, 2-OCH₃), 3.97 (s, 3H, 3-OCH₃), 4.45 (s, 2H, 2-CH₂), 7.85 (d, *J* 2.3 Hz, 1H, 8-H), 8.20 (d, *J* 2.3 Hz, 1H, 6-H), 9.28 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃+DMSO-*d*₆): δ 43.4 (2-CH₂), 51.9 (2-OCH₃), 52.6 (3-OCH₃), 114.3 (C4a), 123.0 (C5a), 123.2 (C3), 124.1 (C9), 124.4 (C6), 130.4 (C7), 135.5 (C8), 140.5 (C4), 149.7 (C9a), 159.6 (C-10a), 161.9 (C2), 164.3 (C2''), 169.4 (C2'), 174.7 (C-5). LC–MS (ESI, 1.65 eV): *m/z* (%) 450/452/454 [60, (M + Na + MeOH)⁺], 434/436/438 [50, (M + K)⁺], 418/420/422 [100, (M + Na)⁺], 396/398/400 [10, (M + H)⁺]. Anal. Calcd for C₁₇H₁₁Cl₂NO₆ (396.18): C, 51.54; H, 2.80; N, 3.54. Found: C, 51.62; H, 2.73; N, 3.58.

Ethyl 2-methyl-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (5a). Yield: 0.108 g (76%), yellow solid, mp 161–163 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1731 (C=O), 1676 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 1.45 (t,** *J* **7.2 Hz, 3H, 3-CH₂CH₃), 2.99 (s, 3H, 2-CH₃), 4.44 (q,** *J* **7.2 Hz, 2H, 3-CH₂CH₃), 7.47 (ddd,** *J* **8.0, 7.2, 0.8 Hz, 1H, 7-H), 7.63 (dd,** *J* **8.3, 0.8 Hz, 1H, 9-H), 7.80 (ddd,** *J* **8.3, 7.2, 1.5 Hz, 1H, 8-H), 8.32 (dd,** *J* **8.0, 1.5 Hz, 1H, 6-H), 9.20 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 14.1 (3-CH₂CH₃), 25.3 (2-CH₃), 61.6 (3-CH₂CH₃), 114.0 (C4a), 118.6 (C9), 121.7 (C5a), 123.5 (C3), 125.0 (C7), 126.6 (C6), 135.6 (C8), 140.5 (C4), 155.5 (C9a), 160.4 (C10a), 161.6 (C2), 164.8 (3-CO), 176.9 (C5). LC–MS (ESI, 1.65 eV):** *m***/***z* **(%) 338 [100, (M + Na + MeOH)⁺], 306 [10, (M + Na)⁺], 284 [39, (M + H)⁺]. Anal. Calcd for C₁₆H₁₃NO₄ (283.28): C, 67.84; H, 4.63; N, 4.94. Found: C, 67.97; H, 4.75; N, 5.02.**

Ethyl 2-phenyl-5-oxo-5H-chromeno[2,3-b]pyridine-3-carboxylate (**5b**). Yield: 0.121 g (70%), yellow solid, mp 146–148 °C (ethanol); IR (KBr, v_{max} , cm⁻¹): 1713 (C=O), 1664 (C=O). ¹H NMR (500 MHz, CDCl₃): δ 1.17 (t, *J* 7.2 Hz, 3H, 3-CH₂CH₃), 4.26 (q, *J* 7.2 Hz, 2H, 3-CH₂CH₃), 7.46–7.51 (m, 4H, 3',4',5',7-H), 7.65 (d, *J* 8.3 Hz, 1H, 9-H), 7.65–7.68 (m, 2H, 2',6'-H), 7.82 (ddd, *J* 8.3, 7.5, 1.4 Hz, 1H, H-8), 8.35 (dd, *J* 8.0, 1.4 Hz, 1H, 6-H), 9.12 (s, 1H, 4-H). ¹³C NMR (125 MHz, CDCl₃): δ 13.8 (3-CH₂CH₃), 62.0 (3-CH₂CH₃), 114.4 (C4a), 118.6 (C9), 121.7 (C5a), 125.16 (C3), 125.18 (C7), 126.9 (C6), 128.3 (C2',6'), 128.9 (C3',5'), 130.0 (C4'), 136.0 (C8), 138.4 (C1'), 140.2 (C4), 155.8 (C9a), 160.1 (C10a), 163.8 (C2), 166.5 (CO), 176.9 (C5). LC–MS (ESI, 1.65 eV): *m/z* (%) 400 [100, (M + Na + MeOH)⁺], 368 [80, (M + Na)⁺], 346 [15, (M + H)⁺]. Anal. Calcd for C₂₁H₁₅NO₄ (345.35): C, 73.03; H, 4.38; N, 4.06. Found: C, 72.94; H, 4.32; N, 4.11.

Ethyl 2,7-dimethyl-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (5c). Yield: 0.116 g (78%), yellow solid, mp 165–167 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1724 (C=O), 1668 (C=O). ¹H**

NMR (300 MHz, CDCl₃): δ 1.44 (t, *J* 7.2 Hz, 3H, 3-CH₂CH₃), 2.49 (s, 3H, 7-CH₃), 2.98 (s, 3H, 2-CH₃), 4.44 (q, *J* 7.2 Hz, 2H, 3-CH₂CH₃), 7.51 (d, *J* 8.5 Hz, 1H, 9-H), 7.60 (dd, *J* 8.2, 2.0 Hz, 1H, 8-H), 8.10 (d, *J* 2.0 Hz, 1H, 6-H), 9.20 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): δ 14.3 (3-CH₂CH₃), 20.9 (7-CH₃), 25.6 (2-CH₃), 61.7 (3-CH₂CH₃), 114.0 (C4a), 118.4 (C9), 121.3 (C5a), 123.4 (C3), 126.2 (C6), 135.2 (C7), 137.1 (C8), 140.6 (C4), 153.0 (C9a), 160.5 (C10a), 165.0 (3-CO), 166.6 (C2), 177.1 (C5). LC–MS (ESI, 1.65 eV): *m*/*z* (%) 386 [50, (M + MeOH + K +H₂O)⁺], 352 [100, (M + Na + MeOH)⁺], 330 [10, (M + MeOH + H)⁺], 298 [37, (M + H)⁺], Anal. Calcd for C₁₇H₁₅NO4 (297.31): C, 68.68; H, 5.09; N, 4.71. Found: C, 68.77; H, 4.99; N, 4.60.

Ethyl 7-methyl-2-phenyl-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (5d). Yield: 0.129 g (72%), yellow solid, mp 174–176 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1733 (C=O), 1681 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 1. 46 (t,** *J* **7.2 Hz, 3H, 3-CH₂CH₃), 3.00 (s, 3H, 7-CH₃), 4.44 (q,** *J* **7.2 Hz, 2H, 3-CH₂CH₃), 7.46–7.70 (m, 7H, 2-Ph,8,9-H), 8.11 (s, 1H, 6-H), 9.10 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 15.0 (3-CH₂CH₃), 25.6 (7-CH₃), 61.8 (3-CH₂CH₃), 113.7 (C4a), 120.3 (C9), 122.5 (C5a), 123.8 (C3), 126.1 (C6), 128.0 (C2',6'), 129.0 (C3',5'), 131.0 (C7), 133.0 (C4'), 135.9 (C8), 140.5 (C4), 140.6 (C1'), 153.8 (C9a), 160.1 (C10a), 164.7 (3-CO), 167.1 (C2), 175.9 (C5). LC–MS (ESI, 1.65 eV):** *m/z* **(%) 414 [100, (M + Na + MeOH)⁺], 382 [80, (M + Na)⁺], 360 [15, (M + H)⁺]. Anal. Calcd for C₂₂H₁₇NO₄ (359.37): C, 73.53; H, 4.77; N, 3.90. Found: C, 73.44; H, 4.66; N, 4.01.**

Ethyl 7-chloro-2-methyl-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (5e). Yield: 0.122 g (77%), yellow solid, mp 166–168 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1741 (C=O), 1659 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 1.46 (t,** *J* **7.2 Hz, 3H, 3-CH₂CH₃), 3.00 (s, 3H, 2-CH₃), 4.44 (q,** *J* **7.2 Hz, 2H, 3-CH₂CH₃), 7.58 (d,** *J* **8.9 Hz, 1H, 9-H), 7.74 (dd,** *J* **8.9, 2.5 Hz, 1H, 8-H), 8.27 (d,** *J* **2.5 Hz, 1H, 6-H), 9.19 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 14.3 (3-CH₂CH₃), 25.6 (2-CH₃), 61.8 (3-CH₂CH₃), 113.7 (C4a), 120.3 (C9), 122.5 (C5a), 123.9 (C3), 126.2 (C6), 131.1 (C7), 135.9 (C8), 140.6 (C4), 153.8 (C9a), 160.2 (C10a), 164.8 (3CO), 167.2 (C2), 175.9 (C5). LC–MS (ESI, 1.65 eV):** *m/z* **(%) 372/380 [100, (M + Na + MeOH)⁺], 356/358 [20, (M + K)⁺], 318/320 [80, (M + H)⁺]. Anal. Calcd for C₁₆H₁₂ClNO₄ (317.72): C, 60.48; H, 3.81; N, 4.41. Found: C, 60.57; H, 3.90; N, 4.30.**

Ethyl 7-chloro-2-phenyl-5-oxo-5*H***-chromeno[2,3-***b***]pyridine-3-carboxylate (5f). Yield: 0.144 g (76%), yellow solid, mp 167–169 °C (ethanol); IR (KBr, v_{max}, cm⁻¹): 1736 (C=O), 1685 (C=O). ¹H NMR (300 MHz, CDCl₃): \delta 1.17 (t,** *J* **7.1 Hz, 3H, 3-CH₂CH₃), 4.25 (q,** *J* **7.1 Hz, 2H, 3-CH₂CH₃), 7.45–7.52 (m, 3H, 3',4',5'-H), 7.60 (d,** *J* **8.8 Hz, 1H, 9-H), 7.63–7.68 (m, 2H, 2',6'-H), 7.74 (dd,** *J* **8.8, 2.0 Hz, 1H, 8-H), 8.30 (d,** *J* **2.0 Hz, 1H, 6-H), 9.09 (s, 1H, 4-H). ¹³C NMR (75 MHz, CDCl₃): \delta 13.8 (3-CH₂CH₃), 62.0 (3-CH₂CH₃), 114.2 (C4a), 120.4 (C9), 122.8 (C3), 125.7 (C5a), 126.3 (C6), 128.4 (C2',6'), 129.0 (C3',5'), 130.1 (C7), 131.3 (C4'), 136.0 (C8), 138.4 (C1'), 140.3 (C4), 154.2 (C9a), 159.5 (C10a), 164.1 (3-CO), 166.4 (C2), 175.9 (C5). LC–MS (ESI, 1.65 eV):** *m/z* **(%) 434/436 [100, (M + Na + MeOH)⁺], 402/404 [75, (M + Na)⁺], 380/382 [15, (M + H)⁺]. Anal. Calcd for C₂₁H₁₄ClNO₄ (379.79): C, 66.41; H, 3.72; N, 3.69. Found: C, 66.34; H, 3.68; N, 3.78.**

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References

- 1. Luche, J.-L. Synthetic Organic Sonochemistry; Plenum Press: New York, 1998.
- 2. Mason, T. J. Advances in Sonochemistry; JAI Press: London, 1990.
- 3. Cintas, P.; Luche, J.-L. Green Chem. 1999, 1, 115–125.
- 4. Puri, S.; Kaur, B.; Parmar, A.; Kumar, H. *Curr. Org. Chem.* **2013**, *17*, 1790–1828. http://dx.doi/10.2174/13852728113179990018
- 5. Cella, R.; Stefani, H. A. *Tetrahedron* **2009**, *65*, 2619–2641. http://dx.doi/10.1016/j.tet.2008.12.027
- 6. Roth, H. J.; Kleemann, A.; Beiswenger, T. Drug Synthesis, In *Pharmaceutical Chemistry*; Wiley: New York, 1988, Vol. 1.
- 7. Costantino, L.; Barlocco, D. *Curr. Med. Chem.* **2006**, *13*, 65–85. <u>http://dx.doi/10.2174/092986706775197999</u>
- 8. Muegge, I. *Med. Res. Rev.* **2003**, *23*, 302–321. http://dx.doi/10.1002/med.10041
- Hegab, M. I.; Abdel-Fattah, A.-S. M.; Yousef, N. M.; Nour, H. F.; Mostafa, A. M.; Ellithey, M. Arch. Pharm. Chem. Life Sci. 2007, 340, 396–403. http://dx.doi/10.1002/ardp.200700089
- Unangst, P. C.; Capiris, T.; Connor, D. T.; Heffner, T. G.; Mackenzie, R. G.; Miller, S. R.; Pugsley, T. A.; Wise, L. D. J. Med. Chem. 1997, 40, 2688–2693. <u>http://dx.doi./10.1021/jm970170v</u>
- Giannouli, V.; Kostakis, I. K.; Pouli, N.; Marakos, P.; Kousidou, O. C.; Tzanakakis, G. N.; Karamanos, N. K. J. Med. Chem. 2007, 50, 1716–1719. <u>http://dx.doi./10.1021/jm061410m</u>
- 12. For recent reviews, see:

(a) Moreau, J.; Hurvois, J.-P.; Mbaye, M. D.; Renaud, J.-L. In: *Targets in Heterocyclic Systems*, Attanasi, O. A; Spinelli, D. Eds.; Italian Society of Chemistry, Rome, 2009; Vol. 13, pp. 201–230.

(b) Tanaka, H.; Namekata, I.; Komikado, C.; Kawanishi, T.; Shigenobu, K. *Curr. Top. Pharmacol.* **2007**, *11*, 1–15.

(c) Quirion, J.-C.; Leclerc, E.; Jubault, P. Product subclass 18: 1,2-dihydropyridines, 1,4-dihydropyridines, and derivatives. In *Science of Synthesis*, Thieme Verlag, Stuttgart, 2007, Vol. *33*, pp 601–628.

(d) Lavilla, R. *Curr. Org. Chem.* **2004**, *8*, 715–737. http://dx.doi/10.2174/1385272043370690

(e) Lavilla, R. J. Chem. Soc., Perkin Trans. 1 2002, 1141–1156.

- 13. Meyer, M. D.; Altenbach, R. J.; Basha, F. Z.; Carroll, W.A.; Drizin, I.; Kerwin, J. F. Jr.; Wendt, M. D.; Haight, A. R.; Zhang, W. U.S. Patent 6 046 207, April, 4, 2000.
- 14. Brown, R. E.; Puchalski, C.; Shavel, J. Jr. U.S. Patent 3 962 266, May, 1, 1975.
- 15. Connor, D. T.; Unangst, P. C.; Schwender, C. F.; Sorenson, R. J.; Carethers, M. E.; Puchalski, C.; Brown, R. E. J. Heterocyclic Chem. **1984**, 21, 1561–1564.
- 16. (a) Zarganes-Tzitzikas, T.; Terzidis, M. A.; Stephanidou-Stephanatou, J.; Tsoleridis, C. A.; Kostakis G. E. J. Org. Chem. 2011, 76, 9008–9014. <u>http://dx.doi/10.1021/jo201732g</u>
 (b) Terzidis, M. A.; Zarganes-Tzitzikas, T.; Tsimenidis, C.; Stephanidou-Stephanatou, J.; Tsoleridis, C. A.; Kostakis G.E. J. Org. Chem. 2012, 77, 9018–9028. http://dx.doi/10.1021/jo3014947
- Dimitriadou, E.; Stephanidou-Stephanatou, J.; Tsoleridis, C. A.; Hadjipavlou-Litina, D. J.; Kontogiorgis, C.; Kostakis, G. E. *Tetrahedron* 2014, 70, 2938–2943. <u>http://dx.doi.org/10.1016/j.tet.2014.03.031</u>
- 18. Ghosh, C. K; SinhaRoy, D. K.; Mukhopadhyay, K. K. J. Chem. Soc., Perkin Trans. 1 1979, 1964–1968.
- 19. Ghosh, C. K. Synth. Comm. 1978, 487-490.
- 20. Kubo, K.; Ukawa, K.; Kuzuna, S.; Nohara, A. Chem. Pharm. Bull. 1986, 1108–1117.
- 21. Ibrahim, M.A. *Synth. Commun.* **2009**, *39*, 3527–3545. http://dx.doi./10.1080/00397910902788141
- 22. Hadjipavlou-Litina, D.; Magoulas, G. E.; Bariamis, S. E.; Drainas, D.; Avgoustakis, K.; Papaioannou, D. *Bioorg. Med. Chem.* **2010**, *18*, 8204–8217. http://dx.doi.org/10.1016/j.bmc.2010.10.012
- 23. Neochoritis, C. G.; Tsoleridis, C. A.; Stephanidou-Stephanatou, J.; Kontogiorgis, C. A.; Hadjipavlou-Litina, D. J. J. Med. Chem. 2010, 53, 8409–8420. <u>http://dx.doi./10.1021/jm100739n</u>