Monosaccharidic mimetics of the sialyl Lewis$^X$ tetrasaccharide based on 2,7-dihydroxynaphthalene

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Dedicated to Professor Rainer Beckert on the occasion of his 60th birthday

Abstract
A potential monosaccharidic mimetic of the sialyl Lewis$^X$ tetrasaccharide (sLe$^X$) was identified based on an in silico docking study using the crystal structure of an E-selectin-sLe$^X$ complex. The chemical synthesis of the mimetic in an ortho-selective C-glycosylation is described. This compound and two close analogues were evaluated in a cell-based selectin binding assay where none of the tested mimetics showed an IC$_{50}$ below 1mM. This result can be explained by an unexpected 1C$_4$ conformation of the mannosyl residue which precludes the required binding of the Ca$^{2+}$-ion in E-selectin.

Keywords: Carbohydrates, phenols, C-glycosides, cell adhesion, molecular modelling

Introduction
Oligosaccharides are known to play an important role in cell recognition processes and in the communication between cells in higher organisms.$^1$ A prominent example is the recruitment of leucocytes during the inflammatory cascade, which is initiated by the interaction between selectins and their cognate oligosaccharide ligands such as sialyl Lewis$^X$ (sLe$^X$, 1) (Fig.1).$^2$ Undesired interactions caused by overexpression or dysregulation have been associated with various diseases e.g. asthma,$^3$ psoriasis,$^4$ reperfusion syndrome$^{1b}$ or the metastasis of tumors.$^5$ Therefore, some effort has been put into the development of selectin ligands$^6$ and their mimetics$^7$ as potential therapeutics.
Previous studies have shown that the three hydroxyl groups of the L-fucose\textsuperscript{8} and the carboxylic acid moiety of the sialic acid\textsuperscript{9} are essential for binding of sialyl Lewis\textsuperscript{X} (1) towards the selectins. However, the L-fucose portion has been successfully substituted by other monosaccharides with similar configuration such as D-arabinose,\textsuperscript{6a,6b} L-galactose\textsuperscript{7e,10} or D-mannose as demonstrated by Kogan’s functional mannosyl mimetic 2.\textsuperscript{7b,7c} Furthermore, the (S)-cyclohexyl lactic acid moiety successfully employed by Ernst and Thoma e.g. in compound 3 turned out to be a privileged mimetic for the sialic acid portion.\textsuperscript{7d,11} A potential general drawback of oligosaccharidic therapeutics is their instability against acidic hydrolysis and their degradation by glycosidases. For instance, O-fucosidic bonds are typical targets for ubiquitous fucosidases.\textsuperscript{12} An approach to overcome the metabolic lability of O-glycosidic bonds can be the use of C-glycosidic mimetics,\textsuperscript{7c,10} the synthesis of which can be effected along various routes.\textsuperscript{13} Here, we report on the preparation and biological evaluation of potential sLe\textsuperscript{X} analogues based on a mannosylated 2,7-dihydroxynaphthalene scaffold.

**Results and Discussion**

Virtual docking experiments using the crystal structure of the E-selectin/sLe\textsuperscript{X} complex and mimetics composed of various aromatic and heteroaromatic core structures in combination with (S)-cyclohexyl lactic acid and D-mannose revealed a good fit of compound 4 containing an α-C-mannosylated 2,7-dihydroxynaphthalene (Figure 1, Figure 2).

![Figure 1. Sialyl Lewis\textsuperscript{X} and mimetic 4 identified by virtual docking.](image-url)
**Figure 2.** Docking of sialyl Lewis\(^X\) (green) and mimetic 4 to E-selectin. The Ca\(^{2+}\)-ion bound by the protein is highlighted in purple.

The key step for the synthesis of 4 was a direct C-glycosylation of an electron rich phenol using a mannosyl trichloracetimidate as a reactive glycosyl donor.\(^{14}\) Reactions of this kind are suggested to proceed in a stepwise fashion and usually yield a single regioisomer in high stereoselectivity.\(^{13b,14-15}\) The triflate of \((S)\)-cyclohexyl lactic acid (6) was prepared from \(D\)-phenylalanine in four steps with 49% overall yield.\(^{16}\) Its reaction with an excess of 2,7-dihydroxynaphthalene in the presence of \(K_2CO_3\) gave the desired phenol ether 7 in 86% yield (Scheme 1).

![Scheme 1](image)

**Scheme 1.** Synthesis of the glycosyl acceptor 7. Reagents and conditions: (a) \(NaNO_2, 1.25\; M\; H_2SO_4\); (b) \(MeOH, DOWEX-50WX8 (H^+)\); (c) \(Rh-Al_2O_3, H_2, THF/H_2O\); (d) \(Tf_2O, 2,6\)-lutidine, DCM; (e) \(K_2CO_3, CH_3CN\).
Glycosyl donor 9 could be synthesized in five steps from D-mannose in 66% overall yield.\(^{17}\) The direct glycosylation of 9 to the acceptor 7 in the presence of TMSOTf gave the C-glycoside 10 in 61% yield but unfortunately with the undesired β-configuration at the anomeric center.\(^{18}\) Hydrolysis of the methyl ester and cleavage of the benzyl ethers gave the corresponding β-configured mimetic 12 in 65% yield (Scheme 2).

![Scheme 2](image)

Scheme 2. Synthesis of β-C-mannoside 12 and α-C-mannoside 4. Reagents and conditions: a) All-OH, AcCl; b) NaH, BnBr, DMF, 0 °C; c) (Ph\(_3\)P)\(_3\)RhCl, toluene/EtOH/H\(_2\)O, reflux; d) I\(_2\), THF/H\(_2\)O; e) Cl\(_3\)CCN, DBU, DCM; f) 7, TMSOTf, DCM, MS 4 Å, 0 °C; g) 1,4-dioxane/MeOH/4N NaOH; h) H\(_2\) (10 bar) Pd(OH)\(_2\)/C; i) 7, ZnCl\(_2\), DCM, MS 4 Å, rt.

It has been demonstrated with other phenolic acceptors that α-C-mannosides can be obtained if ZnCl\(_2\) is used as the promoter.\(^{18}\) However, these conditions only gave a 2:1 mixture of the α-C-glycoside 13C and the α-O-glycoside 13O. The products could be separated after ester hydrolysis giving the acid 14C in 25% over two steps. Unfortunately, the mannosyl residue in 14C was found to adopt the \(^1\)C\(_4\) conformation\(^{18}\) which also prevailed after hydrogenolysis of the benzyl ethers to yield the α-configured mimetic 4, albeit in an unfavorable conformation (Scheme 2).
Scheme 3. Synthesis of α-O-mannoside 17. Reagents and conditions: a) Ac₂O, pyridine; b) H₂NCH₂CH₂NH₂, THF; c) Cl₃CCN, DBU, DCM d) TMSOTf, DCM, MS 4Å; e) 1,4-dioxane /MeOH/4N NaOH.

For comparison of its biological activity with the two C-mannosides, glycoside 17 was synthesized from the acetylated mannosyl trichloroacetimidate 15 which has a strong preference for α-O-glycosylation. Donor 15 was obtained from D-mannose in three steps and reacted with phenol 7 in the presence of TMSOTf to give the α-configured O-glycoside 16 in 70% yield. After removal of the acetyl groups and alkaline hydrolysis of the methyl ester, the O-glycosidic mimetic 17 was obtained in 61% yield (Scheme 3). As expected, the mannose in this compound adopts the 4C₁ conformation while the distance between carboxylate and the carbohydrate is larger than in the C-glycosides 4 and 12.

The three mimetics 4, 12 and 17 were evaluated for their selectin inhibition in a cell-based assay, based on binding of soluble P- or E-selectin-Immunoglobulin chimera to selectin ligand-bearing murine Th1 cells or neutrophils. However, no inhibition exceeding 50% could be observed up to concentrations of 1 mM.

Assuming that the mannosyl residue in 4 adopts a 1C₄ conformation in solution we performed docking calculations in order to test whether 4 could fit favorably into the binding pocket of E-selectin in this conformation. None of the 100 best poses showed a twofold coordination of the calcium and the binding energy score was lower than for 4 with the mannosyl residue in a 4C₁ conformation. This result might explain why the compound did not show the expected inhibition in the biological test. DFT calculations (B3LYP/SOLV, 6-31G**) predict the 4C₁ conformation of a simplified model of 4 to be 1.7 kJ·mol⁻¹ more stable in the gas phase while the 1C₄ conformer is more stable in solution by 11.5 kJ·mol⁻¹, see supporting information.

Conclusions

Three potential monosaccharidic mimetics of the sialyl LewisX tetrasaccharide based on 2,7-dihydroxynaphthalene have been prepared and evaluated for their selectin-inhibitory activity. While mimetic 4 compared favorably with other tested mimetics in an in silico screening, none of the compounds showed a significant biological activity in a cytometric assay up to concentrations of 1 mM. The reason for this may be the unexpected preference of the mannosyl...
residue in 4 for the $^1$C$_4$ conformation, in which the spatial arrangement of the hydroxyl groups in positions 2, 3, and 4 precludes the required binding of the Ca$^{2+}$-ion in E-selectin.

**Experimental Section**

**General.** Moisture sensitive reactions were carried out under argon atmosphere in dried glassware sealed by rubber septa. Unless otherwise specified, chemicals were obtained from commercial suppliers and were used without further purification. CH$_2$Cl$_2$ and acetonitrile were dried over CaH$_2$ and distilled under argon atmosphere prior to use. ZnCl$_2$ was dried at 130 °C in vacuo. Flash chromatography was performed on silica gel 60 (0.035–0.070 mm, Acros). Chromatography solvents (cyclohexane, EtOAc) were distilled prior to use. For analytical TLC, Merck silica gel aluminium sheets (60 F$_{254}$) were used. Visualisation was accomplished by UV (254 nm) and sugar reagent$^{20}$ (1 M ethanolic H$_2$SO$_4$/0.2% ethanolic 3-methoxyphenol solution 1:1). Purification of products was accomplished by flash chromatography on silica gel and the purified compounds showed a single spot in analytical TLC. $^1$H and $^{13}$C NMR spectra were recorded on a Bruker AC 300, AV 400 or DRX 500 in CDCl$_3$ or methanol-$d_4$ using the residual solvent peak as internal reference (CDCl$_3$, $\delta$$_H$ = 7.26, $\delta$$_C$ = 77.16, methanol-$d_4$, $\delta$$_H$ = 3.31, $\delta$$_C$ = 49.0). Optical rotations were measured at room temperature on a Krüss P8000 polarimeter at 589 nm or on a Perkin Elmer 241 polarimeter at 546 and 578 nm; the optical rotation at 589 nm was extrapolated using the Drude equation. IR spectra were recorded on a ThermoNicolet Avatar 370 FT-IR spectrometer. FAB mass spectrometry was carried out on with VG70S (Xe-FAB ionisation) with m-nitrobenzyl alcohol as the matrix. For exact mass determination (FAB-HRMS), PEG 300 or PEG 600 was used as internal standard. ESI mass spectrometry was carried out on an Agilent 1200 LC/MSD Trap XCT. The samples were dissolved in acetonitrile (c ≈ 0.1 g/l) and injected via an Agilent 1200 HPLC with an Ascentis Express C8 (30 x 2.1 mm, 2.7 µm particle size) column (acetonitrile/water 80:20, Flow: 0.5 ml/min). Exact mass determination (ESI-HRMS) was carried out on a Q-ToF-Ultima 3-Instrument with a Lock Spray-interface. NaI/CsI clusters were used as an external reference.

**Docking**

The program Glide 4.5$^{21}$ was used for the docking of a small, manually designed, library of potential sLe$^X$ mimetics. The setup of the receptor was performed using the protein preparation wizard of the Maestro program$^{22}$ based on the X-ray structure of E-selectin complexed with sLe$^X$ (pdb code 1G1T).$^{23}$ The ligands were built and optimized using Maestro. The grid defining the binding site was positioned using the sLe$^X$ ligand as center and had a dimension of 31 Å x 29 Å x 29 Å. The docking was performed in GlideScore SP4.5 mode using default parameters. The 10 best poses were stored for each ligand and manually inspected.
Cellular assay for determination of inhibitory potency for E- and P-selectin
Inhibition of binding of E- and P-selectin to its natural ligands on either T cells or neutrophils was tested by cytometry as described. In short, CD4+ T cells were isolated from mice, activated under Th1 conditions and incubated with soluble E- or P-selectin-IgG chimera in absence or presence of the compounds. Cell-bound selectin was stained with fluorescently labeled anti-human IgG antibody as secondary reagent in HBSS containing Ca2+ and Mg2+ and quantified in a fluorescence-activated cell sorter (FACS). Alternatively, neutrophils were stained in whole blood after erythrocyte lysis as above and identified by anti GR-1 antibody.

(S)-3-Cyclohexyl-2-(7-hydroxynaphthalen-2-yloxy)-propionic acid methyl ester (7)
A solution of 6 (1.68 g, 5.28 mmol) in dry acetonitrile (8 mL) was added to a mixture of 2,7-dihydroxynaphthalene (4.20 g, 26.2 mmol) and K2CO3 (1.80 g) in dry acetonitrile (17 mL) under argon atmosphere. The mixture was stirred for 1.5 h at room temperature and partitioned between CH2Cl2 (70 mL) and H2O (40 mL). The organic layer was washed twice with H2O (10 mL each) and once with saturated aq NaHCO3 (20 mL). Drying over Na2SO4 and removal of the solvent in vacuo furnished a crude residue which was purified by flash chromatography (cyclohexane/EtOAc 5:1) to give 7 as a colorless solid (1.49 g, 4.54 mmol, 86%). Mp 88–90 °C, [α]D25 = −13.6 (c = 1, CDCl3), Rf = 0.23 (cyclohexane/EtOAc 5:1), IR (NaCl, νmax, cm−1): 3416, 2923, 2850, 1736, 1515, 1447, 1203, 1159, 831. 1H NMR (400 MHz, CDCl3): δH 7.67 (d, JH-H-4 = 8.9 Hz, 1 H, H-4), 7.65 (d, JH-H-5 = 8.7 Hz, 1 H, H-5), 7.03 (dd, JH-H-3, H-4 = 8.9 Hz, JH-H-1, H-3 = 2.6 Hz, 1 H, H-3), 7.01 (d, JH-H-6 = 2.5 Hz, 1 H, H-6), 6.95 (dd, JH-H-5, H-6 = 8.7 Hz, JH-H-6, H-8 = 2.5 Hz, 1 H, H-6), 6.89 (d, JH-H-1, H-3 = 2.6 Hz, 1 H, H-1), 4.83 (dd, JH-H-1 = 9.4 Hz, JH-H-1 = 4.1 Hz, 1 H, CHCOOMe), 3.75 (s, 3 H, OCH3), 2.02–1.93 (m, 1 H, CH2CHCOOMe), 1.83–1.58 (m, 7 H, CH2CHCOOMe, CH5, CH2), 1.31–1.13 (m, 3 H, CH2), 1.08–0.91 (m, 2 H, CH2). 13C NMR (100.6 MHz CDCl3): δC 173.5 (COOH), 156.9, 154.5 (C-2, C-7), 136.1 (C-8a), 129.9, 129.8 (C-4, C-5), 125.1 (C-4a) 116.6, 116.1, 109.2, 106.7 (C-1, C-3, C-6, C-8), 75.0 (CHCOOMe), 52.7 (OCH3), 20.7 (CH2CHCOOMe), 34.3 (CH), 34.1, 32.8, 26.7, 26.5, 26.3 (CH2). ESI-MS: m/z = 351.1 (100) [M+Na]+, 329.1 (20) [M+H]+; ESI-HRMS: m/z calcd for [C20H24O4+Na]+: 351.1567, found 351.1574.

(2S)-3-Cyclohexyl-2-[2-hydroxy-1-(2,3,4,6-tetra-O-benzyl-β-D-mannopyranosyl)naphthalen-7-yloxy]propionic acid methyl ester (10)
A mixture of 9 (577 mg, 0.84 mmol), 7 (291 mg 0.89 mmol) and activated molecular sieves (4 Å, 2.00 g) in dry CH2Cl2 (10 mL) was stirred at 0 °C for 20 min under argon atmosphere to remove traces of water from the reactants. Then, TMSOTf (155 µL, 190 mg, 0.86 mmol) in dry CH2Cl2 (2 mL) was added and the mixture was stirred for 2.5 h. The reaction was quenched by addition of saturated aq NaHCO3 (20 mL). The organic layer was separated and the aq phase extracted with CH2Cl2 (3 x 20 mL). The combined organic extracts were dried over Na2SO4 and concentrated in vacuo. The crude residue was purified by flash chromatography (cyclohexane/EtOAc, 15:1) to give 10 (435 mg, 0.51 mmol, 61%) as colorless oil. [α]D25 = +62.3
(c = 1, CHCl₃), Rf = 0.35 (cyclohexane/EtOAc 10:1). ¹H NMR, COSY, NOESY (500 MHz, CDCl₃): δH 9.15 (s, 1 H, OH), 7.68 (d, J₃-H₅-H₆ = 8.9 Hz, 1 H, H-5), 7.63 (d, J = 8.9 Hz, 1 H, H-4), 7.41–7.24 (m, 13 H, H–Ph), 7.19 (dd, J₂ = 7.3 Hz, J₁ = 1.9 Hz, 2 H, H-Ph), 7.08–6.95 (m, 5 H, H-3, H-6, 3 H–Ph), 6.91 (d, J = 7.1 Hz, 2 H, H–Ph), 6.81 (d, J₄-H₆-H₈ = 1.8 Hz, 1 H, H-8), 5.32 (pseudo s, 1 H, H-1'), 4.92 (d, J = 10.8 Hz, 1 H, (C-4')–O–CH₂Ph), 4.82 (d, J = 11.8 Hz, 1 H, (C-3')–O–CH₂Ph). 4.80–4.74 (m, 2 H, (C-3')–O–CH₂Ph, CHCOOME), 4.67 (d, J = 12.2 Hz, 1 H, (C-6')–O–CH₂Ph), 4.57 (d, J = 10.9 Hz, 1 H, (C-4')–O–CH₂Ph), 4.54 (d, J = 12.2 Hz, 1 H, (C-6')–O–CH₂Ph), 4.43 (d, J = 11.8 Hz, 1 H, (C-2')–O–CH₂Ph), 4.28 (m, 2 H, H-4', (C-2')–O–CH₂Ph), 4.13 (d, J = 2.6 Hz, 1 H, H-2'), 3.98 (dd, J = 2.3 Hz, 1 H, H-3'), 3.83 (dd, J = 10.7 Hz, J₃-H₅-H₆a = 3.5 Hz, 1 H, H-6a'), 3.80 (dd, J = 10.7 Hz, J₃-H₅-H₆b = 2.1 Hz, 1 H, H-6b'), 3.75–3.68 (m, 1 H, H-5'), 3.56 (s, 3 H, OCH₃), 2.01–1.94 (m, 1 H, CH₂CHCOOH), 1.85–1.57 (m, 7 H, 1 CH₂CHCOOH, CH, 5 CH₂), 1.33–1.12 (m, 3 H, CH₂), 1.09–0.93 (m, 2 H, CH₂). ¹³C NMR, HSQC, HMBC (100.6 MHz, CDCl₃) δC 173.6 (COOH), 157.0 (C-7), 156.9 (C-2), 139.0, 138.8, 138.5, 138.2 (C-1'), 132.4 (C-8a), 131.1 (C-5), 130.0 (C-4), 128.8, 128.7, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.6, 127.5 (20 CH Ph), 124.5 (C-4a), 118.5 (C-3), 114.6 (C-6), 111.3 (C-1), 102.1 (C-8), 84.6 (C-3'), 80.1 (C-5'), 79.5 (C-1'); 75.9, 75.8, 75.6 (CHCOOME, C-2', (C-4')–O–CH₂Ph), 74.9 ((C-2')–O–CH₂Ph), 74.6 (C-4'), 73.8 ((C-6')–O–CH₂Ph); 72.6 ((C-3')–O–CH₂Ph) 68.8 (C-6'); 52.6 (COOCH₃), 40.5 (CH₂CHCOOMe), 34.2 (CH), 34.0, 32.9, 26.7, 26.5, 26.4 (CH₂). ESI-MS: m/z (%) = 873.5 (100 [M+Na]⁺, ESI-HRMS: m/z calcd for [C₃₅H₅₈O₉+Na]⁺: 873.3973, found: 873.3975.

(2S)-3-Cyclohexyl-2-[2-hydroxy-1-(2,3,4,6-tetra-O-benzyl-β-D-mannopyranosyl)naphthalen-7-yl]propionic acid (11)
Methyl ester 10 (327 mg, 0.39 mmol) was stirred in a mixture of 1,4-dioxane (10 mL), methanol (3.5 mL) and 4 N NaOH (1 mL) for 3 h at room temperature. The reaction was quenched by adding 2 N HCl (10 mL) and H₂O (30 mL) and the mixture was extracted with EtOAc (2 x 20 mL). The combined organic phases were dried over Na₂SO₄ and the solvent was removed in vacuo. The crude residue was purified by flash chromatography (cyclohexane/EtOAc 3:1) to give 11 (272 mg, 0.32 mmol, 83%) as colorless oil. [α]D²⁶ = +75.9 (c = 1, CHCl₃), Rf = 0.18 (cyclohexane/EtOAc 3:1), IR (NaCl, νmax, cm⁻¹): 3340, 3031, 2922, 1724, 1624, 1454, 1212, 1098, 1027, 736. ¹H NMR, COSY (400 MHz, CDCl₃): δH 9.13 (s, 1 H, OH), 7.66 (d, J₃-H₅-H₆ = 8.9 Hz, 1 H, H-5), 7.61 (d, J₃-H₃-H₄ = 8.9 Hz, 1 H, H-4), 7.42–7.24 (m, 13 H, H–Ph), 7.21 (dd, J = 7.3 Hz, J = 2.1 Hz, 2 H, H-Ph), 7.07–6.99 (m, 4 H, H-3, 3 H–Ph), 6.96 (dd, J₃-H₅-H₆ = 8.9 Hz, J₄-H₆-H₈ = 2.3 Hz, 1 H, H-6), 6.92 (m, 2 H, H-Ph), 6.71 (d, J₄-H₆-H₈ = 2.3 Hz, 1 H, H-8), 5.26 (pseudo s, 1 H, H-1'), 4.94 (d, J = 10.9 Hz, 1 H, CH₂Ph), 4.75 (s, 2 H, CH₂Ph), 4.70 (dd, J = 9.2 Hz, J = 4.4 Hz, 1 H, CHCOOH), 4.67 (d, J = 12.2 Hz, 1 H, CH₂Ph), 4.58 (d, J = 10.9 Hz, 1 H, CH₂Ph), 4.54 (d, J = 12.2 Hz, 1 H, CH₂Ph), 4.40 (d, J = 11.7 Hz, 1 H, CH₂Ph), 4.28 (d, J = 11.7 Hz, 1 H, CH₂Ph), 4.27 (pseudo t, Japp = 9.4 Hz, 1 H, H-4') 4.10 (d, J₃-H₂-H₃ = 2.8 Hz, 1 H, H-2'), 3.88 (dd, J₃-H₃-H₄ = 9.4 Hz, J₃-H₂-H₃ = 2.8 Hz, 1 H, H-3'), 3.83 (dd, J = 10.7 Hz, J₃-H₅-H₆a = 3.5 Hz, 1 H, H-6a'), 3.79 (dd, J = 10.7 Hz, J₃-H₅-H₆b = 2.2 Hz, 1 H, H-6b').
(2S)-3-Cyclohexyl-2-[2-hydroxy-1-(β-D-mannopyranosyl)naphthalen-7-yloxy]propionic acid (12)

A mixture of 11 (159 mg, 0.19 mmol) and Pd(OH)$_2$ on charcoal (15 mg, 20 wt% Pd) and methanol (5 mL) was degassed in an autoclave under N$_2$ and flushed with H$_2$ (10 bar). The mixture was stirred for 2 days under H$_2$ (10 bar) at room temperature. The catalyst was removed by filtering through Celite and the solvent was removed in vacuo. The crude residue was purified by flash chromatography (EtOAc/EtOH 9:1) to give 12 as colorless amorphous solid (70 mg, 0.15 mmol, 78%). $\delta^2$S = +45.8 (c = 1, MeOH), $R_f$ = 0.10 (EtOAc/EtOH 9:1). IR (KBr, $\nu_{\text{max}}$, cm$^{-1}$): 3387, 2923, 1727, 1625, 1522, 1451, 1227, 1074, 834, 782. $^1$H NMR, COSY, NOESY (400 MHz, CD$_3$OD): $\delta$H 7.67 (d, $^3$J$_{H,5,H-6}$ = 8.9 Hz, 1 H, H-5), 7.62 (d, $^3$J$_{H,3,H-4}$ = 8.9 Hz, 1 H, H-4), 7.15 (pseudo s, 1 H, H-8), 6.99 (dd, $^3$J$_{H,5,H-6}$ = 8.9 Hz, $^4$J$_{H,6,H-8}$ = 2.3 Hz, 1 H, H-6), 6.92 (d, $^3$J$_{H,3,H-4}$ = 8.8 Hz, 1 H, H-3), 5.50 (pseudo s, 1 H, H-1$''$), 4.83 (dd, $^3$J = 9.2 Hz, $^3$J = 3.9 Hz, 1 H, CHCOOH), 4.12 (d, $^3$J$_{H-2',H-3'}$ = 2.0 Hz, 1 H, H-2$''$), 3.98 (dd, $^3$J = 12.1 Hz, $^3$J$_{H-5',H-6a''}$ = 2.2 Hz, 1 H, H-6a$'$), 3.88 (m, 3 H, H-3$'$, H-4$'$, H-6b$'$), 3.53 (m, 1 H, H-5$'$), 2.00–1.66 (m, 8 H, CH$_2$CHCOOH, CH, 5 CH$_2$), 1.39–0.97 (m, 5 H, CH$_2$). $^{13}$C NMR, HSQC, HMBC (101 MHz, CD$_3$OD): $\delta$C 178.8 (COOH), 158.9 (C-7), 157.8 (C-2), 135.1 (C-8a), 132.1 (C-5), 131.2 (C-4), 126.6 (C-4a), 118.9 (C-3), 117.1 (C-6), 114.4 (C-1), 104.9 (C-8), 84.1 (C-5$'$), 81.3 (C-1$'$), 77.5 (CHCOOH), 77.0 (C-3$'$), 74.1 (C-2$'$), 69.2 (C-4$'$), 63.6 (C-6$'$), 42.4 (CH$_2$CHCOOH), 36.7, 35.9, 34.6, 28.4, 28.3, 28.1 (CH, 5 CH$_2$). FAB-MS: m/z (%) = 515.2 (100) [M+K]$^+$, 499.2 (59) [M+Na]$^+$, 476.2 (31) [M]$^+$, FAB-HRMS: m/z calcd for [C$_{25}$H$_{32}$O$_9$]+: 476.2046, found: 476.2061; calcd for m/z [C$_{25}$H$_{32}$O$_9$+H]: 477.2119, found: 477.2115.

(2S)-3-Cyclohexyl-2-[2-hydroxy-1-(2,3,4,6-tetra-O-benzyl-α-D-mannopyranosyl)naphthalen-7-yloxy]propionic acid methyl ester (13C) and (2S)-3-Cyclohexyl-2-[2-(2,3,4,6-tetra-O-benzyl-α-D-mannopyranosyl)oxy]naphthalen-7-yloxy]propionic acid methyl ester (13O)

A mixture of 9 (320 mg, 0.50 mmol), 7 (174 mg, 0.53 mmol) and molecular sieves 4 Å (1.00 g) in dry CH$_2$Cl$_2$ (10 mL) was stirred at room temperature for 20 min under argon atmosphere to remove traces of water from the reactants. Dried ZnCl$_2$ (206 mg, 1.52 mmol) was added and the
mixture was stirred for 2 h at room temperature. The reaction was quenched by addition of saturated aq NaHCO₃ (20 mL) and extracted with CH₂Cl₂ (3 x 10 mL). The combined organic phases were dried over Na₂SO₄ and the solvent was removed in vacuo. The crude residue was purified by flash chromatography (cyclohexane/EtOAc 12:1) to give a 2:1 mixture (¹H NMR) of 13C (A) and 13O (B) (222 mg total, 0.26 mmol, combined yield 52%) as colorless oil. Rf (both compounds) = 0.20 (cyclohexane/EtOAc 12:1). ¹H NMR, COSY (400 MHz, CDCl₃) δ_H 8.83 (s, 1 H₆, OH₆), 7.69–7.63 (m, 2 H₈, H-4₈, H-5₈, 2 H₉, H-4₉, H-5₉), 7.47–6.91 (m, 21 H, H-3H₆⁻⁶, H-8H₆⁻⁸, 18 H-P₄, 24 H-B, H-1B, H-3B, H-6B, H-8B, 20 H-P₈B), 6.57 (d, 3 J = 7.4 Hz, 2 H₄, 2 H-P₄), 5.82 (d, 3 J_{H1-H2} = 10.1 Hz, 1 H₁A, H-1A), 5.76 (d, 3 J_{H1-H2} = 1.9 Hz, 1 H₁B, H-1B), 4.91 (d, 2 J = 10.7 Hz, 1 H₁B, CH₂Ph), 4.87–4.78 (m, 2 H₁, CHCOOME₁A, CH₂Ph₁, 1 H₈, CHCOOME₈B), 4.74 (d, 2 J = 11.7 Hz, 1 H₇B, CH₂Ph₇B), 4.70 (d, 2 J = 11.7 Hz, 1 H₇B, CH₂Ph₇B), 4.69–4.62 (m, 1 H₁A, CH₂Ph₁, 3 H₈B, CH₂Ph₈B), 4.60 (d, 2 J = 12.2 Hz, 1 H₁A, CH₂Ph₁), 4.56 (d, 2 J = 12.0 Hz, 1 H₁A, CH₂Ph₁), 4.53 (d, 2 J = 9.5 Hz, 1 H₁B, CH₂Ph₁B), 4.50–4.42 (m, 2 H₁A, CH₂Ph₁A, 1 H₈B, CH₂Ph₈B), 4.41 (pseudo t, 3 J = 6.7 Hz, 1 H₁A, H-5₅₈A), 4.23 (dd, 3 J_{H1-H2'} = 10.1 Hz, 3 J_{H2-H1'} = 2.8 Hz, 1 H₄A, H-2₄A), 4.20–4.13 (m, 2 H₈B, H-3B, H-4B), 4.07 (dd, 2 J = 10.3 Hz, 3 J_{H5-H6a} = 7.2 Hz, 1 H₆₄A, H-6₄A), 4.03–4.00 (m, 1 H₁B, H-2B), 3.98 (pseudo t, J_{app} = 3.0 Hz, 1 H₁A, H-3₅₈A), 3.95–3.86 (m, 2 H₈B, CH₂Ph₈B), 3.80 (m, 1 H₁A, H-4₅₄₈A, 1 H₈B, H-6₄B), 3.73 (s, 3 H₅₄B, OCH₃B), 3.69–3.62 (m, 1 H₆₈A, CH₂Ph₆₈A, 1 H₈B, H-6₆B), 3.54 (s, 3 H₅₈B, OCH₃₈B), 1.97 (ddd, 3 J = 14.5 Hz, 3 J = 9.4 Hz, 3 J = 5.4 Hz, 1 H₈B, CH₂CHCOOME₈B), 1.93–1.84 (m, 1 H₁A, CH₂CHCOOME₈B), 1.83–1.55 (m, 7 H₈B, CH₂CHCOOME₈B, CH₂A, CH₂₈B, 7 H₈, CH₂CHCOOME₈B, CH₈B, 5 CH₂B), 1.33–0.79 (m, 5 H₈A, CH₂₅A, 5 H₈B, CH₂₅B).

(2S)-3-Cyclohexyl-2-[2-hydroxy-1-(2,3,4,6-tetra-O-benzyl-α-D-mannopyranosyl)naphthalen-7-yloxy]propionic acid (14C) and (2S)-3-cyclohexyl-2-[2-(2,3,4,6-tetra-O-benzyl-α-D-mannopyranosyl)naphthalen-7-yloxy]propionic acid (14O)

The mixture (2:1) of 13C and 13O (150 mg total, 0.18 mmol) was dissolved in a mixture of 1,4-dioxane (5 mL), methanol (1.8 mL) and 4 N aq NaOH (0.5 mL) and stirred for 3 h at room temperature. The reaction was quenched with 2 N HCl (10 mL) and H₂O (30 mL). The mixture was extracted with EtOAc (3 x 20 mL). The combined organic phases were dried over Na₂SO₄ and the solvent was removed in vacuo. The crude residue was purified by flash chromatography (cyclohexane/EtOAc 3:1) to give 14C and 14O (combined yield 138 mg, 0.16 mmol, 93%). The separation yielded a mixture of 14C and 14O (29 mg, 34 μmol, 19%) along with the pure components:

14C (72 mg, 83 μmol, 49%), colorless oil. [α]²⁸_D = -57.6 (c = 1, CHCl₃), Rf = 0.16 (cyclohexane/EtOAc 3:1), IR (NaCl, v_max cm⁻¹): 3354, 3029, 2923, 1726, 1623, 1453, 1211, 1093, 750, 698. ¹H NMR, COSY, NOESY (500 MHz, CDCl₃): δ_H 8.86 (s, 1 H, OH), 7.68 (pseudo d, 3 J = 8.9 Hz, 2 H, H-4, H-5), 7.41–7.31 (4 H, H₈, 3 H-P₄, H₈-P₄), 7.30–7.20 (m, 12 H, H-P₈, 7.13 (t, 3 J = 7.2 Hz, 1 H, H-P₈), 7.07 (m, 3 H, H₃, 2 H-P₈), 7.02 (dd, 3 J_{H₅-H₆} = 8.9 Hz, 4 J_{H₅-H₆} = 2.0 Hz, 1 H, H₆), 6.59 (d, 3 J = 7.2 Hz, 2 H, H-P₈), 5.85 (d, 3 J_{H₁-H₂} = 10.1 Hz, 1 H, H₁), 4.79 (d, 2 J = 12.4 Hz, 1 H, CH₂P₄), 4.74 (dd, 3 J = 8.5 Hz, 3 J = 4.1 Hz, 1 H, CHCOOH),
4.65–4.56 (m, 2 H, CH₂Ph), 4.54–4.43 (m, 3 H, CH₃Ph), 4.39 (pseudo t, J_app = 6.6 Hz, 1 H, H-5′), 4.23 (dd, 3J_H-1′,H-2′ = 10.1 Hz, 3J_H-2′,H-3′ = 2.4 Hz, 1 H, H-2′), 4.08–4.01 (m, 1 H, H-6a′), 3.95 (pseudo s, 1 H, H-3′), 3.92–3.82 (m, 2 H, H-6b′, CH₂Ph), 3.78 (pseudo d, J_app = 3.6 Hz, 1 H, H-4′), 3.66 (d, 2J = 11.1 Hz, 1 H, CH₂Ph), 1.95–1.87 (m, 1 H, CH₂CHCOOH), 1.71 (m, 2 H, CH₂CHCOOH, CH₂^3Hex), 1.62 (m, 5 H, CH, 4 CH₂), 1.34–1.05 (m, 3 H, CH₃), 0.92 (m, 1 H, CH₂), 0.78 (m, 1 H, CH₂). ^13C NMR, DEPT, HSQC, HMBC (126 MHz, CDCl₃): δC 176.2 (COOH), 156.1, 156.0 (C-2, C-7), 138.7, 138.4, 138.0 (4 C-1′′′′), 134.6 (C-8a), 130.4, 130.1 (C-4, C-5), 128.9, 128.7, 128.2, 128.1, 128.0, 127.9, 127.8, 127.6 (20 CH Ph), 124.9 (C-4a), 118.0 (C-3), 115.2 (C-6), 114.4 (C-1′), 105.2 (C-8), 76.3 (C-5′), 75.8 (C-2′), 75.1 (C-4′), 74.6 (CHCOOH), 74.2 (C-3′), 73.6, 73.3, 73.1, 71.9 (CH₂Ph), 68.3 (C-1′), 67.3 (C-6′), 40.1 (CH₂CHCOOH), 33.9 (CH, CH₂), 32.5, 26.6, 26.4, 26.3 (CH₂). FAB-MS: m/z = 836.5 (15) [M]+, 181.1 (100) [matrix] FAB-HRMS: m/z calc for [C₅₃H₅₀O₇+H]^+: 837.3997, found. 837.3975.

140 (37 mg, 43 μmol, 25%), colorless oil. [α]²⁰_D = +68.5 (c = 1, CHCl₃). R₉ = 0.10 (cyclohexane/EtOAc 3:1), IR (NaCl, v_max, cm⁻¹): 3441, 3029, 2922, 1733, 1633, 1453, 1208, 1095, 1027, 749, 697. ^1H NMR, COSY, NOESY (400 MHz, CDCl₃): δH 7.70 (d, 3J_H-5,H-6 = 8.9 Hz, 1 H, H-5), 7.67 (d, 3J = 8.9 Hz, 1 H, H-4), 7.48–7.17 (m, 21 H, H-1, H-Ph), 7.10 (dd, 3J_H-5,H-6 = 8.9 Hz, 4J_H-6,H-8 = 2.4 Hz, 1 H, H-6), 7.08–7.01 (m, 2 H, H-8, H-3), 5.78 (d, 3J_H-1′,H-2′ = 1.5 Hz, 1 H, H-1′), 4.94 (d, 2J = 10.7 Hz, 1 H, CH₂Ph), 4.88 (dd, 3J = 9.4 Hz, 3J = 3.9 Hz, 1 H, CHCOOH), 4.84 (m, 2 H, CH₂Ph), 4.77 (d, 2J = 11.8 Hz, 1 H, CH₂Ph), 4.73 (d, 2J = 11.8 Hz, 1 H, CH₂Ph), 4.65 (d, 2J = 12.0 Hz, 1 H, CH₂Ph), 4.56 (d, 2J = 10.7 Hz, 1 H, CH₂Ph), 4.42 (d, 2J = 12.0 Hz, 1 H, CH₂Ph), 4.26–4.16 (m, 2 H, H-4′, H-3′), 4.04 (m, 1 H, H-2′), 3.92 (m, 1 H, H-5′), 3.82 (dd, 2J = 10.8 Hz, 3J_H-5,H-6b′ = 4.1 Hz, 1 H, H-6a′), 3.69 (dd, 2J = 10.8 Hz, 3J_H-5,H-6b′ = 1.3 Hz, 1 H, H-6b′), 2.10–1.97 (m, 1 H, CH₂CHCOOH), 1.91–1.62 (m, 7 H, CH₂CHCH₂COOH, CH₅, CH₂), 1.38–0.92 (m, 5 H, CH₂). ^13C NMR (100.6 MHz, CDCl₃): δC 176.6 (COOH), 156.6 (C-7), 155.0 (C-2), 138.8, 138.7, 138.5 (4 C-1′′′′), 135.8 (C-8a), 129.8 (C-5), 129.5 (C-4), 128.8, 128.7, 128.6, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8 (20 CH Ph), 125.9 (C-4a), 117.1 (C-6, C-3), 110.4 (C-1), 107.9 (C-8), 96.6 (C-1′), 80.3 (C-3′), 75.5 (CH₂Ph), 75.0 (C-4′), 74.9 (C-2′), 74.6 (CHCOOH), 73.5, 73.1 (CH₂Ph), 72.8 (C-5′), 72.7 (CH₂Ph), 69.1 (C-6′), 40.4 (CH₂CHCOOH), 34.3 (CH), 34.0, 32.7, 26.7, 26.5, 26.3 (CH₂). ESI-MS: m/z (%) = 854.6 (100) [M+NH₄]^+. ESI-HRMS: m/z calc for [C₅₃H₅₀O₇+Na]^+: 859.3817, found: 859.3797.

(2S)-3-Cyclohexyl-2-[2-hydroxy-1-(α-d-mannopyranosyl)naphthalen-7-yl oxy]propionic acid (4)

A mixture of 14C (52 mg, 62 μmol) and Pd(OH)₂ on charcoal (4 mg, 20 wt% Pd) in MeOH (2 mL) was degassed in an autoclave under N₂ and then flushed with H₂ (10 bar). The mixture was stirred under H₂ (10 bar) at room temperature. The catalyst was removed by filtration through Celite and the solvent was removed in vacuo. The crude residue was purified by flash chromatography (EtOAc/EtOH 9:1) to give 4 (18 mg, 38 μmol, 61%) as a colorless oil. [α]²⁰_D = +16.9 (c = 0.16, MeOH), R₉ = 0.11 (EtOAc/EtOH 9:1). IR (ATR, v_max, cm⁻¹): 3339, 2921, 2850, 1726, 1624, 1522, 1450, 1226, 1137, 1073, 833, 783. ^1H NMR (400 MHz, CD₃OD): δH 7.65 (m, n,
$J_{\text{app}} = 9.5 \text{ Hz}, 3 \text{ H, H-4, H-5, H-8}$), 6.99 (dd, $^3J_{\text{H-5,H-6}} = 8.9 \text{ Hz}$, $^4J_{\text{H-6,H-8}} = 2.3 \text{ Hz}$, 1 H, H-6), 6.96 (d, $^3J_{\text{H-3,H-4}} = 8.8 \text{ Hz}$, 1 H, H-3), 5.73 (d, $^3J_{\text{H-1',H-2'}} = 9.2 \text{ Hz}$, 1 H, H-1'), 4.91 (m, 1 H, CHCOOH), 4.55 (dd, $^3J_{\text{H-1',H-2'}} = 9.2 \text{ Hz}$, $^3J_{\text{H-2',H-3'}} = 3.4 \text{ Hz}$, 1 H, H-2'), 4.33 (dd, $^2J = 12.3 \text{ Hz}$, $^3J_{\text{H-5',H-6a'}} = 7.7 \text{ Hz}$, 1 H, H-6a'), 4.16 (pseudo t, $J_{\text{app}} = 4.0 \text{ Hz}$, 1 H, H-3'), 3.98 (dd, $^3J_{\text{H-3',H-4'}} = 4.6 \text{ Hz}$, $^3J_{\text{H-4',H-5'}} = 2.4 \text{ Hz}$, 1 H, H-4'), 3.95 (m, 1 H, H-5'), 3.78 (dd, $^2J = 12.3 \text{ Hz}$, $^3J_{\text{H-5',H-6b'}} = 3.7 \text{ Hz}$, 1 H, H-6b'), 1.99–1.66 (m, 8 H, 2 CH$_2$CHCOOH, CH, 5 CH$_2$), 1.38–1.23 (m, 5 H, CH$_2$).

$^{13}$C NMR, HSQC (100.6 MHz, CD$_2$OD) $\delta_C$ 178.9 (COOH), 158.4 (C-7), 156.8 (C-2), 136.9 (C-8a), 131.4, 131.3 (C-4, C-5), 126.8 (C-4a), 118.0 (C-3), 117.1 (C-1), 116.9 (C-6), 107.3 (C-8), 82.0 (C-5'), 77.3 (CHCOOH), 72.9 (C-3') 71.3 (C-4'), 70.6 (C-1'), 69.2 (C-2'), 61.5 (C-6'), 42.2 (CH$_2$CHCOOH), 36.0, 35.5, 34.4, 31.2, 27.9, 27.8 (CH, 5 CH$_2$). FAB-MS: $m/z$ (%) = 515.1 (44) [M+K]$^+$, 499.1 (86) [M+Na]$^+$, 476.1 (17) [M]$^+$, 176 (100) [matrix], FAB-HRMS: $m/z$ calcd for [C$_{25}$H$_{32}$O$_9$]$^+$: 476.2046, found: 476.2041; $m/z$ calcd for [C$_{25}$H$_{32}$O$_9$+H]$^+$: 477.2125, found: 477.2116.

(2S)-3-Cyclohexyl-2-[2-(2,3,4,6-tetra-O-acetyl-α-d-mannopyranosyloxy)naphthalen-7-yloxy]-propionic acid methyl ester (16)

A mixture of 7 (192 mg, 0.58 mmol), 15 (270 mg, 0.55 mmol) and molecular sieves 4 Å (1.00 g) in dry CH$_2$Cl$_2$ (10 mL) is stirred at 0 °C for 20 min under argon atmosphere to remove traces of water from the reactants. TMSOTf (20 μL, 24.6 mg, 0.11 mmol) in CH$_2$Cl$_2$ (5 mL) was added and the mixture was stirred for 2 h at 0 °C. The reaction was quenched by addition of saturated aq NaHCO$_3$ (30 mL) and the mixture was extracted with CH$_2$Cl$_2$ (3 x 15 mL). The combined organic phases were dried over Na$_2$SO$_4$ and the solvent was removed in vacuo. The crude residue was purified by flash chromatography (cyclohexane/EtOAc 2:1) to give 16 (252 mg, 0.38 mmol, 70%) as colorless oil. [α]$^2_D$ = +60.1 (c = 1, CHCl$_3$), $R_f$ = 0.13 (cyclohexane/EtOAc 4:1), IR (ATR, wmax, cm$^{-1}$): 2923, 2851, 1749, 1631, 1514, 1437, 1370, 1208, 1129 1036, 835, 755. $^1$H NMR (400 MHz, CDCl$_3$): $\delta_H$ 7.70 (d, $J_{\text{app}} = 8.9 \text{ Hz}$, 2 H, H-4, H-5), 7.33 (d, $^4J_{\text{H-6,H-8}} = 2.4 \text{ Hz}$, 1 H, H-8), 7.12–7.08 (m, 2 H, H-3, H-6), 6.92 (d, $^4J_{\text{H-1,H-2}} = 2.4 \text{ Hz}$, 1 H, H-1), 5.66 (d, $^3J_{\text{H-1',H-2'}} = 1.8 \text{ Hz}$, 1 H, H-1'), 5.60 (dd, $^3J_{\text{H-3',H-4'}} = 10.0 \text{ Hz}$, $^3J_{\text{H-2',H-3'}} = 3.5 \text{ Hz}$, 1 H, H-3'), 5.49 (dd, $^3J_{\text{H-2',H-3'}} = 3.5 \text{ Hz}$, $^3J_{\text{H-1',H-2'}} = 1.8 \text{ Hz}$, 1 H, H-2'), 5.39 (t, $^3J_{\text{H-3',H-4'},H-5'} = 10.0 \text{ Hz}$, 1 H, H-4'), 4.80 (dd, $^2J = 9.5 \text{ Hz}$, $^3J = 4.0 \text{ Hz}$, 1 H, CHCOOMe), 4.31 (dd, $^2J = 12.2 \text{ Hz}$, $^3J_{\text{H-5',H-6a'}} = 5.3 \text{ Hz}$, 1 H, H-6a'), 4.16–4.08 (m, 1 H, H-5'), 4.05 (dd, $^2J = 12.2 \text{ Hz}$, $^3J_{\text{H-5',H-6b'}} = 2.2 \text{ Hz}$, 1 H, H-6b'), 3.75 (s, 3 H, OCH$_3$), 2.22 (s, 3 H, CH$_3$), 2.06 (s, 3 H, CH$_3$), 2.05 (s, 3 H, CH$_3$), 2.00-1.93 (m, 1 H, CH$_2$CHCOOMe) 1.95 (s, 3 H, CH$_3$), 1.82–1.56 (m, 7 H, CH$_2$CHCOOMe, CH, 5 CH$_2$), 1.30–0.90 (m, 5 H, CH$_2$). $^{13}$C NMR, HSQC (101 MHz, CDCl$_3$): $\delta_C$ 173.0 (COOMe), 170.9, 170.3, 170.2, 170.1 (CH$_3$C=O), 157.0, 154.3 (C-2, C-7), 135.7 (C-8a), 129.8 (C-4, C-5), 126.1 (C-4a), 117.9, 116.7 (C-3, C-6), 110.5, 107.5 (C-1, C-8), 96.2 (C-1'), 75.1 (CHCOOMe), 69.8 (C-2'), 69.7 (C-5'), 69.3 (C-3'), 66.4 (C-4'), 62.5 (C-6'), 52.7 (COOCH$_3$), 40.6 (CH$_2$CHCOOMe), 34.3, 34.1, 32.8, 26.7, 26.5, 26.3 (CH, 5 CH$_2$), 21.2, 2x 21.0, 20.9 (CH$_3$C=O).

ESI-MS: $m/z$ (%) = 681.4 (100) [M+Na]$^+$, ESI-HRMS: $m/z$ calcd for [C$_{34}$H$_{42}$O$_{13}$+Na]$^+$: 681.2518, found: 681.2519.
(2S)-3-Cyclohexyl-2-[2-(α-D-mannopyranosyloxy)naphthalen-7-yloxy]-propionic acid (17)
A mixture of 16 (57 mg, 86 µmol) in 1,4-dioxane (10 mL), MeOH (3.6 mL) and 4 N NaOH (1 mL) was stirred 16 h at room temperature. The reaction was quenched with 2 N HCl, the solvent was removed in vacuo and the residue was coevaporated three times with methanol in vacuo. The crude residue was purified by flash chromatography (EtOAc/EtOH 9:1) to give 17 (25 mg, 53 µmol, 61%) as colorless oil. Rf = 0.09 (EtOAc/EtOH 9:1). 1H NMR, COSY (500 MHz, CD3OD): δH 7.69 (d, Japp = 8.9 Hz, 2 H, H-4, H-5), 7.42 (d, 4JH-1,H-3 = 2.1 Hz, 1 H, H-1), 7.09 (dd, 3JH-3,H-4 = 8.9 Hz, 4JH-1,H-3 = 2.1 Hz, 1 H, H-3), 7.04 (m, 2 H, H-6, H-8), 5.61 (d, 3JH-1,H-2' = 1.5 Hz, 1 H, H-1'), 4.80 (dd, 3J = 9.3 Hz, 3J = 3.6 Hz, 1 H, CHCOOH), 4.05 (dd, 3JH-2,H-3' = 3.3 Hz, 3JH-1,H-2' = 1.5 Hz, 1 H, H-2'), 3.95 (dd, 3JH-3,H-4' = 9.4 Hz, 3JH-2,H-3' = 3.3 Hz, 1 H, H-3'), 3.80-3.71 (m, 3 H, H-4', H-6'), 3.64 (ddd, 3JH-4',H-5' = 9.7 Hz, 3JH-5',H-6a' = 4.9 Hz, 3JH-5',H-6b' = 2.7 Hz, 1 H, H-5'), 1.92 (dd, 2J = 14.3 Hz, 3J = 9.3 Hz, 3J = 5.3 Hz, 1 H, CH2CHCOOH), 1.76 (m, 6 H, CH2CHCOOH, 5 1Hex), 1.36–1.15 (m, 4 H, 1Hex), 1.01 (m, 2 H, 1Hex). 13C NMR, HSQC, HMBC (126 MHz, CD3OD): δC 177.2 (COOH), 158.1 (C-7), 156.3 (C-2), 137.1 (C-8a), 130.2 (C-4, C-5), 126.8 (C-4a), 118.1 (C-6), 117.7 (C-3), 111.2 (C-1), 108.1 (C-8), 100.1 (C-1'), 76.2 (CHCOOH), 75.4 (C-5'), 72.5 (C-3'), 72.0 (C-2'), 68.4 (C-4'), 62.7 (C-6'), 41.6 (CH2CHCOOH), 35.5 (CH), 35.0, 33.6, 27.6, 27.4, 27.2 (CH2). ESI-MS: m/z (%) = 494.2 (100) [M+NH4]+, ESI-HRMS: m/z calcd for [C25H32O9+Na]+: 499.1939, found: 499.1957.

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References


