A new construct for the *anti* conformational locking of nucleosides: the dioxomethine transglycosidic tether[†]

Michael P. Groziak*a and Ronghui Linb

^a Pharmaceutical Discovery Division, SRI International 333 Ravenswood Avenue, Menlo Park, CA 94025-3493, USA E-mail: michael.groziak@sri.com

(received 15 Sep 99; accepted 13 Feb 00; published on the web 21 Feb 00)

DOI: http://dx.doi.org/10.3998/ark.5550190.0001.105

Abstract

Two members of a new class of *anti* conformationally locked 2'-deoxynucleoside mimics were synthesized starting from uridine through key 1-(β -D-arabinofuranosyl)uracil-6-carboxaldehyde intermediates. O5'-Mesylation of 1-(β -D-arabinofuranosyl)uracil-6-carboxaldehyde (5c) followed by pyridine-mediated transglycosidic displacement by the dominant 7,O2'-cyclic hemiacetal gave the locked 2',5'-dideoxyuridine mimic (7). Transglycosidic transacetalation in the 7,O2'-cyclic hemiacetal of 1-(β -deoxy-5,5-dimethoxy- β -D-arabinofuranosyl)uracil-6-carboxaldehyde (18) gave the locked O5'-methyl-2'-deoxyuridine mimic (6b). These transglycosidic dioxomethine tether constructions involve proximity-assisted displacement reactions and provide an entry into new, highly biomimetic, *anti* conformationally locked 2'-deoxynucleoside mimics for use as probes of conformation-activity relationships.

Keywords: Transglycosidic displacement, dioxomethine tether, 2'-deoxynucleoside mimics

Introduction

Conformational restriction is a very useful tool for studying the relationship between molecular topography and biochemical or medicinal activity of nucleosides. Transglycosidically tethered analogs are particularly useful when the focus of attention is the glycosidic bond, but in most cases the rotation of this bond is incompletely restricted—some residual rotational flexibility remains. The C6-spiro-fused 5,6-dihydrouridines 1 (X, Y = F, Cl, Br, NO₂) reported by Honjo are a rare exception. They are 5,6-dihydrouridine arabinofuranosides in which both the 2' and 5' hydroxyl group oxygens have been attached to the pyrimidine C6 position. They are, in essence, completely rigidified mimics of the corresponding 2',5'-dideoxy-5,6-dihydrouridines (2). The cage-like carbohydrate moiety in 1 is related in a near-enantiomeric sense to 1,2,5-O-benzylidyne- β -L-arabinofuranose orthoesters reported by Kochetkov³ and to 1,2,5-O-ethylidyne- α -D-galactofuranose orthoesters reported by Bertolini and Glaudemans. 4

The structural differences between 1 and its closest natural counterparts in RNA and DNA (dihydrouridine 3 and thymidine 4, respectively) limit their utility as probes of conformation/activity relationships.⁵ Nevertheless, the idea of transforming arabinofuranosides

ISSN 1551-7004 Page 25 [©]ARKAT USA, Inc

^b Drug Discovery Research, R. W. Johnson Pharmaceutical Research Institute 920 U.S. Route 202, P.O. Box 300, Raritan, NJ 08869-0602, USA E-mail: rlin@prius.jnj.com

into conformationally locked 2'-deoxyribonucleoside mimics by constructing a tether over the β face of the furanose ring has merit and would be particularly valuable if a high degree of biomimicry could be maintained by minimizing structural disturbances.

Some of the findings from our previous investigations⁶ into the chemistry of 6-formyluridine, 6-formyl-2'-deoxyuridine, 6-formyl-1-(β-D-arabinofuranosyl)uracil, and 6-formylthymidine (5ad, respectively) convinced us that this could be accomplished. The structures displayed by these compounds are dictated by their highly electrophilic carboxaldehyde group. While the ribonucleosides 5a,b and d exist as a solvent-sensitive collection of open (hydroxy-aldehyde) and 7,05'-cyclic hemiacetal solution structures, the arabinofuranoside 5c exists exclusively as the 7,02'-cyclic hemiacetal in all solvents examined. When taken together with the fact that 5'carboxaldehyde nucleoside derivatives are known to be susceptible to hydrate and hemiacetal formation, this raised the intriguing possibility that the 5'-carboxaldehyde derivative of 5c might exist at least partly in transglycosidically tethered form (6a). This form would be a highly biomimetic. anti conformationally locked and yet fully *O*-functionalizable deoxyribonucleoside mimic. As the nucleoside or even the nucleotide (i.e., 6, $R = PO_3H_2$), it would be a valuable bioprobe of conformation-activity relationships by virtue of an exceptionally close structural resemblance to naturally occurring counterparts. The anticipation of access not only to separate C5' epimeric versions but also to purine-based versions (from 8-formyl purine nucleosides⁸) only enhanced our interest in this attractive design construct, and so we began working on the construction of dioxomethine-tethered nucleosides.

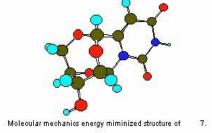
Results and Discussion

We searched for conditions that would assemble the desired dioxomethine tether, and first examined the ease of intramolecular dehydration in 5c to give a cyclic acetal. Even when heated at 65 °C, 5c was inert to exposure to TsOH or TfOH in (CH₃)₂SO solution. The fact that the expected dioxomethine-tethered 2',5'-dideoxyuridine mimic 7 did not form under these conditions reflects the known difficulty of acetal formation in electron-deficient carbonyl

ISSN 1551-7004 Page 26 [©]ARKAT USA, Inc

groups,⁹ seen before for 5.^{6b,c} Conversion of the 5'- or hemiacetal hydroxyl group in 5c to a good leaving group for intramolecular displacement was explored next. Treatment of 5c with an equivalent of MsCl in pyridine at increasingly elevated temperatures eventually produced 7, which was isolated in a 43% yield. Under different conditions, exposure of 5c to (EtO)₃CH in the presence of TfOH gave 7 in a 57% yield.

The dioxomethine-tethered 7 featuring a five-, a six-, a seven, and an eight-membered carbohydrate-associated ring was characterized by ¹H, ¹³C, COSY, and short-range ¹H-¹³C HETCOR NMR, and both low- and high-resolution FAB mass spectral analyses. The ¹H NMR spectrum was especially diagnostic. Both the large (14.2 Hz) geminal coupling of the diastereotopic 5'-CH₂ hydrogens and the moderate (6.6 Hz) vicinal one between H1' and H2' are consistent with the severe conformational restriction caused by the tether. A similar coupling constant pattern was found in the ¹H NMR spectrum of 2',3'-*O*-(isopropylidene)orotidine 5'-lactone, a reference compound we prepared from 2',3'-*O*-isopropylidenated 5a. In addition, only one of the H5' resonances (the downfield *pro-R*) is coupled (5.2 Hz) to the H4' one in 7. This same pattern is displayed not only by 2',3'-*O*-(isopropylidene)orotidine 5'-lactone, but also by certain 6,O5'-methanouridines reported by others. ^{1k} By an MM calculation (Figure 1), 7 has a γvalue (O4'-C1'-N1-C2 dihedral) of 276°, placing the glycosidic torsion in the "high" *anti* range.



The task of preparing a nucleoside like 7 but with an additional oxygen functionality at C5' was addressed next. Deprotection of 2',3'-O-isopropylidene-6-formyluridine 5'-carboxaldehyde¹³ gave dialdehyde 8 (Scheme 1), but we were unable to generate its 2,2'-anhydride with (PhO)₂CO, and thus could not access the arabinofuranoside. As shown in Scheme 1, only the 5'-carboxaldehyde of 8 could be acetalated under standard conditions (60%), and unfortunately a dehydration with (PhO)₂CO intended to give the 2,2'-anhydride gave instead a complex mixture from which the epoxy acetal 10 was isolated as the major product. ¹⁰ The multistep pathway from 9 to 10 apparently involves a hydride transfer from C1' to the carboxaldehyde at some point.

Scheme 1

As shown in Scheme 2, the simultaneous masking of both carboxaldehyde groups in 8 as 1,3-diphenylimidazolidin-2-yls (DPIs) gave 11 (40%), but the reaction was sluggish and actually produced the 6-DPI 5'-dimethyl acetal (14, Scheme 3) to a greater extent (47%).

Scheme 2

Our first attempt to convert 11 to its 2,2'-anhydride employed 1.3 equivalents of (PhO)₂CO. This did give some of the desired 12, but the yield was very low (7%) and the O3'-phenoxycarbonylated derivative and a substantial amount of starting material were isolated. In the second attempt, 3.5 equivalents of (PhO)₂CO and a slightly higher temperature were used to give the O3'-phenoxycarbonyl derivative of 12 in a satisfactory yield (66%). Rapid saponification (NaOH, THF, H₂O, 23 °C, 1 h) of this gave the desired 12 (77%). Longer reaction times produced inseparable mixtures of 12 and the 6,O2'-cyclonucleoside 13, a compound generated by proximity-assisted intramolecular conjugate addition reaction in the open arabinofuranoside. Nucleoside 12 was treated with pTsOH in Me₂CO/CH₂Cl₂ in an attempt to remove both DPI groups prior to opening the 2,2'-anhydro linkage, but this failed.

The same 6-DPI 5'-dimethyl acetal 14 obtained previously from 8 was also obtained from 9, but in a better yield (86%). As shown in Scheme 3, 2,2'-anhydro formation in 14 gave 15 (31%) together with its O3'-phenoxycarbonylated derivative (58%). Saponification of either of these gave the same 6,O2'-cyclonucleoside 16. Selective deprotection of 16 gave the hydrate 17 in a quantitative yield. Facile hydration in the corresponding aldehyde can be attributed to its two electronegative α-heteroatoms. In a transformation critically important to the construction of the dioxomethine tether, hydrate 17 afforded hemiacetal 18 (quantitative) simply upon desiccation. Just as had been found for 5c,^{6b,c} 18 exists as a 3:1 mixture of 7,O2'-cyclic hemiacetal diastereomers. Acid-catalyzed hydrolysis of the 5'-dimethyl acetal in 18 was sluggish, but eventually proceeded to give not the desired 6a, but its open 7, O2'-cyclic hemiacetal instead (60%, 2:1 mixture of diastereomers).

Scheme 3

Attempts to close this to 6a by heating $(CH_3)_2SO$ containing 4Å molecular sieves, in C_6H_6/DMA containing PPTS, in HMDS containing $(NH_4)_2SO_4$, or in pyridine containing Ac_2O were unsuccessful. Similar attempts to access 6b by heating 18 in $(CH_3)_2SO$ containing TfOH or TsOH also failed, but intramolecular acetal formation in 18 was promoted by the action of TsOH in C_6H_6/DMA , and 6b was isolated in a 25% yield. This dioxomethine-tethered O5'-methylated 2'-deoxynucleoside mimic was characterized by 1H and ^{13}C NMR and low- and high-resolution mass spectral analyses. Interestingly, it was isolated as a single diastereomer tentatively assigned the 5'-R configuration because the $^3J4'$ -5' value (3.8 Hz) is close to the non-zero (5.2 Hz) $^3J4'$ -5'-pro-R one observed in 7. The fate of the absent 5'-S diastereomer will be investigated, as will other, milder methods for generating 6a and methods for generating the nucleotide $(6, R = PO_3H_2)$ analogs.

Conclusions

Our synthesis of 7 and 6b demonstrates how a β -facial dioxomethine tether can be constructed onto a natural β -ribonucleoside framework by the careful and deliberate management of proximity-assisted addition and displacement reactions in arabinofuranoside intermediates. The new *anti* conformationally locked nucleosides now accessible are highly biomimetic and should prove to be valuable bioprobes for the study of conformation-activity relationships involving 2'-deoxynucleosides, 2'-deoxynucleotides, and related species.

Experimental Section

General Procedures. Melting points were determined on a Thomas-Hoover UniMelt capillary apparatus and are uncorrected. Radial preparative-layer chromatography was performed on a Chromatotron instrument (Harrison Research, Inc., Palo Alto, CA) using Merck silica gel60 PF₂₅₄ as the adsorbent, flash column chromatography was performed using 230-400 mesh ASTM Merck silica gel-60, and TLC analyses were performed on Analtech 250 µm silica gel GF Uniplates. Lyophilizations were conducted on a Labconco Lypho-Lock 4.5 liter bench-top freeze-dryer. ¹H and ¹³C NMR spectra were recorded on a Varian VXR-300 (300 and 75 MHz) or VXR-500 (500 and 125 MHz) instrument using (CH3)4Si or 2,2-dimethyl-2-silapentane-5sulfonic acid, sodium salt (DSS) (δ = 0.0 for ¹H), and CDCl₃ (δ = 77.0 for ¹³C), (CD₃)2SO (δ = 39.5 for 13 C), or 1,4-dioxane (δ = 66.5 for 13 C in D_2 O) as internal reference. Short-range 1 H- 13 C heteronuclear shift correlation (HETCOR) 2D NMR spectra were obtained on the VXR-300 instrument. Except where noted, the purity of compounds was shown to be >95% by TLC and high-field ¹H NMR. BuLi, iPr2NH, HCO₂Et, (PhNHCH₂)2, 2-iodobenzoic acid, KBrO₃, TsOH, NaBH₄, 2,2'-biquinoline, anhydrous iBuOH, and 1 M TBAF in THF solution were purchased from the Aldrich Chemical Co. TBDMS-Cl and TBDPS-Cl were obtained from Hüls America, Inc. The BuLi was titrated by the modified Watson-Eastham procedure. 11 THF and Et₂O were dried by distillation from Na-benzophenone ketyl under argon. Pyridine and iPr₂NH were dried by distillation from CaH₂ under argon. The HCO₂Et was dried by distillation from P₂O₅ under argon. Dowex-50(H⁺) was obtained from the Sigma Chemical Co., and before use was washed with 1N HCl and then rinsed with distilled water until pH neutral. The Dess-Martin periodinane

ISSN 1551-7004 Page 29 [©]ARKAT USA, Inc

reagent was prepared according to the literature procedure.¹² Elemental microanalyses and mass spectral analyses were obtained from the University of Illinois. MM calculations were performed using Chem3D Pro v.5.0 from Cambridge Scientific Computing, Inc.

(7s)-1-(β-D-Arabinofuranosyl)-6-(dihydroxymethyl)-7,O2':7,O5'-dianhydrouracil (7). Method A. A solution of $5c^{6b,c}$ (30.0 mg, 0.110 mmol) in 0.5 mL of anhydrous pyridine was treated with one equivalent of MsCl (0.2 mL) and the solution was stirred at 0 °C for 4 h, 23 °C for 24 h, and then was heated at reflux for 2 h. The mixture was evaporated to dryness in vacuo and the residue was separated by ascending preparative chromatography on silica gel (15% MeOH/CH₂Cl₂ as eluent) to give 12.0 mg (43%) of 7: ¹H NMR ((CD₃)₂SO) δ 11.6 (1H, bs, NH, exchanges with D₂O), 6.34 (1H, d, H1'), 5.92 (1H, s, H7), 5.79 (1H, s, H5), 5.67 (1H, d, 3'-OH, exchanges with D₂O), 4.67 (1H, d, H2'), 4.41 (1H, d, H3'), 4.25 (1H, d, H4'), 4.08 (1H, d of d, H5'-*pro-R*), 3.49 (1H, d, H5'-*pro-S*); $^3J1'$ -2' = 6.6, $^3J3'$ -3'-OH = 4.8, $^3J4'$ -5'-*pro-R* = 5.2, $^3J4'$ -5'*pro-S* = 0, $^2J5'$ -*pro-R*-5'-*pro-S* = 14.2 Hz. 13 C NMR ((CD₃)₂SO) δ 163.0 (C4), 150.8 (C2), 146.4 (C6), 102.1 (C5), 91.3 (C7), 82.9 (C4'), 79.8 (C2'), 77.3 (C3'), 76.2 (C1'), 68.4 (C5'). Low-resolution FAB-mass spectrum, m/e 255.0 (MH⁺). High-resolution FAB-mass spectrum for C₁₀H₁₁N₂O₆(MH⁺): calcd 255.0617, found 255.0618.

Method B. A solution of $5c^{6b,c}$ (15.0 mg, 0.055 mmol) in 0.5 mL of $(CH_3)_2SO$ was treated with 50 μ L of TfOH and 0.2 mL (excess) of (EtO)₃CH. The reaction mixture was stirred first at 23 °C for 1 h and then at 65 °C for 6.5 h, and then it was evaporated to dryness in vacuo. The residue was separated by the same procedure used in Method A to give 8.0 mg (57%) of 7.

2',3'-*O*-Isopropylideneorotidine 5'-lactone. A solution of 2',3'-*O*-isopropylidenated 5a^{6c} (312 mg, 1.0 mmol) and DCC (0.8 g, 3.9 mmol) in 10 mL of anhydrous (CH₃)₂SO was treated with anhydrous pyridine (0.1 mL) and TFA (0.05 mL), and the mixture was stirred at 23 °C for 50 h. Water (1 mL) was added and the mixture was stirred for additional 0.5 h. The dicyclohexylurea precipitate was removed by suction filtration, and the filtrate was evaporated to dryness at 50 °C in vacuo. Radial chromatography (5% CH₃OH/CH₂Cl₂ as eluent) gave 161 mg (52%) of the lactone as a white solid: mp 270-275 °C (dec.). ¹H NMR (CDCl₃) δ 8.32 (1H, bs, NH), 6.08 (1H, s, H5), 5.98 (1H, d, H1'), 5.02 (1H, m, H5'), 4.97 (1H, d of d, H2'), 4.78 (1H, d, H3'), 4.72 (1H, m, H4'), 4.25 (1H, m, H5'), 1.58 (3H, s, CH₃), 1.37 (3H, s, CH₃). ¹H NMR ((CD₃)₂SO) δ 11.74 (1H, bs, NH), 5.76 (1H, s, H5), 5.73 (1H, d, H1'), 5.11 (1H, d, H5'), 4.68 (1H, t, H3'), 4.63 (1H, t, H2'), 4.55 (d of d, 1H, H4'), 4.17 (1H, t, H5''), 1.45 (3H, s, CH₃), 1.28 (3H, s, CH₃). ¹³C NMR ((CD₃)₂SO) δ 165.5 (C7), 162.8 (C4), 148.7 (C2), 143.1 (C6), 112.0 (C5 or *C*(CH₃)₂), 103.0 (C5 or *C*(CH₃)₂), 95.0, 86.4, 85.4, and 79.7 (each C1', C2', C3', or C4'), 66.2 (C5'), 26.3 and 24.7 (C(CH₃)₂). Low-resolution ACE-mass spectrum, *m/e* 310.2 (M+'), 311.2 (MH+').

6-Formyluridine 5'-carboxaldehyde (8). A solution of 2',3'-*O*-isopropylidene-6-formyluridine 5'-carboxaldehyde¹³ (51.4 mg, 0.17 mmol) in 50% aqueous TFA (1 mL) was stirred at 23 °C for 2 h. The reaction mixture was evaporated to dryness, and residual TFA was removed from the residue by repetitive azeotropic coevaporation with water. Lyophilization afforded NMR-pure 8 in a quantitative yield: 210-220 °C (dec.). The NMR spectral features of 8 in D₂O solution were consistent with dihydrate structure: ¹H NMR (D₂O) δ 6.14 (s, 1H, H5), 5.98 (m, 2H, H1' and H7), 5.12 (d, 1H, H5'), 4.84 (s, H2' under HOD), 4.50 (pseudo-t, 1H, H3'), 3.80 (pseudo-t, 1H, H4'); ³J1'-2' not well resolved, ³J2'-3' = 6.2, ³J3'-4' = 5.9, ³J4'-5' = 5.7 Hz. ¹³C NMR (D₂O) δ 165.7 (C4), 156.0 (C2), 151.6 (C6), 100.2 (C5), 92.1 (C1'), 89.9 (C5'), 85.8 (C7), 85.5 (C4'), 71.6 (C2'), 70.3 (C3'). UV λmax, nm (ε× 10⁻³): (H₂O) 261 (8.9), 204 (9.2); (pH 1) 262 (7.8), 209 (7.2).

ISSN 1551-7004 Page 30 [©]ARKAT USA, Inc

Low-resolution CIMS, m/e 271.1 (MH⁺); High-resolution CIMS calcd for $C_{10}H_{11}N_2O_7$ (MH⁺): calcd 271.0566, found 271.0564.

6-Formyluridine 5'-carboxaldehyde, 5'-dimethyl acetal (9). A suspension of 8 (2.00 g, 7.40 mmol) in 20 mL of absolute MeOH containing TfOH (100 μL) was heated at reflux under argon. After 20 min, the mixture became homogeneous and 8 had been consumed, by TLC analysis. The solution was concentrated in vacuo and the residue was purified by radial chromatography (10% MeOH/CH₂Cl₂ as eluent) to give, after Abderhalden (P₂O₅) drying, 1.40 g (60%) of 9 as a yellow foam: ¹H NMR (D₂O) δ 6.12 (1H, s, H5), 5.96 (1H, d, H1'), 5.95 (1H, s, hemiacetal CH), 4.81 (1H, m, H2'), 4.62 (1H, d, H5'), 4.54 (1H, pseudo-t, H3'), 3.89 (1H, pseudo-t, H4'), 3.51 and 3.44 (each 3H, each s, each OCH₃); ³J1'-2' = 3.3, ³J3'-4' = 6.6, ³J4'-5' = 6.9 Hz. ¹³C NMR (D₂O) δ165.7 (C4), 155.9 and 151.4 (C₂/C6), 104.2 (C5'), 100.2 (C5), 92.4 (C7), 85.8 (C1'), 82.2 (C4'), 72.0 (C2'), 70.3 (C3'), 55.3 and 54.2 (two CH₃O).

1-(1-Dehydro-1-hydroxy-5-deoxy-5,5-dimethoxy-1,2-anhydro-α-lD-arabinofuranosyl)-6hydroxymethyluracil (10). A solution of monoacetal 9 (100.0 mg, 0.316 mmol) and (PhO)₂CO (101.5 mg, 0.474 mmol) in 0.3 mL of anhydrous DMA was treated with NaHCO₃ (2.0 mg) and then was heated at 140-150 °C for 15 min. The solution was concentrated in vacuo, and the oily residue treated with 5 mL of anhydrous Et₂O. The resulting solid was collected, rinsed with additional anhydrous Et₂O (3 × 3 mL), and then purified by ascending preparative chromatography (10% MeOH/CH₂Cl₂ as eluent) to give 26.6 mg (27%) of 10: ¹H NMR (D₂O) δ 5.74 (1H, s, H5), 5.49 (1H, s, H2'), 4.94 and 4.74 (each 1H, each d, each H7), 4.55 (1H, d, H5'), 3.99 (1H, d, H3'), 3.95 (1H, d of d, H4'), 3.48 and 3.42 (each 3H, each s, each OCH₃); ${}^{3}J2'-3'=0$, $^{3}J3'-4' = 2.8$, $^{3}J4'-5' = 7.3$, $^{2}J7a-7b = 16.8$ Hz. ^{1}H NMR ((CD₃)₂SO) $\delta 11.5$ (1H, bs, NH, exchanges with D₂O), 7.3 (1H, bs, 7-OH, exchanges with D₂O), 5.7 (1H, d, 3'-OH, exchanges with D₂O), 5.53 (1H, s, H5), 5.30 (1H, s, H2'), 4.65 (2H, q, CH2), 4.28 (1H, d, H5'), 4.71 (1H, d of d, H4'), 3.62 (1H, d of d, H3'), 3.26 and 3.22 (each 3H, each s, each OCH₃); ${}^{3}J2'-3'=0$, ${}^{3}J3'-4'$ = 2.2, ${}^{3}J3'$ -3'-OH = 6.0, ${}^{3}J4'$ -5' = 7.7 Hz. ${}^{13}C$ NMR ((CD₃)₂SO) δ 162.0 (C4), 151.0 and 148.0 (C2/C6), 103.1 (C5'), 99.0 (C1'), 96.7 (C5), 82.8 (C4'), 78.2 (C2'), 74.4 (C3'), 57.6 (CH₂), 54.0 and 52.7 (two CH₃O). Low-resolution FAB-mass spectrum, m/e 317.1 (MH⁺). High-resolution FAB-mass spectrum for $C_{12}H_{17}N_2O_8(MH^+)$: calcd 317.0985, found 317.0986.

5'-Deoxy-5',6-bis-(1,3-diphenylimidazolidin-2-yl)uridine **(11)** and 6 - (1.3 diphenylimidazolidin-2-yl) uridine-5'-carbox- aldehyde, dimethyl acetal (14). A solution of crude 8 prepared from 200 mg of its 2',3'-O-isopropylidene derivative, 11 (PhNHCH₂)₂ (303 mg, 1.42 mmol), and 0.12 mL of glacial AcOH in 10 mL of absolute CH₃OH was stirred for 3 d at 23 °C. The reaction mixture was partitioned between saturated aqueous NaHCO₃ and CH₂Cl₂, and the layers were separated and the aqueous phase was extracted with fresh CH2C12. The organic solutions were combined, dried (MgSO₄), and then rotary evaporated to dryness. Column chromatography (5% CH₃OH/CH₂Cl₂ as eluent) gave 168 mg (40%) of 11 as a pink powder and 155 mg (47%) of the 6-DPI, 5'-dimethyl acetal derivative (14) as a yellowish foam. For 11: ¹H NMR (CDCl₃) δ 9.6 (1H, bs, NH), 7.35-6.58 (20H, m, four C₆H₅); 5.90 (1H, s, H1'), 5.78 (1H, s, H5), 5.68 (1H, d, H5'), 5.46 (1H, s, H7), 4.64-4.61 (2H, m, H2' and H3'), 3.90 (1H, d of d, H4'), 3.72-3.25 (10H, m, 2'-OH, 3'-OH, and two NCH₂CH₂N); ${}^{3}J1'-2'=0$, ${}^{3}J3'-4'=7.2$, ${}^{3}J4'-5'=1.9$ Hz. ¹³C NMR (CDCl₃) δ 163.0 (C4), 153.4, 150.8, 147.6, 146.4, 146.3, and 145.2 (C2, C6, and four C₆H₅), 129.5, 129.4, 129.1, 128.9, 122.5, 119.6, 118.3, 117.6, 117.4, 114.3, 113.5 and 113.0 (four C_6H_5), 102.7 (C5), 91.7 (C7), 83.3 (C4'), 75.6 (C1'), 73.3 (C5'), 72.1 and 69.7 (C2'/C3'), 50.8, 47.0, 46.8 and 46.1 (two NCH₂CH₂N). Low-resolution CI-mass spectrum, m/e 335.2 (80%, B+1), 306.2 (30%, M⁺-B-H₂O), 223.1 (100%). Low-resolution EI-mass spectrum, m/e 334.1

ISSN 1551-7004 Page 31 [©]ARKAT USA, Inc

(15%, B⁺), 306.1 (10%, M⁺-B-H₂O), 223.1 (100%). Low-resolution FAB-mass spectrum, m/e 659.2 (80%, MH⁺). 14: ¹H NMR (CDCl₃) δ 10.2 (1H, bs, NH), 7.28-6.69 (10H, m, two C₆H₅), 5.90 (1H, s, H5), 5.82 (1H, s, H7), 5.62 (1H, d, H1'), 4.75 (1H, d of d, H2'), 4.53-4.49 (2H, m, H3' and H5'), 3.64 (1H, pseudo-t, H4'), 3.61-3.40 (6H, m, NCH₂CH₂N, 2'-OH, and 3'-OH), 3.39 and 3.37 (each 3H, each s, each CH₃O); ³J1'-2' = 2.4, ³J2'-3' = 6.3, ³J4'-5' = 7.2 Hz. ¹³C NMR (CDCl₃) δ 63.2 (C4), 153.4, 151.4, 147.3, 145.7 (C2, C6, and two C₆H₅), 129.5, 129.4, 122.2, 120.3, 118.1 and 115.3 (two C₆H₅), 104.0 (C5'), 102.5 (C5), 92.3 (C7), 82.1 (C4'), 75.9 (C1'), 72.5 and 70.5 (C2'/C3'), 54.7 and 53.4 (two CH₃O), 50.5 and 47.5 (NCH₂CH₂N). Low-resolution CI-mass spectrum, m/e 511.3 (25%, MH⁺), 479.3 (40%, MH⁺-CH₃OH), 447.3 (40%, MH⁺-2CH₃OH), 223.2 (100%). Low-resolution CI-mass spectrum for C₂₆H₃₁N₄O₇ (MH⁺): calcd 511.2192, found 511.2170.

5'-Deoxy-5',6-bis-(1,3-diphenylimidazolidin-2-yl)-2,2'-anhydrouridine (12). A solution of 11 (110.0 mg, 0.167 mmol) and (PhO)₂CO (46 mg, 0.215 mmol) in 0.3 mL of anhydrous DMF was treated with 1 mg of NaHCO3 and was heated at 110-130 °C for 0.5 h and then concentrated to dryness in vacuo. The mixture was then separated by ascending preparative chromatography using 5% MeOH/CH₂Cl₂ as eluent to give 12 (8.0 mg, 7%) and its 3'-phenyl carbonate (25.2 mg, 20%). Treatment of 11 (80 mg, 0.121 mmol) with excess (PhO)₂CO (92 mg, 0.429 mmol), additional NaHCO3 (2.0 mg), and at 135-155 °C in this procedure gave the O3'phenoxycarbonylated derivative of 12 in a 66% yield. 12: ¹H NMR (CDCl₃) δ 7.35-6.55 (20H, m, four C6H5), 6.10 (1H, d, H1'), 6.06 (1H, s, H5), 6.03 (1H, s, H7), 5.84 (1H, s, 3'-OH), 5.53 (1H, s, H5'), 4.89 (1H, d of d, H2'), 4.52 (1H, m, H3'), 4.25 (1H, d, H4'), 3.80-3.32 (8H, m, two NCH₂CH₂N); ${}^{3}J1'-2' = 5.7$, ${}^{3}J2'-3' = 2.1$ Hz, ${}^{3}J3'-4' = 6.6$. ${}^{13}C$ NMR (CDCl₃) δ 172.4 (C4), 160.7 (C2), 148.0, 147.2, 146.5, 145.8, 144.8 (C6 and four C_6H_5), 129.7, 129.6, 129.4, 129.2, 122.5, 118.8, 118.2, 118.1, 117.8, 113.4, 113.3, and 112.8 (two C₆H₅), 108.1 (C₅), 89.4 (C₂'), 86.7 (C1'), 85.7 (C4'), 74.9 (C3'), 73.8 (C7), 73.5 (C5'), 50.8, 47.0, 46.9 and 45.6 (two NCH₂CH₂N). Low-resolution FAB-mass spectrum, m/e 641.3 (70%, MH⁺), 223.2 (100%). High-resolution FAB-mass spectrum for C₃₈H₃₇N₆O₄(MH⁺): calcd 641.2876, found 641.2887. O3'-Phenoxycarbonylated 12: ¹H NMR (CDCl₃) δ 7.45-6.62 (25H, m, five C₆H₅), 6.47 (1H, d, H1'), 6.14 (1H, s, H5), 5.96 (1H, s, H7), 5.79 (1H, d, H5'), 5.60 (1H, d, H3'), 5.27 (1H, d, H2'), 4.82 (1H, d of d, H4'), 3.80-3.35 (8H, m, two NCH₂CH₂N); ${}^{3}J1'-2' = 5.4$, ${}^{3}J3'-4' = 2.7$, ${}^{3}J4'-5' = 1.2$ Hz. ¹³C NMR (CDCl₃) δ171.6 (C4), 160.4 (C2), 152.6, 150.5, 147.8, 147.3, 146.2, 145.8, 144.9 $(C_6, C_6H_5OCO_2, and five C_6H_5), 129.8, 129.7, 129.6, 129.5, 129.4, 126.7, 122.5, 120.6, 119.6,$ 119.3, 118.7, 117.8, 114.8, 113.6 and 113.2 (three C₆H₅), 109.6 (C5), 88.5 (C4'), 88.3 (C1'), 85.8 (C2'), 78.0 (C3'), 74.5 (C7), 73.4 (C5'), 50.7, 49.3, 47.6 and 46.0 (two NCH₂CH₂N). Lowresolution FAB-mass spectrum, m/e 761.3 (60%, MH⁺).

(6R)-6,O2'- Anhydro-1-[5-deoxy-5-(1,3-diphenylimidazolidin-2-yl)-β]-D-arabinofuranosyl]-6-(1,3-diphenylimidazo-lidin-2-yl)-5,6-dihydrouracil (13). Saponification of 60 mg (0.079 mmol) of the O3'-phenoxycarbonylated 12 in a mixture of 0.123 mL of 1 M NaOH, 0.9 mL of H₂O, and 1.4 mL of THF at 23 °C for 0.5 h gave, after chromatographic separation, 39.1 mg (77%) of 12. Saponification of 12 in THF/MeOH/H₂O at 23 °C for 2.5 h afforded an inseparable mixture of 12 and the dihydrouracil 13 by ¹H NMR analysis. 13: ¹H NMR (CDCl₃) δ 2.94 and 2.29 (each 1H, each d, H5a and H5b), 2.8 (1H, bs, 3'-OH); ²*J*5a-5b = 15.9 Hz).

5'-Deoxy-5',5'-dimethoxy-6-(1,3-diphenylimidazolidin-2-yl)-2,2'-anhydrouridine (15). A solution of 14 (120 mg, 0.235 mmol), (PhO)₂CO (100.7 mg, 0.47 mmol) and NaHCO₃ (2.0 mg) in 0.5 mL of DMF was heated at 135-155 °C for 1 h. The solution was rotary evaporated to

ISSN 1551-7004 Page 32 [©]ARKAT USA, Inc

dryness in vacuo, and the residue purified by chromatography to afford 15 (35.8 mg, 31%) and 3'-phenoxycarbonylated 15 (83.7 mg, 58%). 15: 1 H NMR (CDCl₃) δ 7.32-6.66 (10H, m, two C₆H₅), 6.53 (1H, d, H1'), 6.23 and 6.03 (each 1H, each s, H5 and H7), 6.20 (1H, bs, 3'-OH, exchanges with D₂O), 5.43 (1H, d, H2'), 4.75 (1H, s, H3'), 4.42 and 4.24 (each 1H, each d, H4'/H5'), 3.75-3.45 (4H, m, NCH₂CH₂N), 3.30 (6H, s, two CH₃O); 3 J1'-2' = 6.3, 3 J4'-5' = 3.3 Hz. 13 C NMR (CDCl₃) δ 173.4 (C4), 161.4 (C2), 148.7, 148.2, and 144.5 (C6 and two C₆H₅), 129.5, 129.4, 122.8, 118.8, 118.1, and 112.4 (two C₆H₅), 107.3 and 103.3 (C5/C5'), 90.1, 88.9, 88.8, 76.5, and 73.1 (C1', C4', C2', C3', and C7), 56.2 and 55.9 (each CH₃O), 51.7 and 45.0 (NCH₂CH₂N). Low-resolution FAB-mass spectrum, *m/e* 493.1 (35%, MH⁺). High-resolution FAB-mass spectrum for C₂6H₂9N₄O₆(MH⁺): calcd 493.2087, found 493.2090.

O3'-Phenoxycarbonylated 15: 1 H NMR (CDCl₃.) δ 7.42-6.65 (15H, m, three C₆H₅), 6.66 (1H, d, H1'), 6.27 and 6.12 (each 1H, each s, H5 and H7), 5.42 (1H, s, H3'), 5.33 (1H, d, H2'), 4.45 (1H, d, H4'), 4.25 (1H, d, H5'), 3.75-3.35 (8H, m, two NCH₂CH₂N), 3.32 (6H, s, two CH₃O); 3 J1'-2' = 6.0, 3 J4'-5' = 3.0 Hz. 13 C NMR (CDCl₃) δ 173.2 (C4), 161.1 (C2), 152.4, 150.5, 148.9, 147.8, 144.6 (C6, C₆H₅OCO₂, and three C₆H₅), 129.6, 129.5, 129.3, 126.5, 123.0, 120.5, 118.2, 115.4, and 112.5 (three C₆H₅), 107.7 (C5), 102.9 (C5'), 89.9 (C1'), 86.0 (C4'), 85.1 (C3'), 81.4 (C2'), 73.1 (C7), 56.8 and 56.6 (each CH₃O), 52.2 and 44.9 (NCH₂CH₂N). Low-resolution FAB-mass spectrum, m/e 613.3 (40%, MH⁺). High-resolution FAB-mass spectrum for C₃₃H₃₃N₄O₈(MH⁺): calcd 613.2298, found 613.2288.

(*6R*)-6,O2'-Anhydro-1-(5-deoxy-5,5-dimethoxy-β-D-arabinofuranosyl)-6-(1,3-diphenylimidazolidin-2-yl)-5,6-dihy-drouracil (16). Saponification of the O3'-phenoxycarbonylated 15 from above (83.7 mg, 0.136 mmol) in a mixture of 0.2 mL of 1M NaOH and 2.0 mL of 50% aqueous THF at 23 °C overnight gave, after ascending chromatographic purification (silica gel, 5% MeOH/CH₂Cl₂ as eluent), the dihydrouridine 16 (62.7 mg, 90%). The intermediacy of 15 in this transformation was verified by TLC. A similar saponification of 15 (25.0 mg, 0.136 mmol) for 24 h also gave 16 (23.8 mg, 92%) in a reaction which when halted after only 12 h was found to afford 1-(5-deoxy-5,5-dimethoxy-β-D-arabinofuranosyl)-6-(1,3-diphenylimidazolidin-2-yl)uracil (19%) in addition to 16 (66%).

16: 1 H NMR (CDCl₃) δ 7.5 (1H, bs, NH), 7.25-6.70 (10H, m, two C₆H₅), 5.92 (1H, d, H1'), 5.71 (1H, s, H7), 4.49 (1H, d, H5'), 4.43 (1H, d of d, H2'), 4.29 (1H, d, H3'), 3.85-3.30 (5H, m, H4' and NCH₂CH₂N), 3.47 and 3.46 (each 3H, each s, each CH₃O), 2.97 and 2.43 (each 1H, each d, H5a and H5b), 2.5 (1H, bs, 3'-OH); 3 J5a-5b = 15.9, 3 J1'-2' = 4.2, 3 J3'-4' = 6.6, 3 J4'-5' = 4.5 Hz. 13 C NMR (CDCl₃) δ 165.7 (C4), 150.1, 147.8 and 146.3 (C2 and two C₆H₅), 129.1, 128.9, 119.8, 119.0 116.3 and 114.1 (two C₆H₅), 104.1 (C5'), 100.2 (C6), 89.2 (C2'), 88.8 (C1'), 84.6 (C4'), 77.7 (C7), 74.4 (C3'), 56.7 and 55.1 (each CH₃O), 49.8 and 48.4 (NCH₂CH₂N), 38.5 (C5). Low-resolution CI-mass spectrum, m/e 511.3 (30%, MH⁺), 479.2 (30%, MH⁺-MeOH). High-resolution CI-mass spectrum for C₂₆H₃₁N₄O₇(MH⁺): calcd 511.2193, found 511.2183.

1-(5-Deoxy-5,5-dimethoxy-β-D-arabinofuranosyl)-6-(1,3-diphenylimidazolidin-2-yl)uracil. ¹H NMR (CDCl₃) δ 9.0 (1H, br, NH, exchanges with D₂O), 7.32-6.50 (10H, m, two C₆H₅), 6.22 (1H, d, H1'), 6.00 and 5.81 (each 1H, each s, H7/H5), 4.56 (2H, pseudo-t, H5'/H2'), 4.48 (1H, d, H2'), 4.29 (1H, d, H3'), 3.75 (1H, d of d, H4'), 3.58 and 3.27 (each 3H, each m, NCH₂CH₂N), 3.44 and 3.40 (each 3H, each s, each CH₃O), 2.6 and 1.8 (each 1H, each br, 2'-OH/3'-OH, exchanges with D₂O); 3 J5a-5b = 15.9, 3 J1'-2' = 4.2, 3 J3'-4' = 6.6, 3 J4'-5' = 4.5 Hz. 13 C NMR (CDCl₃) δ162.4 (C4), 150.9, 150.8, 147.8 and 146.0 (C2, C6, and two C₆H₅), 129.3, 129.2, 122.2, 119.3, 117.4 and 112.8 (two C₆H₅), 103.6 and 100.7 (C5/C5'), 82.5, 81.2, 80.8, 76.8 and 76.0 (C7/C1'/C4'/C2'/C3'), 56.6 and 55.4 (each CH₃O), 47.5 and 41.7 (NCH₂CH₂N). Low-

ISSN 1551-7004 Page 33 [©]ARKAT USA, Inc

resolution FAB-mass spectrum, m/e 511.3 (40%, MH⁺), 479.2 (20%, MH⁺-MeOH). High-resolution FAB-mass spectrum for $C_{26}H_{31}N_4O_7(MH^+)$: calcd 511.2193, found 511.2199.

(*6R*)-6,O2'-Anhydro-1-(5-deoxy-5,5-dimethoxy-β-D-arabinofuranosyl)-6-(dihydroxy-methyl)-5,6-dihydro-uracil (17). When an attempt at hydrolyzing the DPI group in 16 using TsOH in Me₂CO/CH₂Cl₂ gave a chromatographically inseparable mixture, this deprotection was instead effected by stirring a mixture of 16 (32.9 mg, 0.064 mmol) and Dowex 50 (500 mg, H⁺form) in 2.0 mL of 67% aqueous THF at 23 °C for 3 d. Upon a removal of the resin and solvents, hydrate 17 was isolated (21.5 mg, quantitative). ¹H NMR ((CD₃)₂SO) δ 10.3 (1H, bs, NH, exchanges with D₂O), 6.45 (1H, d, hydrate OH, exchanges with D₂O), 6.15 (1H, d, exchanges upon addition of D₂O, hydrate OH), 5.89 (1H, d, H1'), 5.54 (1H, d, exchanges, 3'-OH), 4.75 (1H, pseudo-t, hydrate CH), 4.44 (1H, d, H2'), 4.41 (1H, d, H5'), 4.01 (1H, pseudo-t, H3'), 3.73 (1H, d of d, H4'), 3.32 and 3.27 (each 3H, each s, each CH₃O), 2.87 (1H, d, H5a), 2.54 (1H, d, H5b); ²J5a-5b = 15.8, ³J7-7-OH = 4.4 and 5.8, ³J1'-2' = 4.0, ³J3'-3'-OH = 5.4, ³J4'-5' = 7.2 Hz. ¹³C NMR ((CD₃)₂SO) δ 168.7 (C4), 151.1 (C2), 102.9 (C5'), 95.2 (C6), 91.8 (C7), 89.4 (C1'), 87.8 (C2'), 84.8 (C4'), 74.9 (C3'), 54.1 and 52.2 (two CH₃O), 34.2 (C5). Low-resolution FAB-mass spectrum, m/e 317.0 (20%, MH⁺-H₂O), 309.0 (100%, MH⁺-2H₂O).

1-(5-Deoxy-5,5-dimethoxy-β-D-arabinofuranosyl)uracil-6-carboxaldehyde 7,O2'-cyclichemiacetal (**18**). Compou-nd 17 in $(CD_3)_2SO$ solution was observed by 1H NMR to slowly equilibrate to a quaternary mixture of hydrate 17, its corresponding aldehyde, and the diastereomers of 18. This process was inhibited by the presence of water and promoted by heating or desiccation with 4Å molecular sieves. Aldehyde: 1H NMR ($(CD_3)_2SO$) δ 10.8 (1H, br, NH, exchanges with D₂O), 9.35 (1H, s, CHO), 5.99 (1H, d, H1'), 5.71 (1H, d, 3'-OH, exchanges with D₂O), 4.65 (1H, d, H2'), 4.42 (1H, d, H5'), 4.13 (1H, pseudo-t, H3'), 3.79 (1H, pseudo-t, H4'), 3.30 and 3.29 (each 3H, each s, each CH₃O), 3.04 (1H, d, H5a), 2.85 (1H, d, H5b); 2J 5a-5b = 15.6, 3J 1'-2' = 3.8, 3J 3'-3'-OH = 5.5, 3J 4'-5' = 5.2 Hz.

Thus, 18 was obtained (quantitative by ¹H NMR) by heating 17 in (CD₃)₂SO solution at 45 °C for 6 h over 4Å molecular sieves. As had been found for 5c, 6b,c the ¹H NMR spectrum of 18 in (CD₃)₂SO solution revealed the presence of a 3:1 mixture of the two 7,O2'-cyclic hemiacetal diastereomers. Major diastereomer: ¹H NMR ((CD₃)₂SO) δ 11.5 (1H, bs, NH, exchanges with D₂O), 7.63 (1H, d, hemiacetal OH, exchanges with D₂O), 5.77 (1H, d, 3'-OH, exchanges with D₂O), 5.67 (1H, d, hemiacetal CH), 5.63 (1H, d, H1'), 5.62 (1H, s, H5), 4.39 (1H, d, H2'), 4.26 (1H, d, H5'), 4.04 (1H, d, H3'), 3.73 (1H, d, H4'); $^3J7-7-OH = 6.4, ^3J1'-2' = 2.6,$ $^{3}J3'-3'-OH = 4.8$, $^{3}J4'-5' = 7.6$ Hz. ^{13}C NMR ((CD₃)₂SO) δ 162.5 (C4), 150.6 and 148.8 (C2/C6), 102.9 (C5'), 99.5 (C5), 87.1 (C7), 84.2 (C4'), 78.0 (C1'), 75.4 (C3'), 73.8 (C2'), 54.2 and 54.8 (two CH₃O). Minor: ¹H NMR ((CD₃)₂SO) δ 11.5 (1H, br, NH, exchanges with D₂O), 7.92 (1H, d, hemiacetal OH, exchanges with D₂O), 5.78 (1H, d, 3'-OH, exchanges with D₂O), 5.76 (1H, s, H5), 5.54 (1H, d, H1'), 5.46 (1H, d, hemiacetal CH), 4.32 (1H, d, H2'), 4.23 (1H, d, H5'), 4.04 (1H, d, H3'), 3.72 (1H, d, H4'); ${}^{3}J7-7-OH = 7.7$, ${}^{3}J1'-2' = 2.4$, ${}^{3}J3'-3'-OH = 4.9$, ${}^{3}J4'-5' = 8.0$ Hz. 13 C NMR ((CD₃)₂SO) δ 162.4 (C4), 151.0 and 150.8 (C2/C6), 102.9 (C5'), 98.1 (C5), 89.3 (C7), 84.3 (C4'), 78.6 (C1'), 77.7 (C2'), 75.4 (C3'), 54.0 and 52.5 (two CH₃O). Low-resolution CI-mass spectrum, m/e 317.1 (50%, MH⁺), 309.0 (100%, MH⁺-CH₃OH). High-resolution CI-mass spectrum for $C_{12}H_{17}N_2O_8(MH^+)$: calcd 317.0985, found 317.0987.

Deprotection of 18. Hydrolysis of the 5'-(dimethyl) acetal in 18 (18.3 mg, 0.058 mmol) with 50% aqueous TFA (0.5 mL) was sluggish (23 °C, 4 d), but proceeded. The residue obtained upon rotary evaporation to dryness in vacuo was separated by ascending preparative chromatography (silica gel, 15% MeOH/CH₂Cl₂ as eluent), and the purified compound was dried (Abderhalden,

ISSN 1551-7004 Page 34 [©]ARKAT USA, Inc

P₂O₅, 45-50 °C). Lyophilization of an aqueous solution followed by desiccation gave the deprotected material (8.4 mg, 54%) found to exist as a 2:1 mixture of 7,O2'-cyclic hemiacetal diastereomers in (CD₃)₂SO solution, by ¹H NMR. Major diastereomer: ¹H NMR ((CD₃)₂SO) δ11.6 (1H, bs, NH), 9.41 (1H, s, 4'-CHO), 7.77 (1H, d, 7-OH), 6.20 (1H, d, 3'-OH), 5.84 (1H, d, H1'), 5.64 (1H, d, H7), 5.63 (1H, s, H5), 4.47 (1H, d, H2'), 4.30 (1H, s, H4'), 4.25 (1H, d, H3'). ¹³C NMR ((CD₃)₂SO) δ201.4 (C5'), 158.1 (C4), 150.7 and 148.1 (C2/C6), 99.9 (C5), 89.2 (C7), 87.1 (C4'), 79.6 (C1'), 76.9 and 71.4 (C2'/C3'); 3J7-7-OH = 6.5 Hz. Minor: ¹H NMR ((CD₃)₂SO) δ 11.6 (1H, bs, NH), 9.42 (1H, s, 4'-CHO), 8.00 (1H, d, 7-OH), 6.20 (1H, d, 3'-OH), 5.79 (1H, d, H1'), 5.60 (1H, s, H5), 5.50 (1H, d, H7), 4.37 (1H, d, H2'), 4.26 (1H, s, H4'), 4.24 (1H, d, H3'); 3J7-7-OH = 7.7 Hz. ¹³C NMR ((CD₃)₂SO) δ 201.6 (C5'), 157.7 (C4), 150.6 and 148.3 (C2/C6), 98.3 (C5), 89.1 (C7), 89.0 (C4'), 79.9 (C1'), 77.2 and 75.9 (C2'/C3').

(5'-R)-1-(5-Methoxy-β-D-arabinofuranosyl)-6-(dihydroxymethyl)-7,O2':7,O5'-

dianhydrouracil (**6b**). A solution of 18 (20.0 mg) in a mixture of anhydrous DMA (0.2 mL) and anhydrous C6H6 (0.4 mL) was treated with pTsOH monohydrate (0.3 mg). The resultant mixture was heated at reflux for 24 h under argon and then was evaporated to dryness in vacuo. The residue was dissolved in a small amount of methanol and separated by ascending preparative chromatography twice (silica gel, 10% MeOH/CH₂Cl₂ as eluent) to give 6b as a gum (4.5 mg, 25%). ¹H NMR ((CD₃)₂SO) δ 11.5 (1H, br, NH), 6.37 (1H, d, H1'), 5.83 and 5.77 (each 1H, each s, H5 and H7), 5.71 (1H, d, 3'-OH), 4.93 (1H, d, H5'), 4.67 (1H, d, H2'), 4.38 (1H, d, H3'), 4.21 (1H, d, H4'), 3.23 (3H, s, CH3O); 3 J1'-2' = 6.4, 3 J3'-3'-OH = 4.3, 3 J4'-5' = 3.8 Hz. 13 C NMR ((CD₃)₂SO) δ 162.4 (C4), 150.8 and 149.1 (C2/C6), 99.4 and 99.2 (C5'/C5), 88.0 (C7), 83.1 (C4'), 79.7 (C1'), 76.7 and 76.4 (C2'/C3'), 55.3 (CH₃O). Low-resolution FAB-mass spectrum, *m/e* 285.1 (40%, MH⁺). High-resolution FAB-mass spectrum for C₁₁H₁₃N₂O₇(MH⁺): calcd 285.0723, found 285.0722.

References

[†]This work was performed at Southern Illinois University where it was supported in part by grants from SIU's Office of Research Development and Administration. Current support from NIH (NIGMS56878) is gratefully acknowledged. This work has been communicated in brief, preliminary form: Groziak, M. P.; Lin, R. *Nucleosides Nucleotides* 1997, *16*, 1419.

(a) Kittaka, A.; Kato, H.; Tanaka, H.; Nonaka, Y.; Amano, M.; Nakamura, K. T.; Miyasaka, T. Tetrahedron 1999, 55, 5319. (b) Wang, G.; Girardet, J.-L.; Gunic, E. Tetrahedron 1999, 55, 7707. (c) Marquez, V. E.; Ezzitouni, A.; Russ, P.; Siddiqui, M. A.; Ford, H.; Feldman, R. J.; Mitsuya, H.; George, C.; Barchi, J. J. J. Am. Chem. Soc. 1998, 120, 2780. (d) Björsne, M.; Szabó, T.; Samuelsson, B.; Classon, B. Bioorg. Med. Chem. 1995, 3, 397. (e) Nielsen, P.; Pfundheller, H. M.; Wengel, J. J. Chem. Soc., Chem. Commun. 1997, 825. (f) Bar, N. C.; Patra, R.; Achari, B.; Mandal, S. B. Tetrahedron 1997, 53, 4727. (g) Steffens, R.; Leumann, C. J. J. Am. Chem. Soc. 1997, 119, 11548. (h) Bolli, M.; Lubini, P.; Leumann, C. Helv. Chim. Acta 1995, 78, 2077. (i) V. Tetrahedron Lett. 1995, 36, 7375. (j) Rao, S. N. Nucleosides Nucleotides 1995, 14, 1179. (k) Megati, S.; Ealick, S. E.; Naguib, F. N. M.; el Kouni, M. H.; Klein, R. S.; Otter, B. A. Nucleosides Nucleotides 1994, 13, 2151. (l) Plavec, J.; Tong, W.; Chattopadhyaya, J. J. Am. Chem. Soc. 1993, 115, 9734. (m) Chemla, P. Tetrahedron Lett. 1993, 46, 7391. (n) Hsu, L.-H.; Wise, D. S.; Kucera, L. S.; Drach, J. C.;

ISSN 1551-7004 Page 35 [©]ARKAT USA, Inc

Townsend, L. B. *J. Org. Chem.* **1992**, *57*, 3354. (o) Koole, L. H.; Wu, J.-C.; Neidle, S.; Chattopadhyaya, J. *J. Am. Chem. Soc.* **1992**, *114*, 2687. (p) Otter, B. A.; Patil, S. A.; Spada, M. R.; Jelicks, L. A.; Yoshimura, Y.; Matsuda, A.; Klein, R. S. *Nucleosides Nucleotides* **1992**, *11*, 615. (q) Hsu, L.-Y.; Wise, D. S.; Drach, J. C.; Townsend, L. B. *Chin. Pharm. J.* **1991**, *43*, 275. (r) Ueda, T. *Nucleosides Nucleotides* **1985**, *4*, 67. (s) Kameyama, K.; Sako, M.; Hirota, K.; Maki, Y. *J. Chem. Soc., Chem. Commun.* **1984**, 1658.

- 2. (a) Maruyama, T.; Kimura, S.; Sato, Y.; Honjo, M. Chem. Pharm. Bull. 1986, 34, 3623.
- 3. (b) Maruyama, T.; Kimura, S.; Sato, Y.; Honjo, M. *J. Org. Chem.* **1983**, 48, 2719. (c) Kumadaki, I.; Nakazawa, M.; Kobayashi, Y. Maruyama, T.; Honjo, M. *Tetrahedron Lett.* **1983**, 24, 1055.
- (a) Voznyi, Ya. V.; Kochetkov, N. K. Carbohydr. Res. 1977, 54, 300. (b) Bochkov, A. F.; Chernetskii, V. N.; Kochetkov, N. K. Carbohydr. Res. 1975, 43, 35. (c) Bochkov, A. F.; Dashunin, V. M.; Shashkov, A. S.; Kochetkov, N. K. Izv. Akad. Nauk SSSR, Ser. Khim. 1975, 1061. (d) Bochkov, A. F.; Chernetskii, V. N.; Kochetkov, N. K. Izv. Akad. Nauk SSSR, Ser. Khim. 1975, 465. (e) Bochkov, A. F.; Obruchnikov, I. V.; Chernetskii, V. N.; Kochetkov, N. K. Carbohydr. Res. 1974, 36, 191.
- 5. Bertolini, M.; Glaudemans, C. P. J. Carbohydr. Res. 1971, 18, 131.
- 6. See reference 1k for the comment that "...the majority of cyclonucleosides suffer from electronic and/or conformational shortcomings that make them less than ideal probes."
- 7. (a) Groziak, M. P.; Koohang, A. *J. Org. Chem.* **1992**, *57*, 940. (b) Groziak, M. P.; Koohang, A.; Stevens, W. C.; Robinson, P. D. *J. Org. Chem.* **1993**, *58*, 4054. (c) Groziak,
- 8. M. P.; Lin, R.; Stevens, W. C.; Wotring, L. L.; Townsend, L. B.; Balzarini, J.; Witvrouw, M.; De Clercq, E. *Nucleosides Nucleotides* **1996**, *15*, 1041.
- 9. (a) Myers, A. G.; Gin, D. Y.; Rogers, D. H. J. Am. Chem. Soc. 1994, 116, 4697. (b) Ueda,
- 10. T. In *Chemistry of Nucleosides and Nucleotides*; Townsend, L. B., Ed.; Plenum: New York, 1988; Vol.1, pp 1–112; and related references cited therein. (c) Jones, G. H.; Taniguchi, M.; Tegg, D.; Moffatt, J. G. *J. Org. Chem.* **1979**, *44*, 1309. (d) Jones, G. H.; Moffatt, J. G. *J. Am. Chem. Soc.* **1968**, *90*, 5337.
- 11. Hayakawa, H.; Haraguchi, K.; Tanaka, H.; Miyasaka, T. Chem. Pharm. Bull. 1987, 35, 72.
- 12. (a) Simmons, H. E.; Wiley, D. W. J. Am. Chem. Soc. **1960**, 82, 2288. (b) Newkome, G. R.; Sauer, J. D.; McClure, G. L. Tetrahedron Lett. **1973**, 18, 1599.
- 13. A similar nucleoside 1,2-epoxide reportedly forms by the action of *m*-CPBA on a D-*erythro*-pent-1-enofuranosyluracil: Kittaka, A.; Tanaka, H.; Miyasaka, T.; Yamaguchi, K. *Nucleosides Nucleotides* 1992, *11*, 37.
- 14. Watson, S. C.; Eastham, J. F. J. Organomet. Chem. 1967, 9, 165.
- 15. Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155.
- 16. Groziak, M. P.; Lin, R.; Robinson, P. D. Acta Crystallogr. 1995, C51, 1204.

ISSN 1551-7004 Page 36 [©]ARKAT USA, Inc