

A new aromatic ester from the mangrove plant *Lumnitzera racemosa* Willd⁺

Ammanamanchi S.R.Anjaneyulu*, Yellajosyula L.N.Murthy,
Vadali Lakshmana Rao, Karanam Sreedhar

School of Chemistry, Andhra University, Visakhapatnam 530 003, India

E-mail: anjaneyuluasr@yahoo.com

Dedicated to Prof. Sukh Dev on the occasion of his eightieth birthday
(received 07 Oct 02; accepted 13 Jan 03; published on the web 16 Jan 03)

Abstract

Chemical examination of the Indian-mangrove plant *Lumnitzera racemosa* Willd has resulted in the isolation of a new aromatic ester besides the known triterpenoids, friedelin, betulin and betulinic acid. The structure of the new compound was established as 3-(4-hydroxyphenyl)-propyl-3¹-(3,4-dihydroxyphenyl)-propionate by a study of its spectral data.

Keywords: Mangrove plant, *Lumnitzera racemosa*, a new aromatic ester

Introduction

Lumnitzera racemosa Willd (Fam: Combretaceae) is a handsome shrub or a small tree found on the coast of India and on the Andaman and Nicobar Islands. The wood of *L. racemosa* is used as a fuel for its calorific value and the leaves of the plant are eaten in South Pacific Island during periods of scarcity. The reddish brown bark contains 15-19% tannin while the leaves and wood contain smaller quantities. A fluid obtained from incisions made in the stem was reported to be employed as an external application for the treatment of herpes and itches¹. Antihypertensive activity has been recently reported for the aqueous acetone extract of the plant². Chemical examination of this plant occurring in various parts of the world was reported to give a large number of compounds, long chain rubber like polyisoprenoid alcohols in leaves³, flavonoids and long chain fatty acids⁴ and low molecular weight carbohydrates⁵. Chemical examination of the Indian species was reported to give friedelin, β -amyrin, taraxerol, betulin, β -sitosterol and triacontanol⁶. The presence of trace elements was also reported⁷. In our continuing interest on the chemical constituents of Indian mangrove plants⁸⁻¹⁷ we have examined this species collected from the Bhiravapalem Island in the Godavary estuary and the results are reported herein.

Results and Discussion

The air dried and powdered stem of *L. racemosa* was exhaustively extracted with CH₂Cl₂: MeOH (1:1). Removal of the solvent from the combined CH₂Cl₂: MeOH extracts gave a residue which was extracted with EtOAc. Removal of the ethyl acetate under reduced pressure gave a residue which on repeated chromatographic separations over silica gel columns furnished a new aromatic ester **1**, in addition to the known triterpenoids, friedelin, betulin and betulinic acid.

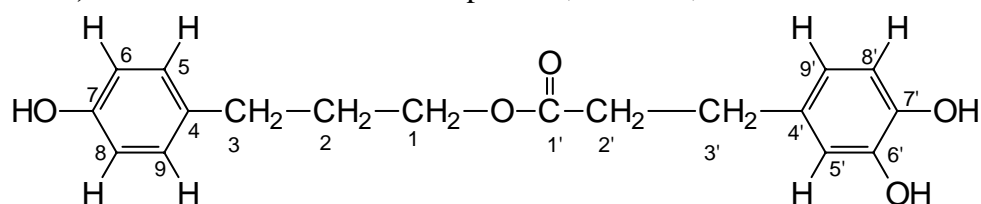


Figure 1

The new aromatic ester **1**, mp. 185⁰C was analysed for C₁₈H₂₀O₅ and its identity supported by the mass ion at m/z 299 [M+ H-H₂O]⁺ in its positive FAB mass spectrum. The IR band at 1708cm⁻¹ indicated the presence of an ester carbonyl and a strong peak at 3443 cm⁻¹ attributed to the phenolic hydroxyls. The molecular formula suggested nine double bond equivalents suggesting the presence of two aromatic rings besides the ester functionality. The ¹H NMR spectrum of **1** exhibited a splitting pattern consistent with a p-hydroxyphenyl substituted A₂B₂ system, δ 6.99 (2H, d, J=7.1Hz) and δ 7.29 (2H, d, J=7.1Hz) and a 3,4-dihydroxy-substituted phenyl ABC system with the signals for the three protons appearing at δ 6.82 (1H, d, J=7Hz), δ 6.59 (1H, d/d, J=7Hz, 1Hz) and δ 5.29 (1H, d, J=1Hz). The spectrum also indicated two benzylic methylenes and three aliphatic methylenes (**Table 1**). The chemical shifts and the multiplicities of the methylene protons clearly suggested 3-phenyl propoxy system and 3-phenylpropionic acid system linked as an ester. The ¹³C NMR spectrum of **1** showed only sixteen signals for the eighteen present suggesting two sets of identical carbons accounting for the p-hydroxyphenyl substituted ring system. The chemical shifts of the respective carbon atoms were assigned on the basis of comparative data and HMQC data. The ester carbonyl appeared at 173.9 (s) and the oxymethylene carbon at 63.8(t). The ¹H-¹H COSY data (**Table 1**) supported the expected connectivities. Similarly, the HMBC data, for example, the connectivities between the oxymethylene protons at δ 4.05 and C-2 and C-3 carbons and the connectivities between the methylene protons at δ 2.25 adjacent to the carbonyl, with carbonyl carbon C-1¹ and benzylic methylene carbon C-3¹ and other connectivities (**Table 1**) were in full agreement with the structure of the new aromatic ester as 3-(4-hydroxyphenyl)-propyl-3'-(3,4-dihydroxyphenyl)-propionate (**1**). Final proof for the structure of **1** was provided by its hydrolysis with alcoholic KOH and obtaining 3-(4-hydroxyphenyl)-1-propanol²¹ and 3,4-dihydroxydihydrocinnamic acid²² identified with the literature compounds (m.p & ¹H NMR).

Table 1. ^1H , ^{13}C NMR, ^1H - ^1H COSY and HMBC data of compound 1

Carbon no.	^1H (δ) (300MHz)	^{13}C (δ) (75MHz)	COSY	HMBC
C-1	4.05 (2H, t, 4.5)	63.8	H-2	C-3, C-2, C-1 ¹
C-2	2.09 (2H, t/t, 4.5,6.6)	28.6		C-3, C-1, C-4
C-3	2.80 (2H, t, 6.6)	33.9	H-2	C-5, C-9, C-4, C-1, C-2
C-4		137.8		
C-5	7.29 (1H, d, 7.5)	132.1	H-6	C-3, C-6, C-8, C-7, C-9
C-6	6.99 (1H, d, 7.5)	123.5		C-8, C-5, C-9, C-4, C-7
C-7		154.2		
C-8	6.99 (1H, d, 7.5)	123.5		
C-9	7.29 (1H, d, 7.5)	132.1		
C-1 ¹		173.9		
C-2 ¹	2.25 (2H, t, 5.5)	32.7		C-3 ¹ ,C-4 ¹ ,C-1 ¹
C-3 ¹	2.82 (2H, t, 5.5)	26.9	H-2 ¹	C-2 ¹ ,C-1 ¹ ,C-5 ¹ ,C-9 ¹ ,C-4 ¹
C-4 ¹		132.6		
C-5 ¹	5.29 (1H, d, 1.2)	112.6		C-3 ¹ ,C-9 ¹ ,C-4 ¹ ,C-7 ¹ ,C-6 ¹
C-6 ¹		149.1		
C-7 ¹	5.64 (OH)	142.5		C-8 ¹ ,C-7 ¹ ,C-6 ¹
C-8 ¹	6.82 (1H, d, 8.0)	115.0	H-9 ¹	C-9 ¹ ,C-4 ¹ ,C-7 ¹ ,C-6 ¹
C-9 ¹	6.59 (1H, dd, 8.0, 1.2)	121.4		C-3 ¹ ,C-5 ¹ ,C-7 ¹

Chemical shifts in δ from TMS taken in CDCl_3 with multiplicity and J values given in the brackets.

Experimental Section

General experimental procedures. Melting points were determined on a VEB-analytic Dreder HMK hot plate and are uncorrected. IR spectra were recorded on a Perkin-Elmer-841 IR spectrometer in CHCl_3 solution. ^1H NMR spectra were measured on a Bruker Advance DRX 300 and Jeol JNM EX-90 spectrometers. ^{13}C NMR spectra were measured on a Bruker Advance DRX 300 spectrometers at 75 MHz and Jeol JNM EX-90 spectrometer at 22.5 MHz using CDCl_3 as a solvent and tetramethylsilane as an internal reference. Elemental analyses were determined on a Carlo Ebra 1108 instrument. Mass spectra were obtained on a Jeol JMS-300 spectrometer.

Plant material. The stems of *Lumnitzera racemosa* were collected at the Bhiravapalem Island in the Godavari estuary ($16^{\circ} 58^1$ N Latitude and $82^{\circ} 15^1$ E Longitude) in March 1998. The plant material was identified by Prof. B.KondalaRao Dept of Marine Living Resources, Andhra University and the voucher specimens of the material have been kept in the museums of Organic chemistry, School of Chemistry, Andhra university and NIO Goa as AU1-166.

Extraction and isolation. The air-dried and powdered stem of *Lumnitzera racemosa* (4Kg) was exhaustively extracted with CH₂Cl₂ : MeOH (1:1) (8X8L). Removal of the solvent from the combined CH₂Cl₂ : MeOH extracts gave a residue (20 g) which was extracted with EtOAc (3 X 500 mL). Removal of the solvent under reduced pressure gave a residue (15 g) which was subjected to column chromatography over a column of silica gel (Acme brand, 100-200 mesh, 400 g) using solvents of increasing polarity from n-hexane through EtOAc. In all, 260 fractions (750 mL) were collected. The fractions showing similar spots were combined and the residues from therein were subjected to chromatography over silica gel or silver nitrate (20%) impregnated silica gel columns to yield four pure compounds as given below.

Fraction I. The residue (800 mg) from the column fractions 95-125 (n-hexane:EtOAc,8.75:1.25) was rechromatographed over a small column of silica gel using n-hexane and ethyl acetate mixtures as eluant to afford pure compound **1**, an aromatic compound (90 mg), crystallised from methanol as colourless needles. m.p 185⁰C, IR (Nujol) : 1708 cm⁻¹ (C=O), 3443 cm⁻¹ (OH), EIMS: m/z 299 [M + H-H₂O]⁺, Anal: Calcd for C₁₈H₂₀O₅, C, 68.35 ; H, 6.32 Found: C, 68.31 ; H, 6.28. For ¹H and ¹³CNMR, ¹H-¹H COSY, and HMBC data see **Table 1**

Fraction II. The residue (2 g) from the column fractions 35-60 (n-hexane : EtOAc, 9.5:5) was chromatographed through a small column of silica gel using n-hexane and ethyl acetate as eluants. The initial eluant fractions afforded compound **2**, friedelin (200 mg) (crystalised from methanol) mp 268⁰ [α]_D²⁵(-)-27.6⁰(c,1.2 in CHCl₃). IR (Nujol) : 1695 cm⁻¹(C=O), EIMS: m/z 426 [M]⁺, ¹H NMR (90 MHz CDCl₃) and ¹³C (22.5 MHz CDCl₃). Comparison of the physical and spectral data of **2** with the literature value^{6,18,20} of friedelin confirmed the characterization.

Fraction III. The residue (1.2 g) from the column fractions 60-95 (n-hexane:EtOAc,9.0:1.0) was rechromatographed through a small column of silica gel using n-hexane and ethyl acetate as eluants. The initial eluant fractions afforded compound **3**, betulin (120 mg) crystalised from MeOH- CHCl₃, m.p 252⁰C, [α]_D²⁵(+)-20.0⁰(c,1.2 pyridine) IR (Nujol) : 3560 cm⁻¹ (OH), EIMS: m/z 442 [M]⁺, ¹H NMR (90 MHz CDCl₃) and ¹³C (22.5 MHz CDCl₃). Comparison of the physical and spectral data of **3** with the literature values^{19,20} of betulin proved their identity.

Fraction IV. The residue (1.5 g) from the column fractions 125-180 (n- hexane:EtOAc,8:2) was green in colour and contained some impurity . The impurity was removed by repeated chromatography over small columns of silicagel using n-hexane:ethyl acetate mixtures as eluants . Repeated crystallisation of the residues from the column fractions from methanol-chloroform , afforded pure compound **4**, betulinic acid (900 mg) as colourless needles. mp 276⁰C [α]_D²⁵(+) 15.2⁰(c, 1.2, pyridine)IR (Nujol): 3485 (OH), 1720(C=O), 885 (CH₂) cm⁻¹,EIMS: m/z 456 [M]⁺,¹H NMR (90 MHz D₅Pyridine) and ¹³C (22.5 MHz D₅Pyridine). Comparison of the physical and spectral data of **4** with the literature values^{19,20} of betulinic acid confirmed the characterization.

Alkaline hydrolysis of compound 1. To compound **1** (25 mg) dissolved in methanol (10 ml) was added methanolic KOH (10%, 5 ml) and the mixture refluxed on a steam bath for 1 hour.

The mixture was diluted with water (20 ml) and then extracted into ether. The ether solution after evaporation gave 3-(4-hydroxyphenyl)-1-propanol m.p 52⁰C identical with the literature compound²¹ (m.p, ¹HNMR taken on 90MHz instrument in CDCl₃ with TMS as internal standard). The alkaline aqueous solution from the reaction was acidified with dil H₂SO₄ and the acid liberated was extracted into ether. The ether solution after evaporation left a residue which on crystallization from chloroform-methanol gave 3,4-dihydroxydihydrocinnamic acid m.p 136-38⁰ identical with the literature compound²² (m.p, ¹HNMR taken on 90MHz instrument in CDCl₃ with TMS as internal standard).

Acknowledgements

The authors are grateful to the Department of Ocean Development, New Delhi for funding and The Head, RSIC, Lucknow and the Co-ordinator, UGC-SAP Programme, School of Chemistry, Andhra University for providing the spectral data.

References and Notes

⁺ Part XI of the series "Chemical constituents of Indian Mangrove plants" for part X see ref.17.

1. The Wealth of India, Raw materials, Published by Publication and Information Directorate, CSIR , New Delhi, **1962**, p182.
2. Lin,Ta-chen.; Hsu,Feng-lin.; Cheng,Juei-Tang. *J.Nat.Prod.* **1993**, *56*, 629.
3. Skoczylas,E.; Swiezewska,E.; Chojnacki,T.;Tanaka,Y. *Plant Physiol.Bioche.***1994**, *32*, 825.
4. Huangliang,Y.; Su,M.; Chen,P. *Tianran Chanwu Yanjiu Yu Kaifa.* **1994**, *6*, 6.
5. Popp, M. *Z.Pflanzenphysiol.* **1984**, *113*, 411.
6. Majumdar, S.G.;Patra, G. *J.Indian Chem Soc.* **1980**, *57*, 568.
7. Bhosale, L. J. *Indian J.Mar.Sci.* **1979**, *8*, 58.
8. Anjaneyulu, A S R.; Lakshmana Rao, V. *Phytochemistry* **2000**, *55*, 891.
9. Anjaneyulu, A S R.; Anjaneyulu, V.; Lakshmana Rao, V. *Indian J. Chem.* **2000**, *39B*, 803.
10. Anjaneyulu, A S R.; Lakshmana Rao, V. *Nat. Prod. Lett.* **2001**, *15*, 13.
11. Anjaneyulu, A S R.; Lakshmana Rao, V.; Sreedhar, K. *J. Nat. Prod.* **2002**, *65*, 382.
12. Anjaneyulu, A S R.; Anjaneyulu, V.; Lakshmana Rao, V. *J. Asian Nat. Prod.* **2002**, *4*, 53.
13. Anjaneyulu, A S R.; Lakshmana Rao, V. *Phytochemistry*. (In press).
14. Anjaneyulu, A S R.; Lakshmana Rao, V.; Lobkovsky, E.;Clardy, J. *J. Nat. Prod.* **2002**, *65*, 592.
15. Anjaneyulu, A S R.; Lakshmana Rao, V.; Sreedhar, K. *Nat. Prod. Lett.* (In press).
16. Anjaneyulu, A S R.; Lakshmana Rao, V. *Phytochemistry* (In press).
17. Anjaneyulu, A. S. R .;Lakshmana Rao, V. *Phytochemistry* (In press).

18. Amarendra, P.; Swapan, K. C.; Heinz, R. *J. Indian Chem. Soc.* **1990**, *67*, 394.
19. Ghosh, A. G.; Mishra, S.; Dutta, A. K.; Chowdary, A. *Phytochemistry*. **1985**, *24*, 1725.
20. Mahato, S. B.; Kundu, A. P. *Phytochemistry*. **1994**, *37*, 1517.
21. The Aldrich Library of ^{13}C and ^1H FT-NMR Spectra, Edition-1, Vol. 2, p 409, CAS [10210-17-0].
22. The Aldrich Library of ^{13}C and ^1H FT-NMR Spectra, Edition-1, Vol. 2, p 1038, CAS [1078-61-1].